

Nov 28, 2022

# MycoFluor Mycoplasma Detection

DOI

dx.doi.org/10.17504/protocols.io.bp2l692nklqe/v1

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Protocol Citation: Ali Albalakhi, Ning Xia 2022. MycoFluor Mycoplasma Detection . protocols.io

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Protocol status: Working

We use this protocol and it's working.

Created: November 23, 2022



Last Modified: December 12, 2024

Protocol Integer ID: 73200

Keywords: ASAPCRN, mycofluor mycoplasma detection, fluorescent mycofluor, mycoplasma infection in cell culture, mycoplasma infection, simple fluorescence microscopic assay, mycoplasma, fluorescence microscope, simple fluorescence, microscopic assay, cell culture, cell, infection, detection

#### Abstract

MycoFluor™ Mycoplasma Detection produces a fast and simple fluorescence microscopic assay that identifies mycoplasma infection in cell cultures. In order to detect mycoplasma, the fluorescent MycoFluor™ reagent is added to the culture medium, with or without cells, and the sample becomes stained and examined under a fluorescence microscope.

#### **Materials**

### **Reagents Needed:**

- 1. MycoFluor Mycoplasma Detection Kit (M-7006)
- 2. Microcentrifuge tubes
- 3. Microscope slides
- 4. Clear coverslips

## **Troubleshooting**



## **Protocol Testing of Culture Media**

- 1 Take about 4mL of cell medium directly form the culture dish in which the cells have been growing centrifuge the sample at 1300 x g for 10min to pellet any cells and debris
- 2 Carefully transfer 1mL of supernatant into labeled microcentrifuge tubes.
- 3 Centrifuge the microcentrifuge tubes at 12,500 x g for 15 minutes
- 4 Carefully remove and discard 0.5mL of supernatant, leaving behind 0.5mL of medium in the tube. Resuspend any pellet that may have formed using this 0.5mL of medium.
- 5 Add 26µL of 20X concentrated MycoFluor reagent to 0.5mL of the medium.
- 6 Pipet 10µL of the stained medium onto a clean microscope slide and cover with a clean coverslip.
- 7 Seal the slide using quick dry clear nail polish topcoat by covering all sides of the coverslip.
- 8 Image the slide using fluorescent microscope.

# Control Slides with Mycoplasma MORFS

- 9 Generating positive controls for the testing of culture media
- 9.1 a. Pipet 5µL of the mycoplasma MORFS stock suspension on a clean, labeled microscope slide
- 9.2 b. Add 5µL of stained medium to the slide
- 9.3 c. Cover with clear coverslip and seal using quick dry topcoat nail polish.



10 Image the positive controls and compare to the samples that have not been spiked with **MORFS** 

# Microscopy

- 11 Prepare the microscope with a near ultraviolet fluorescence filter (Excitation at 365nm and either bandpass 450nm ± 30nm or longpass >400nm emission filter)
- 12 For optimum result, a 100X oil immersion objective is suggested but a 60X oil immersion objective will also work
- 13 First look at the control slide spiked with MORFS to get an idea what it should look like. Then examine the test slide for extranuclear blue fluorescence.