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Microbiome Assay with 96WP Updated April 2020

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Behavioural Genomics

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Protocol status: Working
We use this protocol and it's working

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Troubleshooting

-10 days

1 Pick 10 L4s onto 10×90mm OP50 seeded platese.gon Monday (10 L4 per plate so total 100L4s)

-6 days

2 At 2pm on day of bleaching (eg Friday) follow the protocol *Bleach synchronization of C elegans*

3 Keep the tube with bleached N2s on a rotator at 20C incubator until refeed (make sure not to exceed 5days as the worm behaviour is not consistent post this time frame)

-3 days

4 If tracking is intended to be performed on the following Thursday, then refeed the arrested L1s onto 4 OP50 seeded 150mm plates on the Monday (+72hrs post bleaching, at 2pm following the protocol Bleach synchronization of C elegans

5 Store the refed L1 plates at 20C incubator

-2days

6 At least 2days prior to tracking day (e.g. Tuesday if tracking is planned on Thursday) make about 250ml no peptone NGM

7 Dispense 200ul of agar into each well of the 96WP using the integra viafill following the protocol *Dispensing agar into multi well plates*

8 Let the agar dry and store the plates at 4C (lid side down) until used (plates can be stored for up to a week)-Note:

9 Measure the weight of the plates with lids on and record average plate weight on day of pouring

10 Grow an overnight culture of the bacterial strain from the (shuffled) 96WP library stock plates, 2days prior to tracking (eg on a Tuesday afternoon if tracking is to be intended on Thursday) following the protocol *Growing overnight bacterial culture in 96WP.*

11 Cover the plates with breathable seals and shake at 180-200rpm in a shaking incubator

12 Separately inoculate OP50 and incubate at 37C overnight in a shaking incubator at 200 rpm

-1day

13 Take the bacterial cultures out of the incubators in the morning, and store at 4C until used for seeding later that day

14 Take agar-filled 96WPs out of the cold room and dry under in the drying cabinet for about 2-3hrs (Measure the weight of 3 plates before drying without the lid)

15 After drying measure the weight of the 3 plates again. Ensure that the plates lose between 3-5% of their original weight following drying.

16 Seed the dried plates with the bacterial culture grown the night before using the ViaFlo

17 Turn the ViaFlo ON (Make sure to book it beforehand)

18 Edit the protocol SAUL to have the mixing stage consisting of 15 cycles to initially mix the bacterial cultures and to resuspend each strain

19 Load the tips (long 12ul tips), Adjust the Z height to the one that worked best in the test runs (See lab notes)

20 Mix the bacterial plate using the ViaFlo

21 Change the tips

22 Edit protocol SAUL again to have mixing stage be of 1 cycle when actually seeding plates

23 Run the protocol- To dispense 10ul volume in each well

24 Change tips every time a new plate is seeded

25 After all the plates are seeded dry them under the hood for another 1hr

26 After drying put the plates in a closed box upside down in the cold room (4C)

Day 0: Tracking day

27 On the day of tracking prepare the worms following the protocol *Preparing worms for the COPAS (wormsorter)*

28 Dispense the worms using the COPAS following the protocol *Using the worm sorter (COPAS)*

29 Dry the plates for 30minutes under the hood

30 After drying, place the plates under the Hydra rig, leave for a further 30mins to acclimatize, and then record for 15mins, following the protocol *Tracking on the Hydra rigs*

31 Record with the lids on and lid side facing up inside the rig. Makesure to wipe the lids with lint-free tissues before placing them under the rig. Also use the script that involves using blue light in the recording

32 Record each plate at four time points: 1hr, 3hrs, 5hrs, and 23.5hrs (next day). Between 1hr, 3hr & 5hr the plates are kept in the cave outside the rigs

33 Store the plates in 20C incubator overnight between tracking 5hrs and 23.5hrs timepoint

34 Discard the plates in the biological waste bins post tracking