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Mouse perfusion

 Forked from [Mouse perfusion](#)

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Protocol status: Working

We use this protocol and it's working

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Abstract

General protocol for mouse perfusion fixation. In this version, other tissues outside of the brain are fixed and ready for processing.

Materials

DPBS

pentobarbital sodium and phenytoin sodium

4% paraformaldehyde (diluted from

 Pierce & Warriner; 16% Formaldehyde (w/v), Methanol-free **Thermo Fisher Catalog #28908**) in 1X DPBS

Troubleshooting



Mouse Perfusion

- 1 Anesthetize mouse using pentobarbital sodium and phenytoin sodium. A 30g mouse would need roughly 100 ul of this pentobarbital sodium and phenytoin sodium solution.
- 2 Ensure deep anesthesia with paw pinch. Mouse should not respond.
- 3 Using four 26g needles, secure mouse paws to styrofoam.
- 4 Make an incision along the abdomen, extending up to the rib cage.
- 5 Expose chest cavity and secure with hemostats clamped to the xiphoid process.
- 6 Insert 22g needle into the left ventricle and hold via hemostat.
- 7 Make small incision in the right ventricle.
- 8 Immediately start peristaltic pump connected to sterile filtered DPBS, flowing at a rate of 10 mL/min
- 9 Once the perfusate runs clear, stop pump, switch to 4% PFA solution, and resume flow.
- 10 Perfuse mouse with approximately 100mL of 4% PFA. 
- 11 Remove head with scissors, and carefully dissect out the brain. Remove the skin and limbs and place the skinless, headless carcass into 50mL 4% PFA solution, overnight, at 4C. 
- 12 Skull should be snipped at the base, and peeled away carefully. Care should be taken not to damage underlying brain tissues or olfactory bulb.
- 13 Place dissected brain in 20mL 4% PFA solution, overnight, at 4C.

1d

14 Wash the tissue with DPBS for a minimum of 3 times, 10 minutes per each wash.

10m

15 Place the gut into DPBS + azide until ready for sectioning

16 Place the brain in 15% sucrose DPBS solution overnight.

1d



17 Once brain sinks, Place in 30% sucrose DPBS solution at 4C until sectioning.

