May 13, 2023 Version 1

NCEM Drop - Tissue Dounce Homogenisation (TM-014) V.1

DOI
https://dx.doi.org/10.17504/protocols.io.bp2l69j4dlqe/v1

sandra.crameri[1]
[1]CSIRO ACDP

sandra.crameri

DOI: https://dx.doi.org/10.17504/protocols.io.bp2l69j4dlqe/v1

Protocol Citation: sandra.crameri 2023. NCEM Drop - Tissue Dounce Homogenisation (TM-014). **protocols.io** https://dx.doi.org/10.17504/protocols.io.bp2l69j4dlqe/v1

Protocol status: Working
We use this protocol and it's working

Created: May 13, 2023

Last Modified: May 13, 2023

Protocol Integer ID: 81824

Keywords: cell pellet dounce homogenisation, tissue dounce homogenisation, ncem drop, ncem, tissue

Abstract

cell pellet dounce homogenisation

Protocol materials

nano-W **Catalog** ##2018-5ML

HEADER

1 SAN:

SPEC No:

OPERATOR:

Dounce Homogenisation

2 1. Remove cells from the flasks by scraping with a disposable plastic scraper and poor into a 10 mL centrifuge tube.

3 Pellet the cells in a bench top centrifuge at 2000 rpm 00:02:00 , remove supernatant. 2m

4 Resuspend the cells in equal volume TC water and transfer to glass homogeniser tube.

5 Insert teflon plunger into cordless drill, set drill on max speed.

6 Homogenise(20 strokes) inside Class II BSC cabinet.

7 Clarify resultant solution at 13000 rpm in Eppendorf centrifuge for 00:01:00 1m

8 Stand for 00:05:00 to permit viruses to diffuse back into solution, from the debris, and provide an interface for the sampling of membrane associated viruses. Use supernatant for sample below. 5m

Conventional 11m

9 Adsorb 10 µL sample to grid 00:10:00 , inspect to ensure sample does not dry out. 10m

10 Drain excess sample from grid using filter paper, leave wet.

11 Stain nano-W **Catalog** ##2018-5ML 00:01:00 1m

12 Drain & dry using filter paper