

MojoSort™ Nanobeads Column Protocol - 1

DOI

<https://dx.doi.org/10.17504/protocols.io.69dhh26>

Sam Li¹

¹BioLegend

BioLegend

Tech. support email: tech@biolegend.com



Sam Li

BioLegend

Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account



DOI: <https://dx.doi.org/10.17504/protocols.io.69dhh26>

External link: <https://www.biolegend.com/protocols/mojosort-nanobeads-column-protocol-1/4593/>

Protocol Citation: Sam Li . MojoSort™ Nanobeads Column Protocol - 1. [protocols.io](https://dx.doi.org/10.17504/protocols.io.69dhh26)

<https://dx.doi.org/10.17504/protocols.io.69dhh26>

License: This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Created: September 10, 2019

Last Modified: September 10, 2019

Protocol Integer ID: 27653



Keywords: cell isolation, separation, mojosort, nanobeads, fewer beads than with other commercial supplier, best dilution factor, dilution, used separation column, fewer bead, nanobead, separation column, bead, biolegend mojosort, reagent, biolegend technical service

Abstract

BioLegend MojoSort™ nanobeads work in commonly used separation columns, based on our internal research as well as validation by external testing by academic labs. This simple protocol consists of following the MojoSort™ protocol to label the cells with **pre-diluted** MojoSort™ reagents and using the columns as indicated by the manufacturer.

Note: Due to the properties of our beads, it may be possible to use far fewer beads than with other commercial suppliers. We recommend a titration to find the best dilution factor. However, as a general rule, dilutions ranging from 1:3 to 1:20 for the Nanobeads can be used. Please contact BioLegend Technical Service (tech@biolegend.com) if further assistance is needed.

Guidelines

MojoSort™ magnetic particles can be used with other commercially available magnetic separators, both free standing magnets and column-based systems. Because MojoSort™ protocols are optimized for the MojoSort™ separator, the protocols may need to be adjusted for other systems. Please contact BioLegend Technical Service (tech@biolegend.com) for more information and guidance. We do not recommend using MojoSort™ particles for BD's IMag™ or Life Technologies' DynaMag™.

Materials

MATERIALS

 MojoSort™ Buffer **BioLegend Catalog #480017**

 MojoSort™ Nanobeads **BioLegend**

Additional reagents:

- commercially available cell separation columns
- 5 mL polypropylene tubes
- 70 µm cell strainer



- 1 Prepare cells from your tissue of interest or blood without lysing erythrocytes.
- 2 In the final wash of your sample preparation, resuspend the cells in MojoSort™ Buffer by adding up to 4 mL in a 5 mL (12 × 75 mm) polypropylene tube.
Note: Keep MojoSort™ Buffer on ice throughout the procedure.
- 3 Filter the cells with a 70 µm cell strainer, centrifuge at 300xg for 5 minutes, and resuspend in an appropriate volume of MojoSort™ Buffer. Count and adjust the cell concentration to 1×10^8 cells/mL.
- 4 Aliquot 100 µL (10^7 cells) into a new tube.
- 5 Vortex the antibody-conjugated Nanobeads (to resuspend) at max speed, 5 touches, and prepare the dilutions to test. **Add 10 µL of pre-diluted conjugated Nanobeads.** Mix well and **incubate on ice for 15 minutes.** Scale up the volume accordingly if separating more cells. For example, add 100 µL of pre-diluted Nanobeads for separating 1×10^8 cells in 1 ml of MojoSort™ Buffer. When working with less than 10^7 cells, use indicated volumes for 10^7 cells.
- 6 Add the appropriate amount of MojoSort™ Buffer and proceed to separation. At least 500 µL is needed for column separation.
Note: There are several types of commercially available columns, depending on your application. Choose the one that fits best your experiment:

	Max. number of labeled cells	Max. number of total cells	Cell suspension volume	Column rinse volume	Cell wash volume	Elution volume
Small Capacity	1×10^7	2×10^8	500µL for up to 10^8 cells	1ml	1 ml	1 ml
Medium Capacity	1×10^8	2×10^9	500µL for up to 10^9 cells	3ml	3 ml	5 ml
Large Capacity	1×10^9	2×10^{10}	500µL for up to 10^{10} cells	20-50ml	30 ml	20 ml

Example of magnetic separation with medium capacity columns:

- 7 Place the column in a magnetic separator that fits the column.
- 8 Rinse the column with 3 mL of cell separation buffer.



- 9 Add the labeled cell suspension to the column through a 30 μ m filter and collect the fraction containing the unlabeled cells.
- 10 Wash the cells in the column **3 times** with 3 mL of buffer and collect the fraction containing the unlabeled cells. Combine with the collected fraction from step 3. These cells may be useful as controls, to monitor purity/yield, or other purposes.
- 11 Take away the column from the magnet and place it on a tube. Then add 5 mL of buffer and flush out the magnetically labeled fraction with a plunger or supplied device. These are the positively isolated cells of interest; do not discard. To increase the purity of the magnetically labeled fraction repeat the isolation process with a new, freshly prepared column.