

Mar 20, 2023

Version 2

Measurement of biogenic silica from plankton V.2

DOI

<https://dx.doi.org/10.17504/protocols.io.8epv5jjzjl1b/v2>



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Protocol Citation: Ying-Yu Hu, Nuwanthi Samarasinghe, Zoe V. Finkel 2023. Measurement of biogenic silica from plankton. protocols.io <https://dx.doi.org/10.17504/protocols.io.8epv5jjzjl1b/v2> Version created by [Ying-Yu Hu](#)

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Protocol status: Working

We use this protocol and it's working

Created: December 17, 2022

Last Modified: March 20, 2023

Protocol Integer ID: 74131

Keywords: biogenic silica, dissolution, silicomolybdic acid, measurement of biogenic silica, measuring biogenic silica, biogenic silica from plankton, biogenic silica, amorphous silica, molybdate measurement technique, plankton, diatomaceous earth, sodium carbonate, accuracy of the measurement, production of monosilicic acid

Funders Acknowledgements:

CBIOMES

Grant ID: 549937

SCOPE-Gradients

Grant ID: 723789

Abstract

Here, we present a method for measuring biogenic silica from plankton. Biogenic silica is digested using a wet-alkaline method, in which 2 M sodium carbonate is used to hydrate and depolymerize amorphous silica, resulting in the production of monosilicic acid. The molybdate measurement technique is based on the method described by Shemesh et al. (1988) and follows the JGOFS protocols outlined by UNESCO (1994).

To ensure the accuracy of the measurement, Celite S diatomaceous earth is used as a check standard for the recovery of biogenic silica. Our method yields a recovery rate of 85% to 90%.

Citation

Shemesh, Aldo; Mortlock, Richard A; Smith, R J; Froelich, Philip N (1988)
. Determination of Ge/Si in marine siliceous microfossils: separation, cleaning and dissolution of diatoms and radiolaria.
Marine Chemistry.

[https://doi.org/10.1016/0304-4203\(88\)90113-2](https://doi.org/10.1016/0304-4203(88)90113-2)

LINK



Protocol materials

- ☒ Celite S diatomaceous earth **Merck MilliporeSigma (Sigma-Aldrich) Catalog #06858**
- ☒ Ammonium molybdate **Merck MilliporeSigma (Sigma-Aldrich) Catalog #09878-100G**
- ☒ Oxalic acid dihydrate **VWR International (Avantor) Catalog #BDH4556-500G**
- ☒ Sodium hexafluorosilicate **Merck MilliporeSigma (Sigma-Aldrich) Catalog #250171**
- ☒ Sodium sulfite **Fisher Scientific Catalog #S430-500**
- ☒ 4-(methyldamino)phenol hemisulfate salt **Merck MilliporeSigma (Sigma-Aldrich) Catalog #320013**
- ☒ Methyl orange **Merck MilliporeSigma (Sigma-Aldrich) Catalog #1013230250**



Sample collection

- 1 Estimation:
The low limit of detection is approximately 0.6 uM silicate in the molybdate method. For siliceous plankton, sample requires no less than 4 ug PON (particulate organic nitrogen) per filter when using a 50 mL volumetric flask, or 2 ug PON per filter when using a 25 mL volumetric flask. The sampling volume for biogenic silica samples is approximately 10% of the PON sample volume. For seawater samples, the sampling volume for biogenic silica samples should be determined based on the community composition.
- 2 Filter blank media (without cells, same volume as plankton samples) through polycarbonate filter as blank
- 3 Transfer filter into 2 mL cryogenic vial
- 4 Flash freeze and store at  -20 °C
- 5 Filter plankton sample in liquid media onto polycarbonate filters, using gentle vacuum pressure (130 mmHg)

Equipment

Filter forceps

NAME

blunt end, stainless steel

TYPE

Millipore

BRAND

XX6200006P

SKU

- 6 Rinse filter funnel with filtered artificial seawater without macronutrients
- 7 Transfer filter with sample into 2 mL cryogenic vial



- 8 Flash freeze and store at  -20 °C
- 9 Transfer sample to 50 mL falcon tube with clean filter forceps (rinsed by 95% ethanol and air-dried), dry at  90 °C in the airforce oven.

Equipment

Forced air oven	NAME
VWR	BRAND
89511-410	SKU

Equipment

Falcon® Centrifuge Tubes	NAME
Polypropylene, Sterile, 50 mL	TYPE
Corning®	BRAND
352070	SKU

Standard primary solution and reagents

- 10 Molybdate reagent stock solution

Note

Require 100 uL per sample

10.1



Ammonium molybdate **Merck MilliporeSigma (Sigma-Aldrich) Catalog #09878-100G**

Ammonium paramolybdate:

$[(\text{NH}_4)_6\text{Mo}_7\text{O}_{24} \cdot 4\text{H}_2\text{O}]$

CAS: 12054-85-2

10.2

Add  1.6 g ammonium paramolybdate into a 125 mL plastic bottle and top to 100 g with MilliQ.

10.3

Store out of direct sunlight. Discard if white precipitation forms.

11

HCl stock solution

Note

Require 100 uL per sample

11.1

Use graduated cylinder, measure 95 mL MilliQ and transfer into a 125 mL plastic bottle.

11.2

In the fume hood, add  5 mL 12 N HCl into the bottle, mix well.

12

Metol-sulfite solution

Note

Require 100 uL per sample

12.1



4-(methylamino)phenol hemisulfate salt **Merck MilliporeSigma (Sigma-Aldrich) Catalog #320013**



Sodium sulfite **Fisher Scientific Catalog #S430-500**

12.2

Require:

(1) 50 mL syringe

(2) Syringe filter



Equipment

Syringe filter

NAME

0.2 um PES

TYPE

VWR

BRAND

28145-501

SKU

12.3 In a 100 to 250 mL plastic beaker, add  0.6 g sodium sulphite.

12.4 Add  1 g 4-(methyl amino)phenol hemisulfate.

12.5 Top to 50 g with MilliQ water.

12.6 Fill syringe with Metol-sulfite solution, filter through the syringe filter, collect filtrate into four 15 mL falcon tubes wrapped with foil, keep at room temperature.

12.7 Prepare fresh every month.

13 Oxalic acid solution

Note

Require 100 uL per sample.

13.1  Oxalic acid dihydrate **VWR International (Avantor) Catalog #BDH4556-500G**

13.2 In a 125 mL plastic bottle, add  6 g oxalic acid and top to 100 g.



13.3 Let the solution stand at room temperature overnight.

13.4 Decant the solution from the crystals into a plastic bottle.

13.5 Keep at room temperature.

14 Sulphuric acid (30%)

14.1

Note

Require 100 uL per sample

14.2 Mix 3 part concentrated sulphuric acid into 7 part of MilliQ
Cool down to room temperature

Note

This can be prepared on Day 2 prior to molybdate reaction

15 Primary silica standard solution (~ 1 mM Si)

15.1



Sodium hexafluorosilicate **Merck MilliporeSigma (Sigma-Aldrich) Catalog #250171**

15.2 Transfer 1 g sodium fluorosilicate in a plastic vial

15.3 Keep the vial in a vacuum desiccator overnight to remove excess water (do not heat or fuse)



- 15.4 In a one litre plastic volumetric flask, dissolve ~  0.1881 g (log the actual mass) of dry sodium fluorosilicate in MilliQ water and top to 1 L with MilliQ water.
- 15.5 It takes about 30 min to complete the dissolution. This cannot be rushed.
- 15.6 Store in a plastic bottle at room temperature.

Day 1: Dissolution

16  2 M Na_2CO_3 (18.69%)

Note

Need to be freshly prepared.
The old reagent can yield high blank possibly by leaching silicate from plastic material.

- 16.1 Each sample requires 10 mL 2 M Na_2CO_3
- 16.2 Weigh 186.9 g Na_2CO_3 in a weighing dish.
(CAS: 497-19-8, FW 105.99)
- 16.3 Tare a 1 L plastic erlenmeyer flask
- 16.4 Transfer Na_2CO_3 into the flask
- 16.5 Top to 1000 g with MilliQ and shake until all salt is completely dissolved.
- 16.6 Aliquot the solution into four 250 mL plastic bottles.



17 Turn on airforce oven to  85 °C

18 In the fume hood, transfer diatomaceous into a 5 mL plastic tube for weighing convenience (the original package is 1 kg).

Note

Diatomaceous is used as a check standard for the recovery of biogenic silica

Safety information

Diatomaceous:

Upper respiratory irritant. May cause coughing or throat irritation. Breathing dust containing crystalline silica over a long period may cause lung damage.



Celite S diatomaceous earth **Merck MilliporeSigma (Sigma-Aldrich) Catalog #06858**

18.1 Weigh 100~200 ug diatomaceous into 50 mL falcon tube, in triplicate. Log the actual weight.

Safety information

Do not open the container until the static charge of diatomaceous powder has been neutralized by ionization blower.

Note

Less than 100 ug sample might introduce more error amongst the replicates in recovery.

18.2 Prepare one empty 50 mL falcon tube as the reagent blank for diatomaceous.

19 Add  10 mL 2 M Na₂CO₃ to each tube, including:

- reagent blank for check standards
- check standards



- blank for samples
- samples

20 Vortex

21 Loosen the caps and place all tubes into the airforce oven overnight (for example, from 5 pm to 9 am).

16h

Day 2: Acidification

22 Volume of 12 N HCl required:
 $3.5 \text{ mL} \times N$

23 Transfer 12 N HCl into a 50 mL Falcon tube in the fume hood.

24 Work on one tube at a time, and leave other tubes in the oven.

25 In the fume hood, add  30 μL Methyl orange into the tube.

 Methyl orange **Merck MilliporeSigma (Sigma-Aldrich) Catalog #1013230250**

26 Add MilliQ until the volume of solution in the falcon tube is 10 mL.

Note

The original volume of Na_2CO_3 is reduced due to evaporation of water during 20-h dissolution.

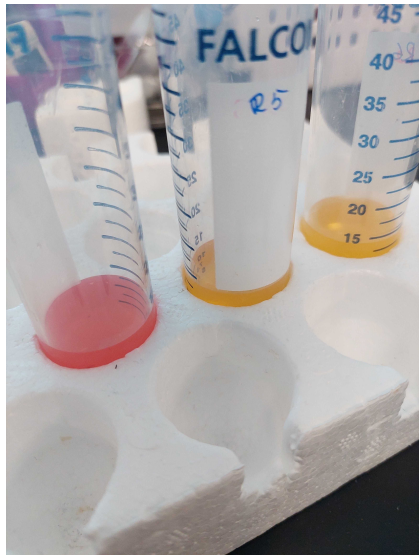
27 Dropwise add  3 mL 12 N HCl by using 1000 uL pipet.

Safety information

Do it slowly. Swirl the tube until reaction stops and then add the next drop. The most vigorous reaction is at about 3 mL 12 N HCl.

28 Switch to a 100 uL pipette, add 100 uL at a time. Near the equivalence point, when the colour starts to change to pink more markedly but after mixing the orange colour returns,

it is necessary to add HCl **drop by drop**. The first drop that causes a permanent colour change to pink determines the equivalence point. Stop adding HCl. Cap the tube, hold tube horizontally, gently invert the tube to wash residue at the inner side of the cap down to the solution. The color may change back to orange, add more drops of HCl until the color turns to permanent pink again (See the color of the left tube).



Note

We have found that the optimal pH for the reaction between silicate and molybdate to form silicomolybdic acid is 3 to 4. Too low or too high pH decreases recovery of biogenic silica.

The acidified solution yields pH at 2 to 3. It is diluted to 10% in the molybdate assay, which gives pH at 3 to 4.

- 29 Transfer resulted solution from falcon tube to 25 or 50 mL polypropylene volumetric flask.

Note

Be careful while transferring the solution and ensure that the filter does not fall out of the tube, which spills the solution and causes sample loss.

- 30 Use MilliQ to rinse the tube **three times** and transfer all samples into the volumetric flask.

Note

If a 50 mL volumetric flask is used, rinse the falcon tube with 5 mL of MilliQ at a time.
If a 25 mL volumetric flask is used, rinse the falcon tube with 1 mL of MilliQ at a time.

31 Use transfer pipet, top final volume to 25 or 50 mL with Milli-Q.

32 Shake and thoroughly mix the solution.

Note

Before mixing, check the cap to avoid leaking

33 Transfer solution from volumetric flask to a clean and labelled Falcon tube.

Day 2: Molybdate reaction

3h

34 Secondary standard solution (Freshly prepared prior to the assay)

50 uL primary stock solution

450 uL MilliQ

35 Standard working solutions (Freshly prepared prior to the assay)

	Standards	Secondary (uL)	MilliQ (uL)	Conc. (uM)
	S1	0	500	0
	S2	5	495	1
	S3	10	490	2
	S4	20	480	4
	S5	40	460	8
	S6	60	440	12
	S7	80	420	16
	S8	100	400	20



36 Vortex and then transfer  50 μL from (1) blank for check standards, (2) check standards, (3) blank for samples, and (4) samples into labelled 2 mL microtubes.

37 Add  450 μL MilliQ into each tube to obtain a 10% dilution.

38 Molybdate working solution

Note

Require 200 μL per sample

38.1 **1 part** Molybdate stock reagent
1 part HCl stock reagent

39

Safety information

The addition of reagent must be operated in the fume hood. Acidified sodium fluorosilicate may contain some hydrofluoric acid.

40 Add  200 μL Molybdate reagent into each tube.

41 Vortex each tube and then shake at  Room temperature for  00:15:00 for the formation of silicomolybdic acid.

15m

42 Reducing solution

Note

Require 300 μL per sample

42.1 **1 part** Metol-sulfite solution
1 part oxalic acid solution
1 part sulphuric acid solution

43 Add  300 μL reducing solution into each tube.

44 Vortex each tube and then shake at 🌡 Room temperature for 🕒 03:00:00

3h

45 Measure pH of each sample (in the Falcon tube)

	Sample code (example)	Sample code	pH
	Blank for check standards		
	Check standard 1		
	Check standard 2		
	Check standard 3		
	Blank for samples		
	Sample 1		
	Sample 2		

Day 2: Colorimetric measurement

3h

46 In the fume hood, vortex each tube and then load 250 µL of the sample into one well of the microplate. Vortex again and load the same sample into another well of the microplate as replicate.

Equipment

96-Well Microplates, Polystyrene, Clear, NAME
Greiner Bio-One BRAND
655101 SKU

47 Setup the layout.

48 Setup the program

	A	B
	Shake duration	00:00:05
	Shaking type	Continuous
	Shaking force	High
	Shaking speed [rpm]	600
	Wavelength [nm]	812
	Use transmittance	No
	Pathlength correction	No
	Measurement Time [ms]	100

Equipment

Varioskan LUX Multimode Microplate Reader NAME
Thermo Fisher BRAND
VL0L00D0 SKU



49 Read the samples.

50 Export data sheet to excel.

Waste disposal

3h

51 Collect all solution with paramethylaminophenol sulphate and sodium fluorosilicate into the waste container.

52 Rinse microtubes and microplate with tap water, dispose in blue recycling bin.

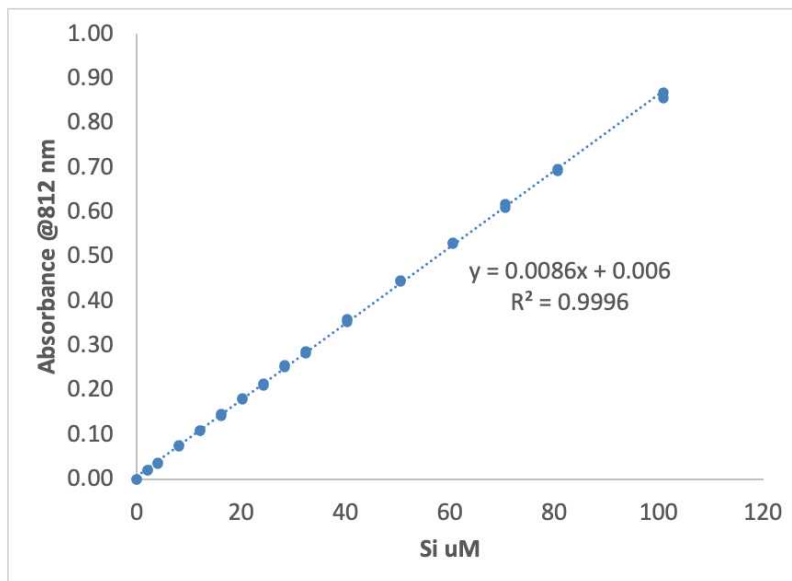
Day 2: Calculation

3h

53 Subtract the average absorbance at 812 nm of the blank standard replicates from the absorbance at 812 nm of all other standard working solutions.

54 Subtract the average absorbance at 812 nm of the blank sample (i.e. blank filter) replicates from the absorbance at 812 nm of all other individual samples.

55 Prepare a standard curve by plotting the average blank-corrected 812 nm absorbance for each standard working solution versus its concentration in μM .



56 Use the standard curve to determine the silicate concentration of each unknown sample by using its blank-corrected 812 nm absorbance.

57 Si per sample = Si X V X (0.001) X DF

	Variable	Unit	Definition
	Si per sample	umol	element Si in the sample collected
	Si	uM	silicate concentration calculated from the standard curve
	V	mL	volume of volumetric flask
	DF		From volumetric flask to the microtube, DF=10

58 % Diatomaceous recovery = 100 X Si X V X (0.001) X MW X DF /(M_{ug} X Purity X 0.01)

	Variable	Unit	Definition
	% Diatomaceous recovery		percentage recovery of diatomaceous
	Si	uM	silicate concentration calculated from the standard curve
	V	mL	volume of volumetric flask



Variable	Unit	Definition
MW	ug/umol	molecular mass of SiO ₂ , i.e. 60.08
DF		From volumetric flask to the microtube, DF=10
M	ug	actual mass of diatomaceous
Purity		purity of SiO ₂ in Celite S diatomaceous earth (06858) is 90.2%

Note

The recovery should be around 85 to 90%.

Citations

Shemesh, Aldo; Mortlock, Richard A; Smith, R J; Froelich, Philip N. Determination of Ge/Si in marine siliceous microfossils: separation, cleaning and dissolution of diatoms and radiolaria

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