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MAIT Cell Expansion in Donor Mice

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Abstract

This is part 3.1 of the "Study of MAIT Cell Activation in Viral Infections In Vivo" collection of protocols.

Collection Abstract: MAIT cells are abundant, highly evolutionarily conserved innate-like lymphocytes expressing a semi-invariant T cell receptor (TCR), which recognizes microbially derived small intermediate molecules from the riboflavin biosynthetic pathway. However, in addition to their TCR-mediated functions they can also be activated in a TCR-independent manner via cytokines including IL-12, -15, -18, and type I interferon. Emerging data suggest that they are expanded and activated by a range of viral infections, and significantly that they can contribute to a protective anti-viral response. Here we describe methods used to investigate these anti-viral functions in vivo in murine models. To overcome the technical challenge that MAIT cells are rare in specific pathogen-free laboratory mice, we describe how pulmonary MAIT cells can be expanded using intranasal bacterial infection or a combination of synthetic MAIT cell antigen and TLR agonists. We also describe protocols for adoptive transfer of MAIT cells, methods for lung homogenization for plaque assays, and surface and intracellular cytokine staining to determine MAIT cell activation.

Abstract: MAIT cells are rare in specific pathogen-free mice [6], typically comprising about 1×10^4 recoverable pulmonary MAIT cells in an infection-naive adult C57BL/6 mouse. Therefore, for adoptive transfer experiments, the MAIT cell population should first be expanded using intranasal infection [15] or immunization (5-OP-RU with TLR agonists) [3, 15] (see Note 5). When planning the adoptive transfer experiment, estimate that one S. Typhimurium BRD509-infected mouse will yield $1-2 \times 10^6$ sorted MAITcells, which are enough for 10-20 recipient mice (10^5 MAIT cells/RAG2^{-/-} γ C^{-/-} mouse in this case). Infect donor mice 7 days earlier than the adoptive transfer.

Attachments



Materials

For materials, please refer to the Guidelines section of the '"<u>Study of MAIT Cell Activation in Viral Infections In</u> <u>Vivo</u>" collection.

Safety warnings

Personal protective equipment (PPE) should be worn at all times (gloves, lab coat, & eye protection) (*see* Notes 3 and 4).

For hazard information and safety warnings, please refer to the SDS (Safety Data Sheet).

- 1 Two days before infection streak out a plate of *S*. Typhimurium BRD509 (an attenuated vaccine strain [14]) on LB agar plates, containing [M] 50 μg/ml streptomycin and incubate plates Overnight at 37 °C.
- The day before infection, pick a single colony under flame and inoculate to
 ▲ 10 mL LB culture medium with IMI 50 µg/ml streptomycin and leave static at
 ③ 37 °C (double contained if working with wild type/virulent SL1344 or equivalent strains) ③ Overnight .
- 3 On the day of infection, re-inoculate into fresh

Δ 10 mL pre-warmed LB culture medium with Δ 0.5 μL , Δ 100 μL , or Δ 20 μL of overnight culture, under flame. This is to ensure an optimal optical density (0.D.) reading (bacteria in log phase growth) for preparing the inoculum later (*see* **Note 6**). The doublingtime for *Salmonella* can vary between 0.5 and 1 h. Make the infection inoculum from culture with 0.D._{600nm} reading between 0.2 and 0.6. Calculate the required CFU of bacteria estimating 1 O.D. = 5–10 × 10⁸ CFU (this constant needs to be established for individual labs). Dilute with PBS to a final concentration of **2 × 10⁷ CFU/mL**, allowing **50** μL inoculum/mouse, i.e., 10⁶ CFU/50 μL/mouse.

- 4 Infect mice i.n. with 10^6 CFU *S.* Typhimurium BRD509 in $\boxed{4}$ 50 µL PBS under isoflurane anesthesia (*see* **Notes 7** and **8**).
- 5 Allow mice to recover and monitor mice for 7 days to allow the infection to take its course and MAIT cell frequencies to expand dramatically from 10⁴ to 5 × 10⁶ MAIT cells, or from [M] 1 % to [M] 20 % [M] 50 % of all alpha-beta T cells [15] (*see* Note 8).