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NucleoSpin® Gel and PCR Clean-up

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We use this protocol and it's working

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Abstract

- Buffer NTI
- NucleoSpin® Gel and PCR Clean-up Column
- NucleoSpin® Gel and PCR Clean-up Collection Tube (2ml)
- Buffer NT3
- Buffer NE / nuclease-free water
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Solubilize gel slice

For each 100 mg of agarose gel < 2 % add 200 μ L Buffer NTI. For gels containing > 2 % agarose, double the volume of Buffer NTI. Incubate sample for 5–10 min at 50 °C. Vortex the sample briefly every 2–3 min until the gel slice is completely dissolved!

Bind DNA

Place a NucleoSpin® Gel and PCR Clean-up Column into a Collection Tube (2 mL) and load up to 700 μ L sample. Centrifuge for 30 s at 11,000 x g. Discard flow-through and place the column back into the collection tube. Load remaining sample if necessary and repeat the centrifugation step.

Wash silica membrane

Add 700 μ L Buffer NT3 to the NucleoSpin® Gel and PCR Clean-up Column. Centrifuge for 30 s at 11,000 x g. Discard flow-through and place the column back into the collection tube.

Dry silica membrane

Centrifuge for 1 min at 11,000 x g to remove Buffer NT3 completely. Make sure the spin column does not come in contact with the flow-through while removing it from the centrifuge and the collection tube. Note: Residual ethanol from Buffer NT3 might inhibit enzymatic reactions. Total removal of ethanol can be achieved by incubating the columns for 2–5 min at 70 °C prior to elution.

Elute DNA

Place the NucleoSpin® Gel and PCR Clean-up Column into a new 1.5 mL microcentrifuge tube (not provided). Add 15–30 μ L Buffer NE and incubate at room temperature (18–25 °C) for 1 min. Centrifuge for 1 min at 11,000 x g.

