



MojoSort™ Mouse CD45 Nanobeads Protocol - Depletion

DOI

dx.doi.org/10.17504/protocols.io.7wkhpcw



Sam Li¹

¹BioLegend

BioLegend

Tech. support email: tech@biolegend.com



Sam Li

BioLegend

Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account



DOI: <https://dx.doi.org/10.17504/protocols.io.7wkhpcw>

External link: <https://www.biolegend.com/protocols/mojosort-mouse-cd45-nanobeads-protocol-depletion/4751/>

Protocol Citation: Sam Li . MojoSort™ Mouse CD45 Nanobeads Protocol - Depletion. **protocols.io**

<https://dx.doi.org/10.17504/protocols.io.7wkhpcw>

License: This is an open access protocol distributed under the terms of the **Creative Commons Attribution License**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Created: October 02, 2019



Last Modified: October 02, 2019

Protocol Integer ID: 28332

Keywords: mojosort, cd45, nanobeads, depletion, isolation, selection, targeted cell, cells per tube, nanobead, specific cell number, cell, functional assay

Abstract

Product description and procedure summary: The cells targeted by the Nanobeads are depleted by incubating your sample with the directly conjugated magnetic particles. The magnetically labeled fraction is retained by the use of a magnetic separator. After collection of the targeted cells, downstream applications include functional assays, gene expression, phenotypic characterization, etc.

Note: This procedure is optimized for the isolation of 10^7 to 2×10^8 cells per tube. If working with fewer than 10^7 cells, keep volumes as indicated for 10^7 cells. For best results, optimize the conditions to your specific cell number and tissue. Prepare fresh MojoSort™ Buffer solution by diluting the 5X concentrate with sterile distilled water. *Scale up volumes if using 14mL tubes and Magnet, and place the tube in the magnet for 10 minutes.*

Guidelines

MojoSort™ magnetic particles can be used with other commercially available magnetic separators, both free standing magnets and column-based systems. Because MojoSort™ protocols are optimized for the MojoSort™ separator, the protocols may need to be adjusted for other systems. Please contact BioLegend Technical Service (tech@biolegend.com) for more information and guidance. We do not recommend using MojoSort™ particles for BD's IMag™ or Life Technologies' DynaMag™.

Application notes: To use this product in magnetic separation columns, a titration of the Nanobeads should be performed. Optimal concentration for magnetic separation columns is lot-specific. Please contact BioLegend Technical Service (tech@biolegend.com) for further assistance on how to use MojoSort™ Nanobeads in magnetic separation columns.

Materials

MATERIALS

⊗ MojoSort™ Magnet **BioLegend Catalog #480019**

⊗ MojoSort™ Buffer **BioLegend Catalog #480017**

⊗ MojoSort™ Mouse CD45 Nanobeads **BioLegend Catalog #480027, 480028**

- Adjustable pipettes
- 70µm filters (one per sample)
- 5mL (12 × 75mm) or 14mL (17 × 100mm) polypropylene tubes
- Reagents for sample preparation
- Reagents and instruments (Flow cytometer) to determine yield and purity

Troubleshooting



- 1 Prepare cells from your tissue of interest or blood without lysing erythrocytes.
- 2 In the final wash of your sample preparation, resuspend the cells in MojoSort™ Buffer by adding up to 4 mL in a 5 mL (12 × 75 mm) polypropylene tube.
Note: Keep MojoSort™ Buffer on ice throughout the procedure.
- 3 Filter the cells with a 70µm cell strainer, centrifuge at 300xg for 5 minutes, and resuspend in an appropriate volume of MojoSort™ Buffer. Count and adjust the cell concentration to 1×10^8 cells/mL.
- 4 Aliquot 100 µL of cell suspension (10^7 cells) into a new tube.
- 5 Resuspend the beads by vortexing, maximum speed, 5 touches. Add **10µL of Antibody Nanobeads**. Mix well and **incubate on ice for 15 minutes**. Scale up the volume accordingly if separating more cells. For example, add 100 µL of Nanobeads for separating 1×10^8 cells in 1 ml of MojoSort™ Buffer. When working with less than 10^7 cells, use indicated volumes for 10^7 cells.
- 6 Wash the cells by adding MojoSort™ Buffer up to 4mL. Centrifuge the cells at 300xg for 5 minutes.
- 7 Discard the supernatant.
- 8 Add 2.5mL of MojoSort™ Buffer.
Note: If you observe aggregates, filter the suspension. To maximize yield, you can disrupt the aggregates by pipetting the solution up and down.
- 9 Place the tube in the magnet for 5 minutes.
Optional: Take a small aliquot before placing the tube in the magnet to monitor purity and yield. Keep unused cells to be used as control or other applications if needed.
- 10 Pour out the unlabeled fraction (THESE ARE YOUR CELLS OF INTEREST). Set aside the labeled fraction.
- 11 Place the **unlabeled** fraction tube in the magnet for 5 minutes. This will be a total of **2 separations (once on the labeled cell mixture and once on the unlabeled fraction)**. Pool the unlabeled fractions. The labeled fractions may be useful as staining controls, to monitor purity/yield, or other purposes.
Optional: Take a small aliquot to monitor purity and yield.

