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Mitophagy detection assay

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Protocol status: Working

We use this protocol and it's working

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Abstract

Detection of mitophagy flux in neurons using Dojindo Mtpthaggy dye and LysoTracker Green

Protocol materials

 Mtpthaggy Dye **Dojindo Catalog #MD01**

 LysoTracker Green **Thermo Fisher Scientific Catalog #L7526**

Before start

Make maintenance media (MM) and imaging media (IM) for cortical neurons



- 1 Plate 120,000-150,000 neurons in an imaging dish at *days in vitro* (DIV) 0

Note

Protocol for primary neuron isolation and in vitro culture can be obtained from:
[dx.doi.org/10.17504/protocols.io.81wgby723vpk/v1](https://doi.org/10.17504/protocols.io.81wgby723vpk/v1)

Preparing reagents

- 2 On the day of imaging, equilibrate 12 ml of fresh maintenance media (MM) per imaging dish in an incubator at  37 °C

Note

Maintenance media composition can be obtained from:
[dx.doi.org/10.17504/protocols.io.81wgby723vpk/v1](https://doi.org/10.17504/protocols.io.81wgby723vpk/v1)

- 3 Prepare intermediate dilution of DMP Red (commercially available as  Mtphagy Dye **Dojindo Catalog #MD01**) in MM: for each imaging dish, make  500 µL of fresh MM containing  0.5 µL of DMP Red

- 4 Prepare intermediate dilution of LysoTracker Green in MM: for each imaging dish make  50 µL of fresh MM containing  0.1 µL of LysoTracker Green
 LysoTracker Green **Thermo Fisher Scientific Catalog #L7526**

- 5 Prepare imaging media (IM) with LysoTracker Green- for each imaging dish make  50 µL of fresh MM containing  0.1 µL of of LysoTracker Green

Note

Imaging media composition for 50 ml:

45% Glucose 660 µL
B-27 1 mL
Hibernate E Up to 50 mL



Imaging steps

3h 30m

- 6 At DIV 6 or 7, aspirate out conditioned media from the imaging dish, wash the neurons once with fresh MM
- 7 Add  1.6 mL of fresh MM and then add  0.4 mL of MM with the intermediate dilution of DMP Red (prepared in step 3)
- 8 Incubate the neurons in an incubator at  37 °C for  00:30:00 30m
- 9 Post incubation, aspirate out the media, and wash once with fresh MM
- 10 Add  2 mL of fresh MM containing 3 nM Antimycin A or an equivalent volume of 100% ethanol
- 11 Incubate the neurons in an incubator at  37 °C for  01:00:00 1h
- 12 Post incubation, aspirate out the media, and replace it with fresh MM **with no Antimycin A or ethanol**
- 13 Incubate the neurons in an incubator at  37 °C for  01:30:00 1h 30m
- 14 Post incubation add  50 µL of MM with the intermediate dilution of LysoTracker Green (prepared in step 4)
- 15 Incubate the neurons in an incubator at  37 °C for  00:30:00 30m
- 16 Post incubation, aspirate out the media, and wash once with fresh IM
- 17 Add  2 mL of fresh IM followed by  50 µL of IM with the intermediate dilution of LysoTracker Green (prepared in step 5)
- 18 Image neurons live under a confocal microscope with a heating chamber kept at  37 °C

