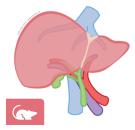
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# O Mouse 2- Step Collagenase Liver Perfusion protocol

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### Protocol status: Working We use this protocol and it's working

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# Abstract

Analyzing the mouse liver requires optimal cell recovery from the tissue in terms of quality (viability) and quantity. Harsh dissociation methods can damage fragile cell populations, resulting in low viability and difficulty in cell culture. We have developed a gentle surgical and enzymatic technique based on the two-step collagenase perfusion protocol for single-cell analysis and primary cell culture.

## **Protocol materials**

Dulbecco's Modified Eagle's Medium - high glucose Merck MilliporeSigma (Sigma-

- Aldrich) Catalog #D5796
- X Heparin LEO® LEO Pharma Catalog #006174-09

X Collagenase from Clostridium histolyticum Merck MilliporeSigma (Sigma-Aldrich) Catalog #C5138

### Preparation

- **Anesthetize** mouse with inhalational anesthesia (4-5% isofluorane) in the induction chamber
- 2 **Heparinize** the animal intraperitoneally by injecting [M] 5 units/g (body weight) of heparin:

Note

Preventing blood clotting improves cell recovery.

X Heparin LEO® LEO Pharma Catalog #006174-09

- **3 Once the animal is anesthetized, transfer** it onto a homeothermic warming pad. The animal will receive 1-3% isofluorane via face-mask inhalation fo the remaining steps.
- 4 **Prepare the abdomen for surgery:** shaved with a trimmer, prepared with iodine-based solution and alcohol-based solution, and draped in a sterile fashion.

### Laparotomy

5 **Midline incision:** cut open the abdomen from the pubic symphysis to the xiphoid process. Flip over the intestines to locate the liver and its portal vein.

Please refer to Figure 14.4.1 of the following publication:

### CITATION

Froh M, Konno A, Thurman RG (2003). Isolation of liver Kupffer cells.. Current protocols in toxicology.

LINK

https://doi.org/10.1002/0471140856.tx1404s14

### Cannulation

6 **Cannulate** the portal vein with the 22-gauge catheter. Secure it cannulation by suture or a micro serrefine.

Equipment		
Insyte™ Autoguard™ BC	NAME	
Intravenous Catheter	TYPE	
BD	BRAND	
382523	SKU	
https://www.bd.com/en-ca/offerings/capabilities/infusion-therapy/iv- catheters/bd-insyte-autoguard-bc-shielded-iv-catheter-with-blood-control- technology		
Non-winged, blue, 22 gauge, 1 inch, 0.9 × 25 mm	SPECIFICATIONS	



# Equipment NAME Schwartz Micro Serrefine NAME Micro Serrefine TYPE Fine Science Tools BRAND 18052-03 SKU https://www.finescience.com/en-US/Products/Vascular-Instruments/Clamps LINK Occluders/Schwartz-Micro-Serrefines/18052-03 SPECIFICATIONS Sharp Bend SPECIFICATIONS

7 **Collect blood:** collect blood from the inferior vena cava (IVC) with a 20-gauge needle.

8 **Cut the IVC and proceed to the next step immediately.** Cutting the IVC allows blood and perfusate to drain.

### Perfusion

9 Perfuse liver with calcium-free buffer without collagenase: I to mL of Hanks' 5m
 Balanced Salt Solution (HBSS) without calcium or magnesium +
 IMJ 0.5 millimolar (mM) EGTA at a flow rate of 2 mL/min until fluid drained from the IVC is no longer red (containing blood). This is done with a pump with no recirculation.
 Note

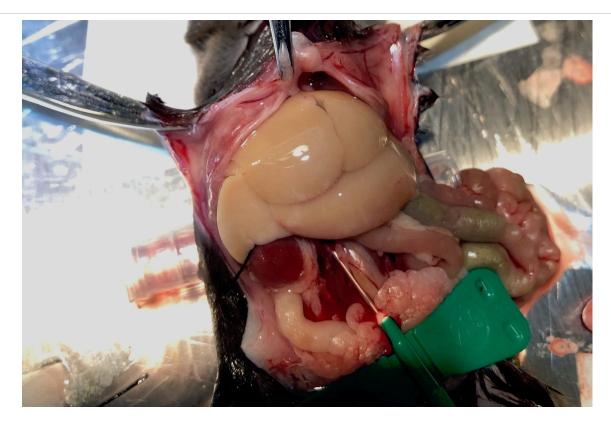
This flow rate **(2 mL/min)** is slower than other liver perfusion protocols. We found that a slower perfusion rate results in hepatocytes with higher quality and quantity and therefore suitable for analyses such as single-cell RNA sequencing.

Equipment	
Model EP-1 Econo Pump	NAME
Peristaltic Pump	TYPE
Bio-Rad	BRAND
7318140	SKU
http://www.bio-rad.com/en-ca/product/model-ep-1-econo-pump? ID=6ee41e9b-5ac5-449c-ae7d-578dca8ed6cf	LINK

10 Perfuse liver with calcium-containing buffer with collagenase: 4 10 mL of Hanks' Balanced Salt Solution (HBSS) **with** calcium and magnesium + [M] 0.1 mg/mL type IV collagenase at the same flow rate (2 mL/min). This is done with the same pump with no recirculation. Collagenase from Clostridium histolyticum Merck MilliporeSigma (Sigma-

Aldrich) Catalog #C5138

5m



A well-perfused liver shows drastic discoloration due to the lack of blood. A welldissociated liver shows white patches.

## Harvest

11 **Excise** the dissociated liver out of the animal and place it in DMEM containing 10% FBS, which inactivates the collagenase enzyme.

Dulbecco's Modified Eagle's Medium - high glucose Merck MilliporeSigma (Sigma-Aldrich) Catalog #D5796

# **Cell Isolation**

12 **Gently cut** the excised liver with a scalpel to release the dissociated cells.

Note

Good dissociation can be visualized by abundance of cells released (in pink) with little cutting. Shake the tissue gently to release cells. Avoid tearing the tissue with the scalpel (horizontal movement); only cut the tissue vertically.

Λ

### 13 **Filter** dissociated cells with a 70 µm filter:

Equipment			
70 μm Cell Strainer	NAME		
Cell Strainer	TYPE		
Falcon	BRAND		
352350	SKU		
https://ecatalog.corning.com/life-sciences/b2c/US/en/Cell-Culture/Cell- Culture-Accessories/Cell-Strainers/Falcon®-Cell-Strainers/p/352350? clear=true			
White, Sterile, Individually Packaged	e, Sterile, Individually Packaged SPECIFICATIONS		

# **Cell Enrichment**

14	Enrich dissociated cells for hepatocytes by centrifugation 🚯 50 x g		5m	
	Room temperature	for 5 minutes. The supernatant can be further pu	irified in the	
	next step. Resuspend th	e pellet (hepatocyte-rich) with DMEM.		
15	Enrich supernatant for	non-parenchymal cells (NPC) by centrifugation	<b>3</b> 400 x g	5m
	Room temperature	for 5 minutes. Discard the supernatant. Resusper	nd the pellet	69
	(NPC-rich) with DMFM			

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## Citations

Step 5

Froh M, Konno A, Thurman RG. Isolation of liver Kupffer cells. https://doi.org/10.1002/0471140856.tx1404s14