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Version 4

noesypr1d_metab.nan V.4

Forked from a private protocol

 In 2 collections



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We use this protocol and it's working

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Abstract

This is a protocol for running the Bruker pulse program "noesypr1d".

Guidelines

This protocol intends to provide concise instructions to carry out the experiment. For more comprehensive information, see Bruker's documentation including "Basic NMR Experiments" by clicking ? → **Manuals (docs)** on the menu bar on TopSpin.

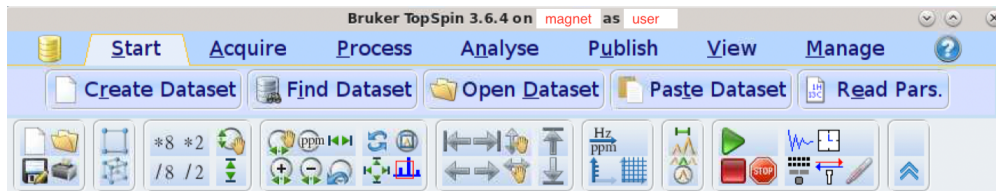
Before start

This protocol assumes your sample is loaded, locked, tuned, and shimmed in the magnet.

Create a new dataset

1

- 1.1 On the menu bar on TopSpin, click on
Start → Create Dataset



(This protocol uses TopSpin 3.6.4, and the interface may look different on other TopSpin versions.)

Note

You can also use the **new** command in the command line to do this step.

- 1.2 Enter
- **NAME**: Name of a set of datasets (e.g., human_serum_study1). Use a single string.
 - **EXPNO**: Dataset number. Use a positive integer.

Select

- **Directory**: Your directory.

Create New Dataset - new

Prepare for a new experiment by creating a new data set and initializing its NMR parameters according to the selected experiment type. For multi-receiver experiments several datasets are created. Please define the number of receivers in the Advanced.

Dataset

NAME: human_serum_study1
EXPNO: 1
Directory: /opt/nmrdata/User
☐ Open in new window

Parameters

☐ Use current parameters
☒ Read parameterset: NOESYPR1D_br600_serum.par [Select]
☒ Set solvent: D2O_salt
Additional action:
☒ No additional action
☐ Execute getprosol
☐ Keep parameters: P 1, O1, PLW 1 [Change]

Advanced

Number of datasets (receivers): 1
Title:

[OK] [Cancel] [More Info...] [Help]

Note

Your new dataset will be stored in **Directory/NAME/EXPNO**

1.3 Select Read parameterset

Click the button

Select

1.4 A new window opens. On the right top bar, select Source = /opt/NAN_METAB/par

In the list, select the one you want to use:

NOESYPR1D_br600_serum.par: Parameter set optimized for serum and plasma samples.

NOESYPR1D_br600_urine.par: Parameter set optimized for urine samples.

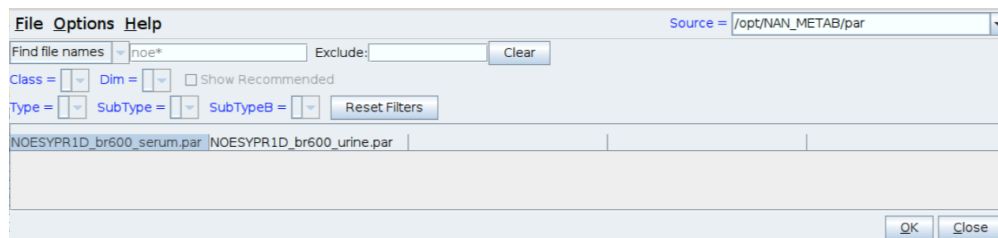
Note

Parameter set names in the list vary between spectrometers (e.g., NOESYPR1D_br800_serum.par).



Click

OK



1.5 Click

OK

2 Go to the "USE DEFAULT" tab below to proceed with the default optimized parameters.

STEP CASE

Use default parameters: 7 steps

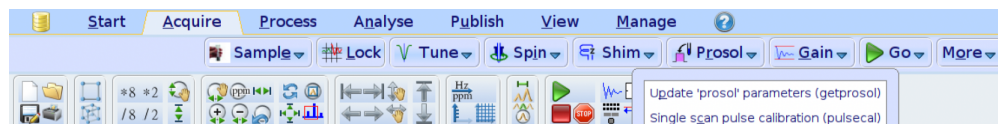
This step case uses the default optimized parameters to acquire a spectrum.

3

3.1 Click on

Acquire → Prosol → Single scan pulse calibration (pulsecal)

to calibrate the 90° proton pulse (i.e., P1).



A new window will appear to show the calibrated P1.

Note

You can also use the **pulseval** command in the command line.

- 3.2 Load the calibrated P1 using the following command in the command line.



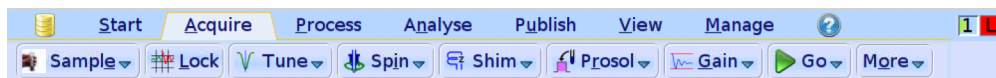
getprosol 1H [calibrated P1 value] [power level for P1]

(e.g., getprosol 1H 10.01 -7.45)

Note

[power level for P1] varies between spectrometers. Never use a wrong **[power level for P1]**

- 3.3 Click on
Acquire → Gain
in the menu bar to automatically set the receiver gain.



Note

You can also use the **rga** command in the command line.

- 3.4 Click
Go
in the menu bar to acquire a spectrum.

Note

You can also use the **zg** command in the command line.



- 3.5 Click on **Process** → **Proc. Spectrum** in the menu bar to execute an automated processing macro.



- 3.6 If you want to modify parameters to improve your spectrum,  go to step #2 and move to a step case "MODIFY PAR".