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Oligo Pool Resuspension and PCR

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<https://www.twistbioscience.com/resources/protocol/twist-dna-resuspension-guidelines>

Twist Bioscience. (n.d.). *Twist Oligo Pool Amplification Guidelines* | *Twist Bioscience*. Retrieved October 9, 2023, from

<https://www.twistbioscience.com/resources/protocol/twist-oligo-pool-amplification-guidelines>



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We use this protocol and it's working

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Abstract

2023 NUS-Singapore iGEM team followed this protocol to resuspend the dried oligo pool and perform PCR to amplify the DNA fragments in the oligo pool.

Materials

KOD One™ PCR Master Mix (Blue)

Protocol materials

 Buffer EB [Qiagen Catalog #19086](#)

Safety warnings

 Proper lab PPE must be worn at all times.



Oligo Pool Resuspension

- 1 Centrifuge the oligo pool for a few seconds before opening the tube.
- 2 Calculate the volume of  Buffer EB **Qiagen Catalog #19086** required for the oligo pool (the final concentration of the sample must be at least 20ng/ μ L).
- 3 Add the respective volume of  Buffer EB **Qiagen Catalog #19086** into the tube with the oligos and resuspend the dried oligos by pipetting the solution.

Oligo Pool PCR

- 4 Prepare a PCR tube, and add the following reagents:

Item	Volume
KOD One™ PCR Master Mix (Blue)	25 μ L
Each Primer (Two Primers In Total)	1.5 μ L
Oligo Pool Sample	1 μ L
DI Water	21 μ L

- 5 Run a PCR with the following setting:

Temperature	Duration
98°C	10s
60°C	5s
68°C	10s
Go to step 1, repeat the cycle 10 times	
68°C	5 minutes
12°C	Infinite Loop



- 6 Follow the PCR clean-up protocol to isolate the oligos from the PCR product and keep it at  Room temperature .

Protocol

NAME

PCR Clean-up

CREATED BY

NUS iGEM

Preview

Protocol references

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