MojoSort™ Streptavidin Nanobeads Column Protocol - Negative Selection V.2

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Abstract

BioLegend MojoSort™ nanobeads work in commonly used separation columns, based on our internal research as well as validation by external testing by academic labs. This simple protocol consists of following the MojoSort™ protocol to label the cells with **pre-diluted** MojoSort™ reagents and using the columns as indicated by the manufacturer.

Note: Due to the properties of our beads, it may be possible to use far fewer beads and less antibody cocktail that with other commercial suppliers. We recommend a titration to find the best dilution factor. However, as a general rule, dilutions ranging from 1:2 to 1:10 for the antibody cocktail can be used. Dilutions ranging from 1:5 to 1:20 for the Streptavidin Nanobeads can be used. Please contact BioLegend Technical Service (tech@biolegend.com) if further assistance is needed.

Guidelines

MojoSort™ magnetic particles can be used with other commercially available magnetic separators, both free standing magnets and column-based systems. Because MojoSort™ protocols are optimized for the MojoSort™ separator, the protocols may need to be adjusted for other systems. Please contact BioLegend Technical Service (tech@biolegend.com) for more information and guidance. We do not recommend using MojoSort™ particles for BD's IMag™ or Life Technologies' DynaMag™.

Protocol Selection: If your target cells are the labeled cells (the positive fraction), use the Streptavidin Nanobeads Column Protocol – Positive Selection. If your target cells are the unlabeled cells (negative fraction), use the Streptavidin Nanobeads Column Protocol - Negative Selection.

Materials

MATERIALS

MojoSort™ Buffer **BioLegend Catalog #**480017

MojoSort™ Streptavidin Nanobeads

Biotin-Conjugated Primary Antibody

Additional reagents:
-commercially available cell separation columns
-5 mL polypropylene tubes
-70 µm cell strainer

Troubleshooting

1 Prepare cells from your tissue of interest or blood without lysing erythrocytes.

2 In the final wash of your sample preparation, resuspend the cells in MojoSort™ Buffer by adding up to 4 mL in a 5 mL (12 × 75 mm) polypropylene tube.
Note: Keep MojoSort™ Buffer on ice throughout the procedure.

3 Filter the cells with a 70 µm cell strainer, centrifuge at 300xg for 5 minutes, and resuspend in an appropriate volume of MojoSort™ Buffer. Count and adjust the cell concentration to 1 × 10^8 cells/mL by adding MojoSort™ Buffer. 5m

4 Aliquot 100 µL of cell suspension (10^7 cells) into a new tube. Check the recommended usage for flow cytometric staining of the Biotin-conjugated antibody indicated in the antibody technical datasheet. Calculate the volume to stain 10^7 cells (or desired amount of cells). **Add the appropriate volume of pre-diluted Biotin-conjugated antibody** to the cell suspension, mix well and **incubate on ice for 15 minutes**. 15m

Note: For the Biotin-conjugated antibodies, we recommend to do a titration to determine the optimal concentration.

5 Wash the cells by adding MojoSort™ Buffer up to 4 mL. Centrifuge the cells at 300xg for 5 minutes. 5m

6 Discard the supernatant and resuspend cells in 100 µL of MojoSort™ Buffer.

7 Resuspend the beads by vortexing, maximum speed, 5 touches. **Add the appropriate volume of pre-diluted Streptavidin Nanobeads**. Mix well and **incubate on ice for 15 minutes**. Scale up the volume accordingly if separating more cells. For example, if the volume of pre-diluted Nanobeads for 1×10^7 cells is 10 µL, add 100 µL for 1 × 10^8 cells. When working with less than 10^7 cells, use indicated volumes for 10^7 cells. 15m
Note: The amount of Nanobeads to use always depends on the frequency of the target, among a few other factors. We recommend to do a titration to determine the optimal concentration.

8 Wash the cells by adding MojoSort™ Buffer up to 4 mL. Centrifuge the cells at 300xg for 5 minutes. 5m

9 Discard the supernatant.

10 Resuspend the cells in appropriate amount of MojoSort™ Buffer and proceed to separation. At least 500 µL is needed for column separation.

Note: There are several types of commercially available columns, depending on your application. Choose the one that fits best your experiment:

	Max. number of labeled cells	Max. number of total cells	Cell suspension volume	Column rinse volume	Cell wash volume	Elution volume
Small Capacity	1×10^7	2×10^8	500µL for up to 10^8 cells	1ml	1 ml	1 ml
Medium Capacity	1×10^8	2×10^9	500µL for up to 10^9 cells	3ml	3 ml	5 ml
Large Capacity	1×10^9	2×10^{10}	500µL for up to 10^{10} cells	20-50ml	30 ml	20 ml

Example of magnetic separation with medium capacity columns:

11 Place the column in a magnetic separator that fits the column.

12 Rinse the column with 3 mL of cell separation buffer.

13 Add the labeled cell suspension in at least 500 µL of buffer to the column through a 30 µm filter and collect the fraction containing the unlabeled cells. These are the cells of interest; do not discard.

14 Wash the cells in the column **2 times** with 3 mL of buffer and collect the fraction containing the untouched cells. Combine with the collected fraction from step 3.

15 If desired, the labeled cells can be collected by taking away the column from the magnet and place it on a tube. Then add 5 mL of buffer and flush out the magnetically labeled fraction with a plunger or supplied device. The labeled cells may be useful as staining controls, to monitor purity/yield, or other purposes.