

Oct 15, 2024

Overview of MultiPathogen Detection Workflow

DOI

<https://dx.doi.org/10.17504/protocols.io.kqdg32jq7v25/v1>

Shannon Fitz¹, Alex Shaw¹, michael Owusu², Dilip Abraham³

¹Imperial College London; ²Kwame Nkrumah University of Science and Technology; ³CMC Vellore

GEMS - Genomic Enviro...



Shannon Fitz

Imperial College London

Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

OPEN  ACCESS



DOI: <https://dx.doi.org/10.17504/protocols.io.kqdg32jq7v25/v1>

Protocol Citation: Shannon Fitz, Alex Shaw, michael Owusu, Dilip Abraham 2024. Overview of MultiPathogen Detection Workflow. protocols.io <https://dx.doi.org/10.17504/protocols.io.kqdg32jq7v25/v1>

License: This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: October 03, 2024

Last Modified: October 15, 2024

Protocol Integer ID: 109068

Keywords: multipathogen detection, hepatitis E, vibrio cholera, salmonella typhi, enterovirus, gems multipathogen detection workflow, overview of multipathogen detection workflow, multipathogen detection workflow, multipathogen detection workflow targets detection, comprehensive overview of the sequential step, sequencing, wastewater sample, detection, enterovirus, sequential step, workflow, vibrio cholera

Funders Acknowledgements:

The Gates Foundation

Grant ID: INV-049092 PA4273

Abstract

This protocol offers a comprehensive overview of the sequential steps involved in the GEMS multipathogen detection workflow, spanning from sample concentration to sequencing. This multipathogen detection workflow targets detection of Hepatitis E, enteroviruses, Vibrio cholera, and Salmonella typhi from wastewater samples.



Concentration

- 1 Invert collected samples several times prior to decanting to ensure even mixing.

Separate sample into appropriate volume per concentration method (PEG = 40 ml, Ceres = 35 ml), and follow the selected protocol:

<https://www.protocols.io/view/ceres-nanoparticles-concentration-and-extraction-u-n2bvjnpwxgk5/v1>

<https://www.protocols.io/view/polyethylene-glycol-precipitation-for-wastewater-s-j8nlk8pzd15r/v1>

Extraction

- 2 Following concentration with the chosen method, carry out extraction using MagMAX Wastewater Ultra Nucleic Acid Isolation Kit (protocol included in the above listed concentration protocols)

PCR purification

- 3 After extraction, carry out the Zymo OneStep PCR Inhibitor Removal protocol.

<https://www.protocols.io/view/zyzo-onestep-pcr-inhibitor-removal-kit-5qpvmrbl4o/v1>

PCR

- 4 Three distinct PCRs will be conducted, so samples will be tested separately at this next stage. Depending on timeline for testing, it may be beneficial to separate samples into different aliquots at the step prior to avoid multiple freeze-thaws.

Run EV/HEV, typhi, and cholera assays.

<https://www.protocols.io/view/hepatitis-e-and-enterovirus-multiplex-nested-rt-pc-rm7vzjd74lx1/v1>

<https://www.protocols.io/view/multiplex-nested-pcr-for-vibrio-cholera-36wgqgnzbkgk5/v1>

<https://www.protocols.io/view/nested-pcr-amplification-of-salmonella-typhi-from-36wgqgnw3ogk5/v1>

Sequencing



- 5 Lastly, amplicon sequencing for all samples will be carried out using the Oxford Nanopore Rapid Barcoding Kit.

<https://www.protocols.io/view/rapid-barcoding-protocol-v14-n92ld8ew8v5b/v1>