MojoSort™ Human anti-PE Nanobeads Protocol

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ABSTRACT

Product description and procedure summary: Target cells are positively selected or depleted by incubating the sample with an anti-human PE conjugated antibody, followed by incubation with magnetic anti-PE Nanobeads. The magnetically labeled fraction is retained by the use of a magnetic separator. These are the PE+ cells, do not discard them if those are the cells of interest. Some of the downstream applications include functional assays, gene expression, phenotypic characterization, etc.

Note: This procedure is optimized for the isolation of 10^7 to 2×10^8 cells per tube. If working with fewer than 10^7 cells, keep volumes as indicated for 10^7 cells. For best results, optimize the conditions to your specific cell number and tissue. Prepare fresh MojoSort™ Buffer solution by diluting the 5X concentrate with sterile distilled water. *Scale up volumes if using 14 mL tubes and Magnet, and place the tube in the magnet for 10 minutes.*

GUIDELINES

MojoSort™ magnetic particles can be used with other commercially available magnetic separators, both free standing magnets and column-based systems. Because MojoSort™ protocols are optimized for the MojoSort™ separator, the protocols may need to be adjusted for other systems. Please contact BioLegend Technical Service (tech@biolegend.com) for more information and guidance. We do not recommend using MojoSort™ particles for BD's IMag™ or Life Technologies' DynaMag™.

Application notes: To use this product in magnetic separation columns, a titration of the Nanobeads should be performed. Optimal concentration for magnetic separation columns is lot-specific. Please contact BioLegend Technical Service (tech@biolegend.com) for further assistance on how to use MojoSort™ Nanobeads in magnetic separation columns.

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MATERIALS

MATERIALS

✘ MojoSort™ Buffer **BioLegend Catalog #480017**

✘ MojoSort™ Magnet **BioLegend Catalog #480019**

✘ MojoSort™ Human anti-PE Nanobeads **BioLegend Catalog #480091**

- Adjustable pipettes
- 70µm filters (one per sample)
- 5mL (12 x 75mm) or 14mL (17 x 100mm) polypropylene tubes
- Reagents for sample preparation
- Reagents and instruments (Flow cytometer) to determine yield and purity

- 1 Prepare cells from your tissue of interest or blood without lysing erythrocytes. Kits for human samples have been optimized for PBMCs, please prepare the cells using a suitable method.
- 2 In the final wash of your sample preparation, resuspend the cells in MojoSort™ Buffer by adding up to 4 mL in a 5 mL (12 x 75 mm) polypropylene tube.
Note: Keep MojoSort™ Buffer on ice throughout the procedure.
- 3 Filter the cells with a 70µm cell strainer, centrifuge at 300xg for 5 minutes, and resuspend in a small volume of MojoSort™ Buffer. Count and adjust the cell concentration to 1×10^8 cells/mL by adding MojoSort™ Buffer.
- 4 Aliquot 100 µL of cell suspension (10^7 cells) into a new tube. **Add 5µL of Human TruStain FcX™ (Fc Receptor Blocking Solution)**, mix well and **incubate at room temperature for 10 minutes**. Scale up the volume accordingly if separating more cells. For example, if the volume of Human TruStain FcX™ for 1×10^7 cells is 5µL, add 50µL for 1×10^8 cells. When working with less than 10^7 cells, use indicated volumes for 10^7 cells.
- 5 Check the recommended usage for flow cytometric staining of the PE-conjugated antibody indicated in the antibody technical datasheet. Calculate the volume to stain 10^7 cells (or desired amount of cells). **Add the appropriate volume of PE-conjugated antibody** to the cell suspension, mix well and **incubate on ice for 15 minutes**.
Optional: Take an aliquot before adding the antibody to monitor purity and yield.
- 6 Wash the cells by adding MojoSort™ Buffer up to 4 mL; centrifuge the cells at 300xg for 5 minutes.
- 7 Discard supernatant and resuspend in 100 µL of MojoSort™ Buffer.

- 8** Resuspend the beads by vortexing, maximum speed, 5 touches. **Add the appropriate volume of Human anti-PE Nanobeads** to the cell suspension, mix well and **incubate on ice for 15 minutes**. The volume of MojoSort™ Human anti-PE Nanobeads should be adjusted depending on starting percentage of cell type to be isolated. For 1×10^7 cells in 100 μ L of buffer, use the following volumes:

Cell type	Cell Frequency in PBMCs	Volume of MojoSort™ anti-PE Nanobeads (for 1×10^7 cells in 100 μ L of buffer)
CD4 (including monocytes)	50-55%	20 μ L
CD8a (including low expressing cells)	15-20%	10 μ L
CD19	5-10%	4 μ L
TCR V β 9	1-3%	2 μ L

For low frequency cells, pre-dilute the Nanobeads in order to pipette a minimum of 5 μ L of any solution. For example, to isolate TCR V γ 9+ cells, pre-dilute 10 μ L of PE Nanobeads in 40 μ L of MojoSort™ buffer and add 10 μ L of that dilution per sample. Avoid working with small volumes.

Scale up the volume accordingly if separating more cells. For example, if the volume of Nanobeads for 1×10^7 cells is 10 μ L, add 100 μ L for 1×10^8 cells. When working with less than 10^7 cells, use indicated volumes for 10^7 cells.

- 9** Wash the cells by adding MojoSort™ Buffer up to 4mL; centrifuge the cells at 300xg for 5 minutes.

- 10** Discard the supernatant.

- 11** Add 2.5mL of MojoSort™ Buffer.
- Note:** If you observe aggregates, filter the suspension. To maximize yield, you can disrupt the aggregates by pipetting the solution up and down.
- 12** Place the tube in the magnet for 5 minutes.
- Optional: Take a small aliquot before placing the tube in the magnet to monitor purity and yield. Keep unused cells to be used as control or other applications if needed.*
- 13** Pour out the unlabeled fraction. If these are your cells of interest, **DO NOT DISCARD**. Resuspend the labeled cells in 2.5mL MojoSort™ Buffer.
- 14** Repeat steps 11-13 on the labeled fraction twice more for a total of **3 separations**. Pool the unlabeled fractions and keep the labeled cells. The fraction that is not of interest may be useful as staining controls, to monitor purity/yield, or other purposes.
- Optional: Take a small aliquot to monitor purity and yield.*