



Jun 06, 2019

Version 2

# 🌐 Machine learning approach yields epigenetic biomarkers of food allergy: A novel 13-gene signature to diagnose clinical reactivity V.2

📖 [PLOS One](#)

DOI

[dx.doi.org/10.17504/protocols.io.x7pfrmn](https://dx.doi.org/10.17504/protocols.io.x7pfrmn)

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DOI: <https://dx.doi.org/10.17504/protocols.io.x7pfrmn>

External link: <https://doi.org/10.1371/journal.pone.0218253>

**Protocol Citation:** Ayush Alag 2019. Machine learning approach yields epigenetic biomarkers of food allergy: A novel 13-gene signature to diagnose clinical reactivity. **protocols.io** <https://dx.doi.org/10.17504/protocols.io.x7pfrmn>

**Manuscript citation:**

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**Protocol status:** Working

**Created:** February 15, 2019

**Last Modified:** June 06, 2019

**Protocol Integer ID:** 20431

**Keywords:** Machine Learning, Food Allergy, DNA Methylation, Gene Signature, epigenetic biomarkers of food allergy, strong epigenetic biomarker, epigenetic biomarker, strong biomarker, food allergy, deep learning classifier, strong epigenetic biomarkers of fa, strong biomarkers of the disease, generalizability to dnam classification problem, classifier, gene signature, dnam classification problem, discriminating gene, using dna methylation, dna methylation, additional classifier, allergy, machine learning, novel gene association, perfect classification accuracy, dangerous oral food challenge, gene, dnam, computational machine learning approach, perfect classifier, dnam study, perfect classifier for this dataset, food, accurate diagnostic tests for those disease, dna

## Abstract

Current laboratory tests have a less than 50% accuracy in distinguishing between people who have food allergies (FA) and those who are merely sensitized to foods, resulting in the use of expensive and potentially dangerous Oral Food Challenges. Our study presents a purely-computational machine learning approach, conducted using DNA Methylation (DNAm) data, to accurately diagnose food allergies and find genes that are strong biomarkers of the disease.

We built two deep learning classifiers with twelve CpG-input features each that achieved perfect accuracy and an AUROC of 1 on the completely hidden cross-validation cohort. In addition, 24 additional classifiers were created that each had an average cross-validation accuracy of 98.35%. These 26 classifiers yielded a total of 18 unique CpGs, which mapped to 13 genes that are strong epigenetic biomarkers of FA.

Biological enrichment on the 13-gene signature yielded new insights. Notably, our FA-discriminating genes were strongly associated with the immune system, which helps validate our findings. Seven of the 13 genes overlapped with previous food-allergy and DNAm studies.

Previous studies have also created a perfect classifier for this dataset, but they used a 96-CpG input feature set built on both data-driven and a priori biological insights. Our study is an improvement on previous work because it maintains a perfect classification accuracy using only 18 highly discriminating CpGs (0.005% of the total available features). In machine learning, simpler models, as used in our study, are preferred over more complex ones (all other things being equal).

In addition, our completely data-driven approach eliminates the need for *a priori* information and allows for generalizability to DNAm classification problems in other disease areas, which may result in novel gene associations or accurate diagnostic tests for those diseases.

## Guidelines

The DNA Methylation data analysis has been done using the Java programming language. To follow the steps it is best to create an Eclipse project.

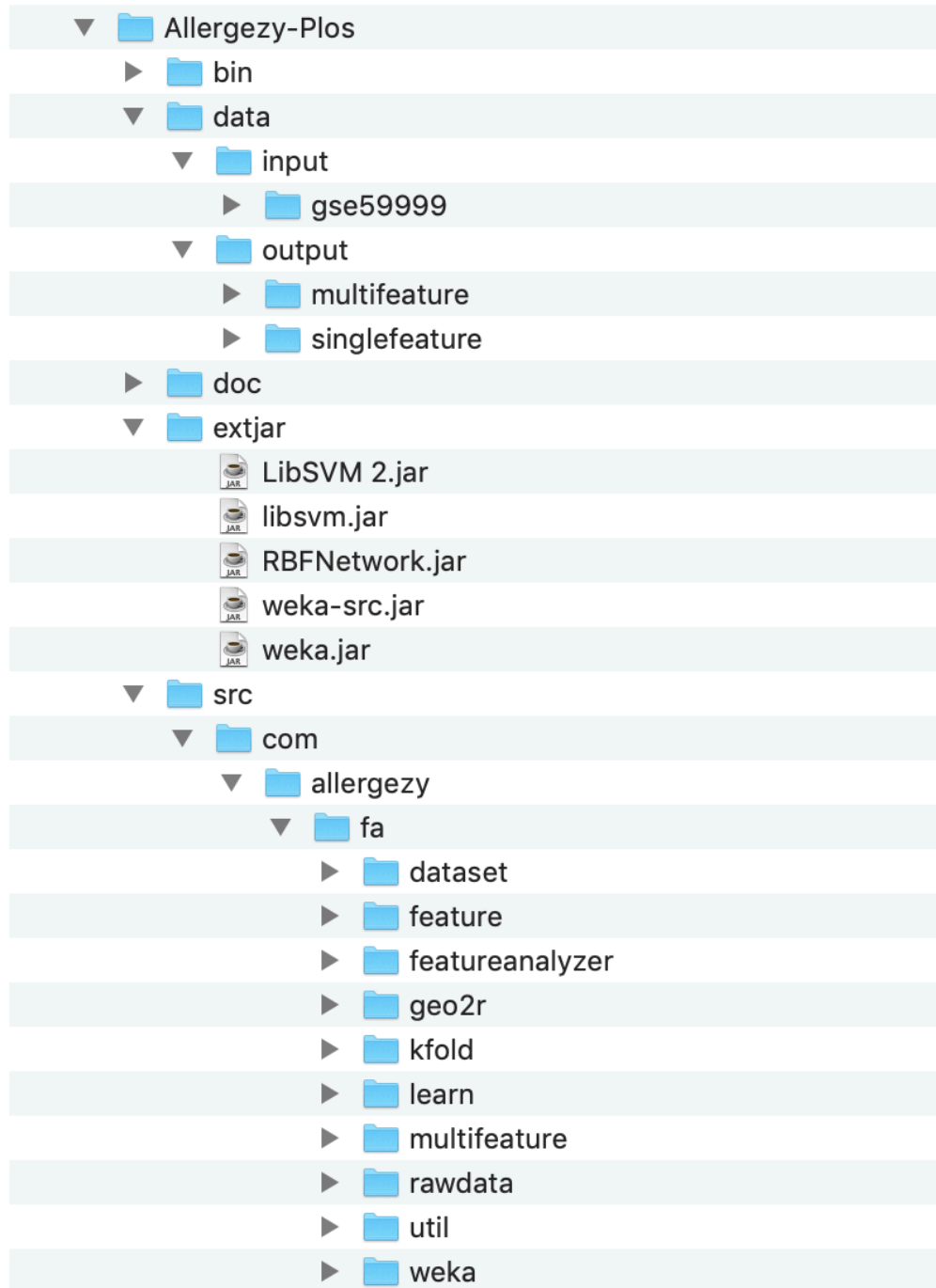
## Troubleshooting

## Before start

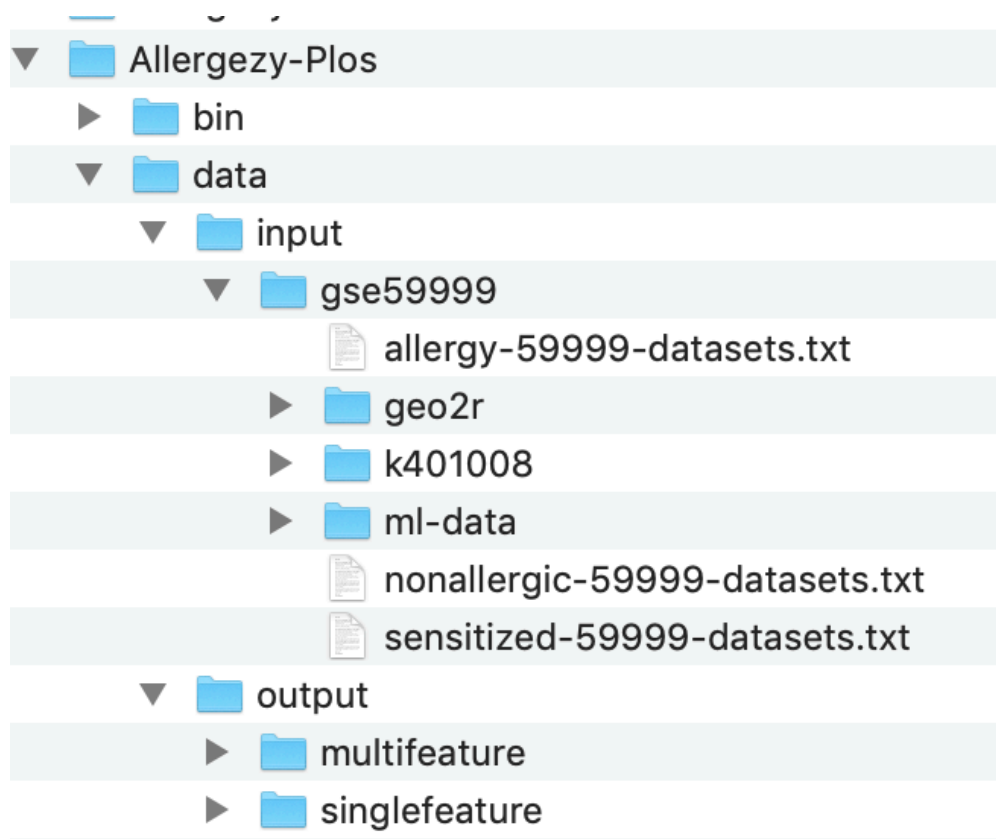
1. Download the Eclipse IDE (<https://www.eclipse.org/downloads/>)

- 1 Create an Eclipse project. Download weka jar files from <https://www.cs.waikato.ac.nz/ml/weka/downloading.html> . The Eclipse project should have the src files with the Java code downloaded in the previous step. Create a directory extjar and put the two reference jar files from Weka (weka-src.jar and weka.jar). Download RBFNetwork.jar from <https://jar-download.com/artifact-search/RBFNetwork> and add it to the extjar directory. Download libsvm.jar from <https://jar-download.com/download-handling.php>. Download the data.jar file and place it within the main project. This directory should have the input and output folders with many sub folders. The doc folder is optional and creates the generated JavaDoc for the project. JavaDoc can be generated by going to Eclipse → Project → Generate JavaDoc.





- 2 Next, we will validate that the CpG data that has already been separated into Food Allergy, Sensitized, and Non-Allergic samples have been downloaded correctly. Expand the data/input/gse59999 folder and verify that there are three files:
- allergy-59999-datasets.txt 215 MB contains 30 rows by 405,659 columns
  - nonallergic-59999-datasets.txt 98.9 MB contains 14 rows by 405,659 columns
  - sensitized-59999-datasets.txt 215 MB contains 30 rows by 405,659 columns



From within Eclipse, run the main program for `com.allergezy.fa.util.FileUtil`. You should see the output shown below.

Note the number of rows and columns and the validation check.

```
com.allergezy.fa.util.FileUtil:main:~/Allergezy-Plos
InMemoryFileReader::Total number of lines data//input/gse59999/allergy-59999-datasets.txt =30
Allergy file has .. 30 rows.
Number of columns = 405659
ID_REF cg00000029 .. cg27666123
InMemoryFileReader::Total number of lines data//input/gse59999/sensitized-59999-datasets.txt =30
Allergy file has .. 30 rows.
Number of columns = 405659
ID_REF cg00000029 .. cg27666123
InMemoryFileReader::Total number of lines data//input/gse59999/nonallergic-59999-datasets.txt =14
Allergy file has .. 14 rows.
Number of columns = 405659
ID_REF cg00000029 .. cg27666123
Allergy and Sensitized column match = true
Allergy and Non-allergic column match = true
```

The research was conducted using a dataset found in the Gene Expression Omnibus (GEO) under accession id GSE59999. The 71 patient samples in this dataset consisted of

29 patients with FA (tested positive on OFCs), 29 patients who were sensitized but not food-allergic, and 13 patients who were neither sensitized nor allergic. Each sample is associated with a DNAm profile that was taken from mononuclear blood cells and consisted of normalized Beta values ranging from 0 (completely unmethylated) to 1 (fully methylated) at 405,658 CpG islands distributed across the genome.

3 In this step, we will create 8-fold test/train/hidden samples.

The 58 samples are randomly split into three datasets: 40 samples for training, 10 samples for testing, and 8 completely hidden samples for cross-validation. Half of the samples in each of the three datasets were children with food allergies and the other half were those that were food-sensitized but not food-allergic. To avoid potential bias, we created eight random splits, shuffling the samples across the three datasets each time such that each of the 58 samples was in the hidden dataset at least once.

The logic is implemented in `com.allergezy.fa.kfold.KFoldDatasetGenerator`. Run the main method in this class to see an output similar to below.

```
InMemoryFileReader::Total number of lines data//input/gse59999/allergy-59999-datasets.txt =30
InMemoryFileReader::Total number of lines data//input/gse59999/sensitized-59999-datasets.txt =30
[14, 16, 28]
[32, 47, 57]

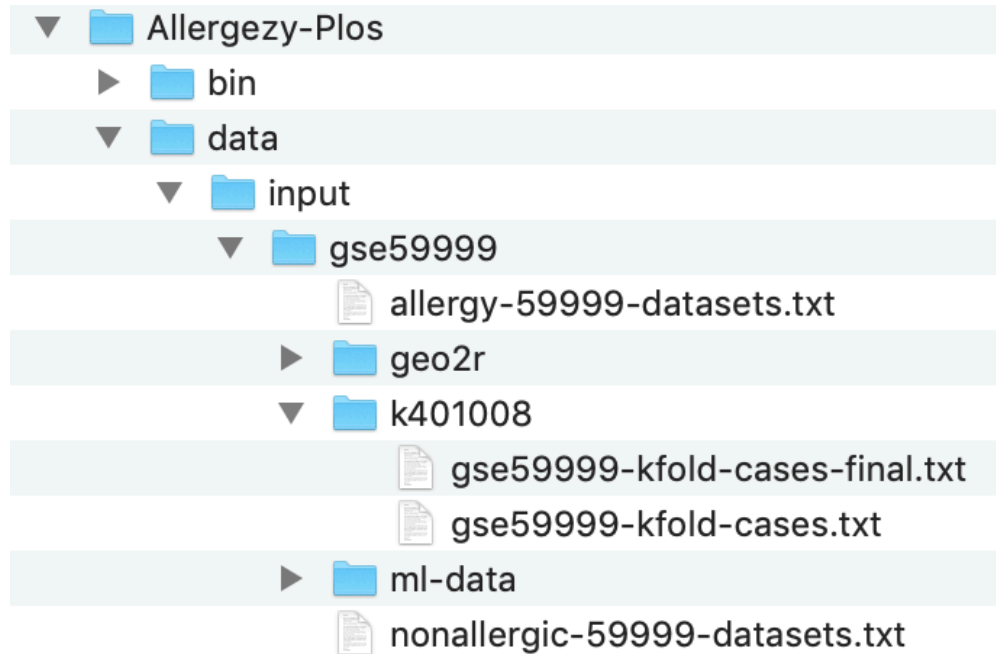
List:
GSM1463330,
GSM1463346,
GSM1463358,
GSM1463376,
GSM1463379,
GSM1463382,
GSM1463394,
GSM1463396,

List:
GSM1463336,
GSM1463341,
GSM1463362
```

Output from running `com.allergezy.fa.kfold.KFoldDatasetGenerator`.

This program produces an output file `data/input/gse59999/k401008/gse59999-kfold-cases.txt` as shown below.

The file `data/input/gse59999/k401008/gse59999-kfold-cases-final.txt` is the final version of this 8-fold split that is used in the rest of the study.



Below is the content of the data/input/gse59999/k401008/gse59999-kfold-cases-final.txt file.

```
Case 0
Training: 0 GSM1463328,GSM1463330,GSM1463342,GSM1463343,GSM1463344,GSM1463352,GSM1463354,GSM1463361,GSM1463362,
Test: 0 GSM1463332,GSM1463337,GSM1463380,GSM1463387,GSM1463389,GSM1463340,GSM1463351,GSM1463357,GSM1463370,GSM1
Hidden: 0 GSM1463334,GSM1463346,GSM1463347,GSM1463348,GSM1463350,GSM1463356,GSM1463358,GSM1463382,
Case 1
Training: 1 GSM1463330,GSM1463332,GSM1463334,GSM1463342,GSM1463344,GSM1463347,GSM1463350,GSM1463354,GSM1463356,
Test: 1 GSM1463328,GSM1463343,GSM1463363,GSM1463378,GSM1463398,GSM1463339,GSM1463367,GSM1463371,GSM1463382,GSM1
Hidden: 1 GSM1463331,GSM1463337,GSM1463338,GSM1463341,GSM1463349,GSM1463352,GSM1463379,GSM1463384,
Case 2
Training: 2 GSM1463328,GSM1463330,GSM1463332,GSM1463347,GSM1463350,GSM1463352,GSM1463354,GSM1463356,GSM1463361,
Test: 2 GSM1463334,GSM1463337,GSM1463372,GSM1463387,GSM1463394,GSM1463336,GSM1463345,GSM1463346,GSM1463368,GSM1
Hidden: 2 GSM1463342,GSM1463343,GSM1463344,GSM1463363,GSM1463371,GSM1463383,GSM1463388,GSM1463396,
Case 3
Training: 3 GSM1463328,GSM1463330,GSM1463334,GSM1463337,GSM1463342,GSM1463343,GSM1463344,GSM1463347,GSM1463350,
Test: 3 GSM1463332,GSM1463363,GSM1463376,GSM1463378,GSM1463380,GSM1463339,GSM1463358,GSM1463368,GSM1463382,GSM1
Hidden: 3 GSM1463354,GSM1463357,GSM1463361,GSM1463386,GSM1463387,GSM1463390,GSM1463392,GSM1463393,
```

Each sample appears at least once in the hidden set. In addition, for every case there are 40 training samples. There is an equal number of food-allergic and food-sensitized samples in each dataset and in each case.

To verify that this file can be accessed, run the main method for `com.allergezy.fa.kfold.KFoldDatasetInfo`. This file follows the Singleton design pattern

and provides information on which samples are in the training, test, and hidden datasets for each of the eight cases. You should see the following output.

```
InMemoryFileReader::Total number of lines data//input/gse59999/k401008/gse59999-kfold-cases-final.txt =33
Training: 0 [40]:GSM1463328, GSM1463330, GSM1463342, GSM1463343, GSM1463344, GSM1463352, GSM1463354, GSM1463361, GSM1463362, GSM14
Test: 0 [10]:GSM1463332, GSM1463337, GSM1463380, GSM1463387, GSM1463389, GSM1463340, GSM1463351, GSM1463357, GSM1463370, GSM1463383,
Hidden: 0 [8]:GSM1463334, GSM1463346, GSM1463347, GSM1463348, GSM1463350, GSM1463356, GSM1463358, GSM1463382,
Normal: 0 [0]:
Training: 1 [40]:GSM1463330, GSM1463332, GSM1463334, GSM1463342, GSM1463344, GSM1463347, GSM1463350, GSM1463354, GSM1463356, GSM14
Test: 1 [10]:GSM1463328, GSM1463343, GSM1463363, GSM1463378, GSM1463398, GSM1463399, GSM1463367, GSM1463371, GSM1463382, GSM1463386,
Hidden: 1 [8]:GSM1463331, GSM1463337, GSM1463338, GSM1463341, GSM1463349, GSM1463352, GSM1463379, GSM1463384,
Normal: 1 [0]:
Training: 2 [40]:GSM1463328, GSM1463330, GSM1463332, GSM1463347, GSM1463350, GSM1463352, GSM1463354, GSM1463356, GSM1463361, GSM14
Test: 2 [10]:GSM1463334, GSM1463337, GSM1463372, GSM1463387, GSM1463394, GSM1463336, GSM1463345, GSM1463346, GSM1463368, GSM1463386,
Hidden: 2 [8]:GSM1463342, GSM1463343, GSM1463344, GSM1463363, GSM1463371, GSM1463383, GSM1463388, GSM1463396,
Normal: 2 [0]:
Training: 3 [40]:GSM1463328, GSM1463330, GSM1463334, GSM1463337, GSM1463342, GSM1463343, GSM1463344, GSM1463347, GSM1463350, GSM14
```

The distribution of the samples can be seen in the following table.

**Table 1. Sample distribution across the eight independent cases**

Case	Test Cases (10)	Hidden Cases (8)
1	GSM1463332, GSM1463337, GSM1463380, GSM1463387, GSM1463389, GSM1463340, GSM1463351, GSM1463357, GSM1463370, GSM1463383	GSM1463334, GSM1463346, GSM1463347, GSM1463348, GSM1463350, GSM1463356, GSM1463358, GSM1463382
2	GSM1463328, GSM1463343, GSM1463363, GSM1463378, GSM1463398, GSM1463339, GSM1463367, GSM1463371, GSM1463382, GSM1463386	GSM1463331, GSM1463337, GSM1463338, GSM1463341, GSM1463349, GSM1463352, GSM1463379, GSM1463384
3	GSM1463334, GSM1463337, GSM1463372, GSM1463387, GSM1463394, GSM1463336, GSM1463345, GSM1463346, GSM1463368, GSM1463386	GSM1463342, GSM1463343, GSM1463344, GSM1463363, GSM1463371, GSM1463383, GSM1463388, GSM1463396
4	GSM1463332, GSM1463363, GSM1463376, GSM1463378, GSM1463380, GSM1463339, GSM1463358, GSM1463368, GSM1463382, GSM1463397	GSM1463354, GSM1463357, GSM1463361, GSM1463386, GSM1463387, GSM1463390, GSM1463392, GSM1463393
5	GSM1463330, GSM1463361, GSM1463384, GSM1463389, GSM1463398, GSM1463331, GSM1463338, GSM1463348, GSM1463351, GSM1463367	GSM1463328, GSM1463346, GSM1463349, GSM1463353, GSM1463365, GSM1463368, GSM1463372, GSM1463380
6	GSM1463328, GSM1463354, GSM1463363, GSM1463372, GSM1463389, GSM1463340, GSM1463341, GSM1463353, GSM1463377, GSM1463397	GSM1463330, GSM1463332, GSM1463344, GSM1463351, GSM1463370, GSM1463371, GSM1463378, GSM1463385
7	GSM1463328, GSM1463350, GSM1463356, GSM1463372, GSM1463380, GSM1463331, GSM1463338, GSM1463340, GSM1463351, GSM1463390	GSM1463339, GSM1463345, GSM1463352, GSM1463362, GSM1463389, GSM1463391, GSM1463397, GSM1463398
8	GSM1463337, GSM1463363, GSM1463380, GSM1463389, GSM1463398, GSM1463331, GSM1463349, GSM1463357, GSM1463388, GSM1463393	GSM1463336, GSM1463340, GSM1463367, GSM1463373, GSM1463376, GSM1463377, GSM1463384, GSM1463394

Each sample appears at least once in the hidden set. In addition, for every case there are 40 training samples that are not shown in the table. These 40 training samples are the remaining samples that are in neither the test nor the hidden cross-validation set for that case. There is an equal number of food-allergic and food-sensitized samples in each dataset and in each case.

- 4 DNAm datasets are characterized as having a small number of samples but a very high number of feature dimensions (HDLSS). To prevent overfitting and increase generalization, it is important to condense the feature list relative to the number of samples available. Computationally, it is very expensive to evaluate the more than 400K CpG features individually. Therefore, in order to limit the evaluation size and begin with a

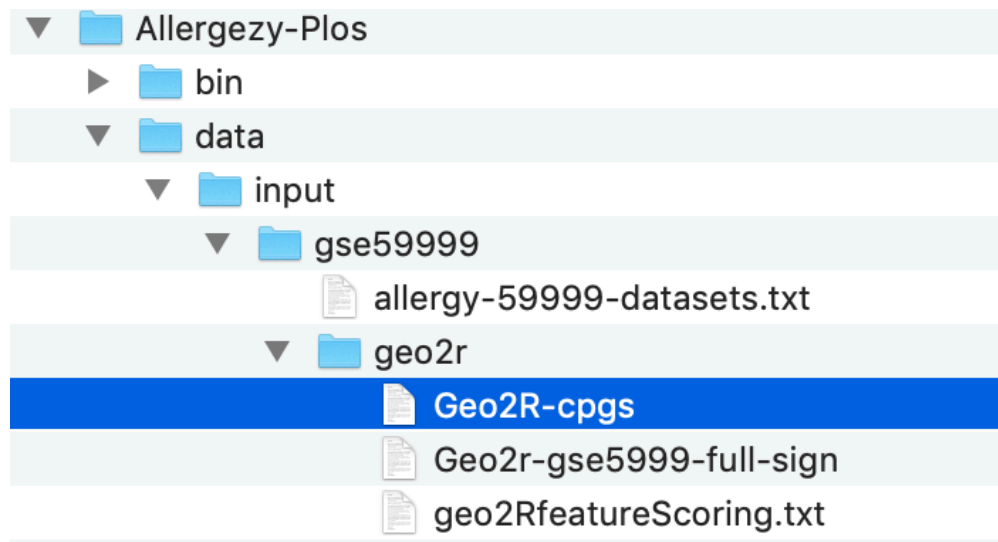
list of potentially highly-relevant CpG points, we used the NCBI **GEO2R tool** to obtain a prioritized list of CpG features differentially expressed across the two groups. For each of the eight independent cases, we split the forty training samples into two cohorts, one with allergic patients and the other with sensitized patients. The GEO2R tool was used to derive eight lists of 100 CpGs each, one for each of the eight cases.

The screenshot below shows the use of the GEO2R tool to create two cohorts of 20 samples each for the first case.

Clicking on

Group	Accession	Source name	Gender	Phenotype	Challenge outcome
Allergic	GSM1463328	gDNA from normal PBMC	FEMALE	Egg allergic	allergic
Allergic	GSM1463330	gDNA from normal PBMC	MALE	Peanut allergic	allergic
-	GSM1463332	gDNA from normal PBMC	FEMALE	Peanut allergic	allergic
-	GSM1463334	gDNA from normal PBMC	FEMALE	Egg allergic	allergic
-	GSM1463337	gDNA from normal PBMC	MALE	Peanut allergic	allergic
Allergic	GSM1463342	gDNA from normal PBMC	MALE	Peanut allergic	allergic
Allergic	GSM1463343	gDNA from normal PBMC	MALE	Egg allergic	allergic
Allergic	GSM1463344	gDNA from normal PBMC	MALE	Peanut allergic	allergic
-	GSM1463347	gDNA from normal PBMC	FEMALE	Egg allergic	allergic
-	GSM1463350	gDNA from normal PBMC	MALE	Egg allergic	allergic
Allergic	GSM1463352	gDNA from normal PBMC	FEMALE	Peanut allergic	allergic
Allergic	GSM1463354	gDNA from normal PBMC	MALE	Egg allergic	allergic
-	GSM1463356	gDNA from normal PBMC	FEMALE	Peanut allergic	allergic
Allergic	GSM1463361	gDNA from normal PBMC	MALE	Egg allergic	allergic
Allergic	GSM1463362	gDNA from normal PBMC	MALE	Egg allergic	allergic

The full signature for the eight cases is in data/input/gse59999/geo2r/Geo2R-cpgs.txt file.



Below are the first twenty lines of the file `data/input/gse59999/geo2r/Geo2R-cpgs.txt` file. This will be used in the next few steps.

Geo2R-cpgs.txt								
1	cg07060505	cg06410630	cg25866059	cg06410630	cg20463995	cg02681173	cg09755579	cg20502977
2	cg02681173	cg20502977	cg24851651	cg09861992	cg03068039	cg07033513	cg24616138	cg18569070
3	cg19714913	cg16978004	cg05861255	cg06243400	cg12050358	cg12050358	cg06410630	cg24851651
4	cg18884295	cg13455434	cg06201372	cg00936790	cg03970350	cg19900821	cg20502977	cg01806508
5	cg15188491	cg24616138	cg01232668	cg04015759	cg04475375	cg04543115	cg13560030	cg09618933
6	cg06410630	cg03356595	cg26401541	cg00076774	cg26124569	cg25890092	cg03946731	cg14552508
7	cg25199372	cg10301401	cg11555257	cg20987610	cg11941920	cg24748191	cg21936550	cg05897163
8	cg09602803	cg17460909	cg09755579	cg27143570	cg07965110	cg26124569	cg00314240	cg08929467
9	cg14789214	cg05406635	cg13632752	cg04958411	cg08378782	cg14014506	cg05223507	cg05791946
10	cg12562479	cg00109551	cg07573872	cg01872024	cg07649835	cg07502333	cg00939931	cg22452122
11	cg09755579	cg27615388	cg00968893	cg07060505	cg19585586	cg15082028	cg18798744	cg10301401
12	cg11390957	cg01432552	cg09618933	cg00442802	cg03716942	cg01893629	cg27517563	cg21113746
13	cg21615831	cg13560030	cg09939673	cg20445094	cg24616138	cg20795023	cg25087507	cg06410630
14	cg23556923	cg26124569	cg11571263	cg09679227	cg04334011	cg07235355	cg24079727	cg25921609
15	cg23216101	cg06447354	cg02313172	cg05008688	cg04623023	cg13632752	cg10461264	cg21610436
16	cg26059153	cg10079327	cg05261496	cg18971416	cg08860346	cg00999904	cg02681173	cg02313172
17	cg21610436	cg05435286	cg09722609	cg04746699	cg06038701	cg11740068	cg12176709	cg27560781
18	cg03274391	cg02124291	cg16434331	cg15250507	cg24584002	cg01962758	cg10301401	cg10336671
19	cg13169968	cg24187345	cg26281728	cg09755579	cg19539826	cg14760797	cg01441698	cg16949103
20	cg24584002	cg27270590	cg13560030	cg04226232	cg18798744	cg08643692	cg17032565	cg04897044

- 5 Next, the eight lists were combined together, which resulted in 636 unique CpGs, the count being less than 800 as some of the CpGs were repeated across the eight cases.

To map the CpGs to genes, we use the output from GEO2R contained in the file `data/input/gse59999/geo2r/Geo2r-gse5999-full-sign.txt`. The Singleton class `com.allergezy.fa.geo2r.CPGAnnotations` maps a CpG to genes using this file. Run the main method in the class `com.allergezy.fa.geo2r.CPGAnnotations`. You should see the output similar to the screen shot below.



```
300000 "cg13581475" "9.66e-01" "6.27e-01" "4.88e-01" "-5.7782727" "2.40e-02" "63924286"
InMemoryFileReader::Total number of lines data//input/gse59999/geo2r/Geo2r-gse5999-full-sign.txt =462177
cg06410630 -> RNF213;LOC100294362
cg06669701 -> FAM190B
cg06628000 -> SARS
cg10461264 ->
cg18988685 ->
cg24616138 -> CTBP2;CTBP2
cg27027230 -> ARID5B
cg00936790 -> KIF13B
cg14414100 -> SLC24A2
cg00939931 -> MAFK
cg06116095 -> PANX1
cg02788266 ->
cg03068039 -> ZNF252;TMED10P
cg25890092 -> CD7
cg19287711 ->
cg07033513 ->
cg07060505 ->
cg26963090 -> TIMP2
cg06410630 -> RNF213;LOC100294362
```

The class `com.allergezy.fa.geo2r.Geo2rCpgInfo` computes the overlap between the CpGs from the GEO2R output. Running the main method produces the output shown below.

```
InMemoryFileReader::Total number of lines data//input/gse59999/geo2r/Geo2R-cpgs.txt =99
300000 "cg13581475" "9.66e-01" "6.27e-01" "4.88e-01" "-5.7782727" "2.40e-02" "63924286" "MTHFD1"
InMemoryFileReader::Total number of lines data//input/gse59999/geo2r/Geo2r-gse5999-full-sign.txt =462177
Case: 99 .... 644
cg06410630 [571-5,0,0,20,90,2,12,] -- {RNF213;LOC100294362}, cg13560030 [442-59,12,19,37,4,27,] -- {NTN4}, cg02681173 [395-1,34,
```

The file `data/input/gse59999/geo2r/geo2RfeatureScoring.txt` contains the result of this analysis.





geo2RfeatureScoring.txt				
1	Id	CPG	Gene	Score Position
2	1	cg06410630	RNF213;LOC100294362	571 5,0,0,20,90,2,12,
3	2	cg13560030	NTN4	442 59,12,19,37,4,27,
4	3	cg02681173	LOC100190940	395 1,34,0,15,55,
5	4	cg09755579	SNORA70B;USP34	395 10,7,18,70,0,
6	5	cg20502977	COL6A3;COL6A3;COL6A3	357 1,39,3,0,
7	6	cg26124569	LPP;LPP;LPP	333 13,5,7,42,
8	7	cg24616138	CTBP2;CTBP2	313 4,12,1,70,
9	8	cg24584002	RNASEH1	294 19,17,39,31,
10	9	cg03946731	PKMYT1;PKMYT1	291 49,22,33,5,
11	10	cg20463995		281 38,43,0,38,
12	11	cg09618933		279 47,11,59,4,
13	12	cg10301401	LMF1	267 6,17,10,
14	13	cg08378782	RASGRP2;RASGRP2;RASGRP2	229 8,26,37,
15	14	cg21615831	KSR1	224 12,58,33,73,
16	15	cg07060505		221 0,10,69,
17	16	cg12176709	DRD2;DRD2	218 35,73,58,16,
18	17	cg02866639		212 27,34,27,
19	18	cg05897163	null	203 25,66,6,

This is summarized as shown below

**Table 2. Top CpGs and associated genes from GEO2R across 8 independent cases**

Number	CpG	Gene	Position
1	cg06410630	RNF213;LOC100294362	6,1,1,21,91,3,13
2	cg13560030	NTN4	60,13,20,38,5,28
3	cg02681173	LOC100190940	2,35,1,16,56
4	cg09755579	SNORA70B;USP34	11,8,19,71,1
5	cg20502977	COL6A3	2,40,4,1
6	cg26124569	LPP	14,6,8,43
7	cg24616138	CTBP2	5,13,2,71
8	cg24584002	RNASEH1	20,18,40,32
9	cg03946731	PKMYT1	50,23,34,6
10	cg20463995	-	39,44,1,39,
11	cg09618933	-	48,12,60,5
12	cg10301401	LMF1	7,18,11
13	cg08378782	RASGRP2	9,27,38
14	cg21615831	KSR1	13,59,34,74
15	cg07060505	-	1,11,70

Note that a CpG may not appear in the top 100 CpGs position across the eight cases. This list is based on the frequency of each CpG across the eight lists as well as its ranking in each list. The order of the genes in this table has no significance in our data-driven methodology but provides insight into how many of these CpGs were eventually selected later through our data-driven methodology.

Note that this ranking really has no material significance in our methodology, since we evaluated all the unique 636 CpGs independently. It does provide some insights into features highlighted by GEO2R and those that appeared in our final CpG signature. The 20% overlap we found between the CpGs across the eight lists seems to suggest that the examples do have an effect on the CpGs that are differentially expressed between the two cohorts (when GEO2R is used). Thus, averaging the results across the eight independent runs, which is performed throughout the rest of the paper, should help in avoiding any potential biases, due to the distribution of the samples across the training, test, and hidden datasets.

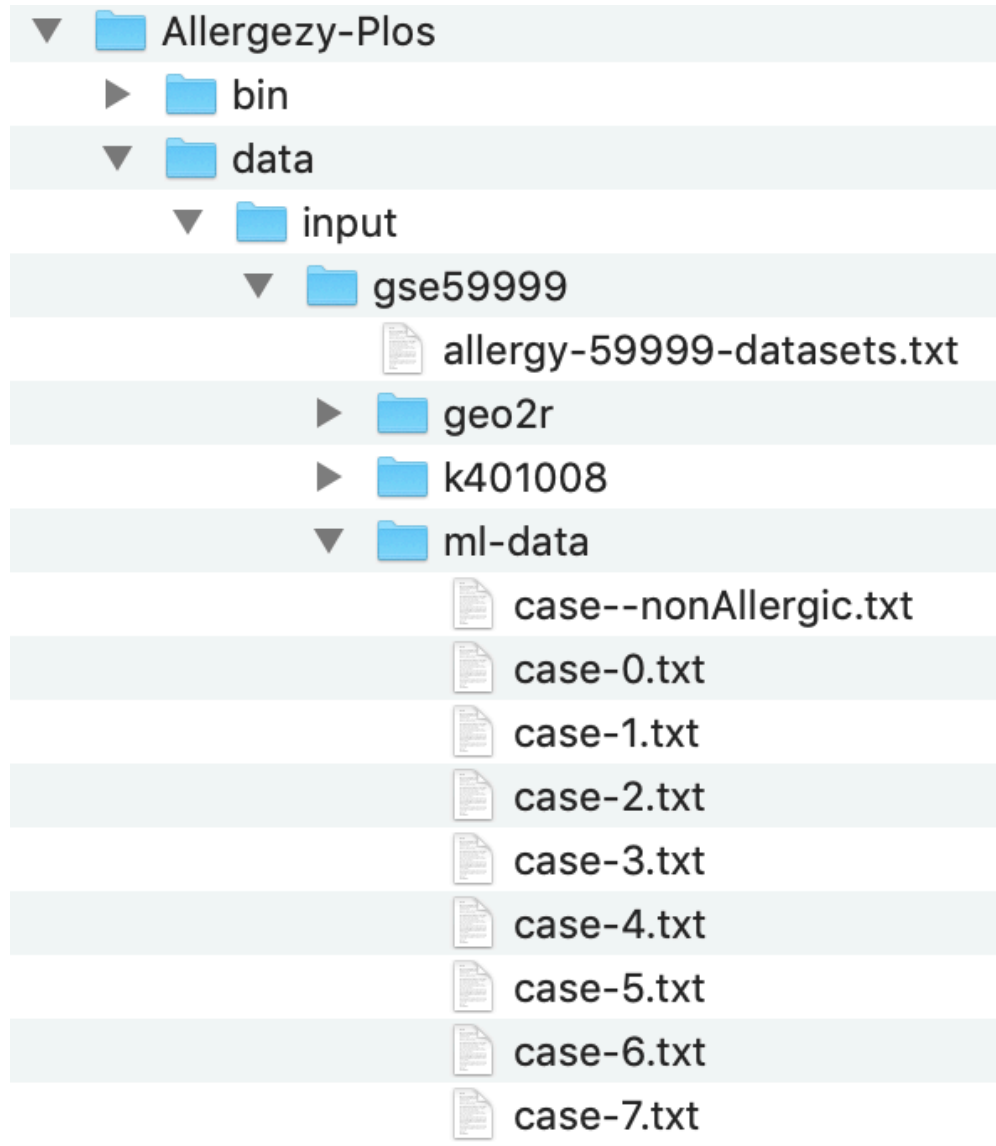
- 6 Next, we will create smaller dataset files that contain the machine learning examples with 636 CpG values as columns.

`com.allergezy.fa.dataset.DatasetCreatorForEachCase --` generates the file that has the examples for each case. The output from the program is shown below.

```

InMemoryFileReader::Total number of lines data//input/gse59999/allergy-59999-datasets.txt =30
InMemoryFileReader::Total number of lines data//input/gse59999/sensitized-59999-datasets.txt =30
InMemoryFileReader::Total number of lines data//input/gse59999/geo2r/Geo2R-cpgs.txt =99
InMemoryFileReader::Total number of lines data//input/gse59999/nonallergic-59999-datasets.txt =14

```



In the input/gse59999/ml-data there are eight files case-0.txt, case-1.txt, and so on for the eight folds.

Attached is a screen shot of a part of the first file case-0.txt.

```
1 ID_REF cg07060505 cg02681173 cg19714913 cg18884295 cg15188491 cg06410630 cg25199372 cg09
* cg02124291 cg04852972 cg00939931 cg14334310 cg00657460 cg24159214 cg09618933 cg09303977
* cg27217474 cg08235883 cg24126361 cg18512963 cg21926875 cg25623524 cg11815480 cg07740525
2 GSM1463328 0.7624729678158 0.352452345606308 0.380357937244544 0.660437367480148 0.175208584450
* 0.217692379904955 0.365743701945769 0.809898102537656 0.276434512610983 0.541775211401801 0.8763
* 0.605049295996472 0.174111396820066 0.278758468675512 0.105603368556034 0.85658240778978 0.8253
* 0.838010124408319 0.802279386019525 0.219379507331627 0.821232999938156 0.732976047763439 0.1861
3 GSM1463330 0.655330467618211 0.349160044261141 0.412464325240356 0.701010793215693 0.2097471035
* 0.173878109200632 0.291083774994774 0.801452705390324 0.286954650319599 0.47530676514792 0.8726
* 0.495186556536599 0.182677712624027 0.274713131364304 0.199966848885264 0.720228392858865 0.8134
* 0.860680856660602 0.830745542618494 0.20813800745818 0.82445583128368 0.697866988372391 0.1577
4 GSM1463332 0.77533863203392 0.275559429158274 0.472355348682406 0.737922445999914 0.2381840839
* 0.221526292697006 0.300411675776253 0.771478779411023 0.288110763677039 0.508922821356991 0.8465
* 0.451175721070312 0.164355961773016 0.329458434490091 0.317466812147663 0.814557363588863 0.7698
* 0.870146925200158 0.832713669520035 0.296001854405043 0.917857480983444 0.736933842419769 0.1511
```

- 7 For each case, and for each of the 636 unique CpGs, we built four different machine learning models: Decision Trees (DT), Logistic Regression (LR), Radial Basis Function (RBF), and a Multi-Layer Perceptron (MLP), a deep learning network with two hidden layers of ten nodes each. Each predictive model only used one CpG and was built on the training data. For each case and each feature, the classifier with the highest testing accuracy was selected. Finally, the average hidden data (cross-validation set) accuracy across the eight independent cases was computed. 636 of these accuracy scores (one for each CpG) and 20,352 models (8 independent cases x 636 features x 4 classifiers) were created in total using this process, for the single input case.

com.allergezy.fa.feature.SingleFeatureScoringRunner is the class that runs this computation. A few class to note

- com.allergezy.fa.weka.WekaHelper interfaces with the Weka APIs for creating models
- Logic for writing details as part of the learning are SummaryRunResultWriterImpl -- summary of results

DetailedRunResultWriterImpl -- detailed output of each model,  
BestModelForCasesResultWriterImpl - best model.

Run the main method com.allergezy.fa.feature.SingleFeatureScoringRunner. You should see an output similar to



```
<terminated> SingleFeatureScoringRunner (2) [Java Application] /Library/Java/JavaVirtualMachines/jdk1.8.0_101.jdk/Contents/Home/bin/java (Dec 9, 2018, 7:46:4
InMemoryFileReader::Total number of lines data//output/singlefeature/kfoldresult/scores.txt =41745
Writing to file ... data//output/singlefeature/kfoldresult/scores.txt
InMemoryFileReader::Total number of lines data//input/gse59999/k401008/gse59999-kfold-cases-final.txt =33
NekaHelper: class com.allergezy.fa.feature.FeatureScoringWekaHelper
NekaDatasetBuilder: class com.allergezy.fa.weka.WekaDatasetBuilder
InMemoryFileReader::Total number of lines data//input/gse59999/ml-data/case-0.txt =59
Completed 1 run out of 352
Completed 2 run out of 352
digraph J48Tree {
V0 [label="cg07060505" ]
V0->N1 [label="<= 0.338886"]
V1 [label="0 (28.0/8.0)" shape=box style=filled ]
V0->N2 [label="> 0.338886"]
V2 [label="1 (12.0)" shape=box style=filled ]
}
```

This program writes output to data/output/singlefeature/kfoldresult. Subdirectories case0, case1, ..., case7 has results for the eight folds. Within each directory are four sub directories: DECISION\_TREE, LOGISTIC\_REGRESSION, MLP, RBF. Within each directory are detailed results for each model and results -- featscore0.txt, featscore1.txt, .....,



▼	folder	Allergezy-Plos
▶	folder	bin
▼	folder	data
▶	folder	input
▼	folder	output
▶	folder	multifeature
▼	folder	singlefeature
▼	folder	kfoldresult
	file	allFeatureScores.txt
	file	Book1.xlsx
▼	folder	case0
▶	folder	DECISION_TREE
▶	folder	LOGISTIC_REGRESSION
▶	folder	MLP
▶	folder	RBF
▶	folder	case1
▶	folder	case2
▶	folder	case3
▶	folder	case4
▶	folder	case5
▶	folder	case6
▼	folder	case7
▼	folder	DECISION_TREE
	file	featscore1.txt
	file	featscore2.txt
	file	featscore3.txt
	file	featscore4.txt

The class `com.allergezy.fa.feature.BestFeatureIdentifierUsingHiddenTestScores` computes the average score for each feature across the eight cases.

The following is a screen shot of the output file `featureScoring.txt`

featureScoring.txt											
Id	Cpg	Algorithm	Epoch	numDimensions	trainCorrect	trainIncorrect	trainROC	testCorrect	testIncorrect	testROC	hiddenCorrect
1	cg21509221	MLP	0	1 50	50	0.5425	50	0.0	50	0.0125	
2	cg21509221	LOGISTIC_REGRESSION	0	1 50	50	0.5	50	0.5	50	0.5	
3	cg21509221	DECISION_TREE	0	1 50	50	0.5	50	0.5	50	0.5	
4	cg21509221	RBF	0	1 57.5	42.5	0.615	60	40	0.52	62.5	37.5
5	cg14086013	MLP	0	2 72.5	27.5	0.8	50	50	0.64	50	0.5
6	cg14086013	LOGISTIC_REGRESSION	0	2 72.5	27.5	0.8	50	50	0.64	62.5	37.5
7	cg14086013	DECISION_TREE	0	2 75	25	0.75	50	50	0.5	50	0.5
8	cg14086013	RBF	0	2 72.5	27.5	0.8175	40	60	0.6	50	0.5625
9	cg07133741	MLP	0	3 80	20	0.8925	30	70	0.52	87.5	12.5
10	cg07133741	LOGISTIC_REGRESSION	0	3 77.5	22.5	0.7775	30	70	0.28	87.5	12.5
11	cg07133741	DECISION_TREE	0	3 85	15	0.8625	30	70	0.3	87.5	12.5
12	cg07133741	RBF	0	3 77.5	22.5	0.855	30	70	0.52	87.5	12.5
13	cg10336671	MLP	0	4 85	15	0.8875	60	40	0.64	87.5	12.5

`com.allergezy.fa.feature.BestFeatureIdentifierUsingHiddenTestScores`

Output from the console

```
Reading file ..data//output/singlefeature/kfoldresult/scores.txt
InMemoryFileReader::Total number of lines data//output/singlefeature/kfoldresult/scores.txt =41753
Writing epoch cpgs ... 5114 data//output/singlefeature/kfoldresult/bestFeatureScoresCpgEpoch.txt
Writing ... 641 data//output/singlefeature/kfoldresult/bestFeatureScoresCpg.txt
```

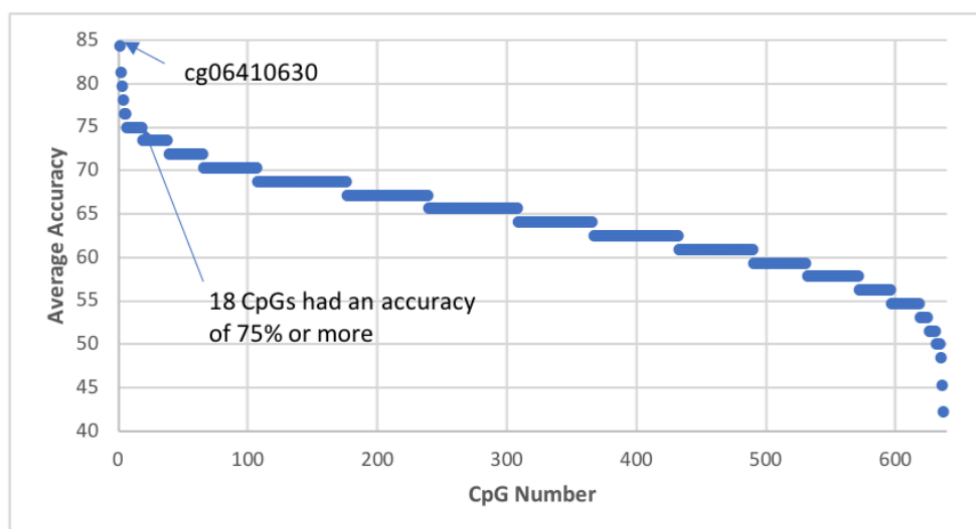
Output file `data/output/singlefeature/kfoldresult.bestFeatureScoresCpg.txt`

```
1 431 RBF 2 13772 89.0625 10.9375 0.9146875 83.75 16.25 0.8075 84.375 15.625 0.8359375
2 87 LOGISTIC_REGRESSION 1 2758 80.0 20.0 0.7734375 83.75 16.25 0.825 81.25 18.75 0.7890625
3 4 LOGISTIC_REGRESSION 6 122 76.875 23.125 0.80625 80.0 20.0 0.75 79.6875 20.3125 0.8359375
4 32 LOGISTIC_REGRESSION 1 998 79.6875 20.3125 0.8178124999999999 76.25 23.75 0.76 78.125 21.875 0.7421875
5 22 LOGISTIC_REGRESSION 5 694 79.0625 20.9375 0.8071875000000001 77.5 22.5 0.79 76.5625 23.4375 0.8125
6 379 MLP 3 12109 76.875 23.125 0.80625 63.75 36.25 0.59 76.5625 23.4375 0.7109375
7 269 LOGISTIC_REGRESSION 5 8598 76.875 23.125 0.7878125 80.0 20.0 0.7899999999999999 75.0 25.0 0.7734375
8 561 LOGISTIC_REGRESSION 2 17930 83.4375 16.5625 0.8505468750000001 77.5 22.5 0.8049999999999999 75.0 25.0 0.828125
9 541 LOGISTIC_REGRESSION 4 17298 74.0625 25.9375 0.7949999999999999 77.5 22.5 0.795 75.0 25.0 0.7421875
10 165 MLP 6 5305 75.625 24.375 0.765625 76.25 23.75 0.7349999999999999 75.0 25.0 0.734375
```

Attached is the screenshot of the final file, sorted by average hidden test accuracy.

7196	cg07193234	RBF	2	431	89.0625	10.9375	0.9146875	83.75	16.25	0.8075	84.375	15.625	0.8359375
3082	cg24854095	LOGISTIC_RE	1	87	80	20	0.7734375	83.75	16.25	0.825	81.25	18.75	0.7890625
16430	cg10336671	LOGISTIC_RE	6	4	76.875	23.125	0.80625	80	20	0.75	79.6875	20.3125	0.8359375
2862	cg17825100	LOGISTIC_RE	1	32	79.6875	20.3125	0.8178125	76.25	23.75	0.76	78.125	21.875	0.7421875
13766	cg04958411	LOGISTIC_RE	5	22	79.0625	20.9375	0.8071875	77.5	22.5	0.79	76.5625	23.4375	0.8125
9721	cg27217474	MLP	3	379	76.875	23.125	0.80625	63.75	36.25	0.59	76.5625	23.4375	0.7109375
14754	cg18512963	LOGISTIC_RE	5	269	76.875	23.125	0.7878125	80	20	0.79	75	25	0.7734375
7714	cg10205430	LOGISTIC_RE	2	561	83.4375	16.5625	0.85054688	77.5	22.5	0.805	75	25	0.828125
13106	cg01962758	LOGISTIC_RE	4	541	74.0625	25.9375	0.795	77.5	22.5	0.795	75	25	0.7421875
17077	cg06868247	MLP	6	166	75.625	24.375	0.7465625	76.25	23.75	0.735	75	25	0.7734375
4413	cg04774040	MLP	1	420	76.5625	23.4375	0.8215625	75	25	0.72	75	25	0.7421875
11857	cg02681173	MLP	4	229	73.75	26.25	0.81625	75	25	0.8	75	25	0.8203125
4146	cg04475375	LOGISTIC_RE	1	353	72.8125	27.1875	0.7771875	75	25	0.77	75	25	0.7734375
17793	cg14336003	MLP	6	345	72.8125	27.1875	0.8240625	72.5	27.5	0.785	75	25	0.8125
9362	cg09095047	LOGISTIC_RE	3	289	69.375	30.625	0.763125	67.5	32.5	0.76	75	25	0.78125
4706	cg05359853	LOGISTIC_RE	1	493	74.6875	25.3125	0.771875	66.25	33.75	0.595	75	25	0.75
7121	cg12596505	MLP	2	413	77.1875	22.8125	0.7965625	66.25	33.75	0.5925	75	25	0.796875
20497	cg12285988	MLP	7	337	79.0625	20.9375	0.81390625	63.75	36.25	0.6	75	25	0.765625
16974	cg02382878	LOGISTIC_RE	6	140	70	30	0.748125	85	15	0.82	73.4375	26.5625	0.7734375
13314	cg01079652	LOGISTIC_RE	4	593	75.3125	24.6875	0.8103125	82.5	17.5	0.8125	73.4375	26.5625	0.8359375

The following graph summarizes the results.



**Figure 1. Average accuracy across eight independent cases for singular CpG features.** Each CpG was used as an input to create classifiers using four different machine learning methods. The classifier with the best score on test data was then selected and evaluated on the hidden dataset. The above accuracy for each CpG is the average accuracy across the eight different cases. cg06410630 was the top CpG with an average accuracy of 84.375%. 18 CpGs had a score of 75% or more.

The top CpGs with associated genes are



**Table 3. Top CpGs and associated genes using single inputs to a classifier across 8 independent cases**

Number	CpG	Gene	Average Accuracy	AUROC
1	cg06410630	RNF213;LOC100294362	84.375	0.8359375
2	cg06669701	FAM190B	81.25	0.7890625
3	cg06628000	SARS	79.6875	0.8359375
4	cg10461264	-	78.125	0.7421875
5	cg18988685	-	76.5625	0.8125
6	cg24616138	CTBP2	76.5625	0.7109375
7	cg27027230	ARID5B	75	0.765625
8	cg00936790	KIF13B	75	0.7421875
9	cg14414100	SLC24A2	75	0.7734375
10	cg00939931	MAFK	75	0.796875
11	cg06116095	PANX1	75	0.7421875
12	cg02788266	-	75	0.7734375
13	cg03068039	ZNF252;TMED10P	75	0.828125
14	cg25890092	CD7	75	0.8203125
15	cg19287711	-	75	0.78125
16	cg07033513	-	75	0.75
17	cg07060505	-	75	0.8125
18	cg26963090	TIMP2	75	0.7734375

These 18 CpGs achieved an accuracy score of 75% or higher when used as the singular input feature in the machine learning models. cg06410630 with an accuracy of 84.375% was the top CpG from the 636 CpGs that were used to create single input classifiers. The scores were averaged over 8 independent cases. For each case, the machine learning models were retrained and accuracy was computed on completely hidden data.

- 8 We sought to increase the number of features used by the classifier while still avoiding over-fitting and encouraging generalization. Therefore, we selected the first eighteen CpGs, based on the accuracy scores computed above, and combined them two at a time, followed by three at a time, and so on until combinations of twelve were reached. Given the large number of potential combinations, each model was limited to a small subset of strong CpG-lists, to which a new input feature was added. On an average, we tried about 200 unique combinations for a given number of input features. Again, each unique input feature combination set was run  $4 \times 8 = 32$  times, to account for the four different classifier methods and 8 independent sample-distribution "cases".

The class `com.allergezy.fa.multifeature.casegenerator.MultiFeatureCasesGenerator` is used to create next set of cases to be tried.

- 9 For a given number of input features, the models were ranked using their average accuracy scores on hidden data cohorts (described above). Odd numbers of models,

starting from 1 to 101, were combined together using a simple voting scheme, i.e., each model independently predicted whether a sample was classified as FA or sensitized and the final prediction was the majority of predictions made across the different models.

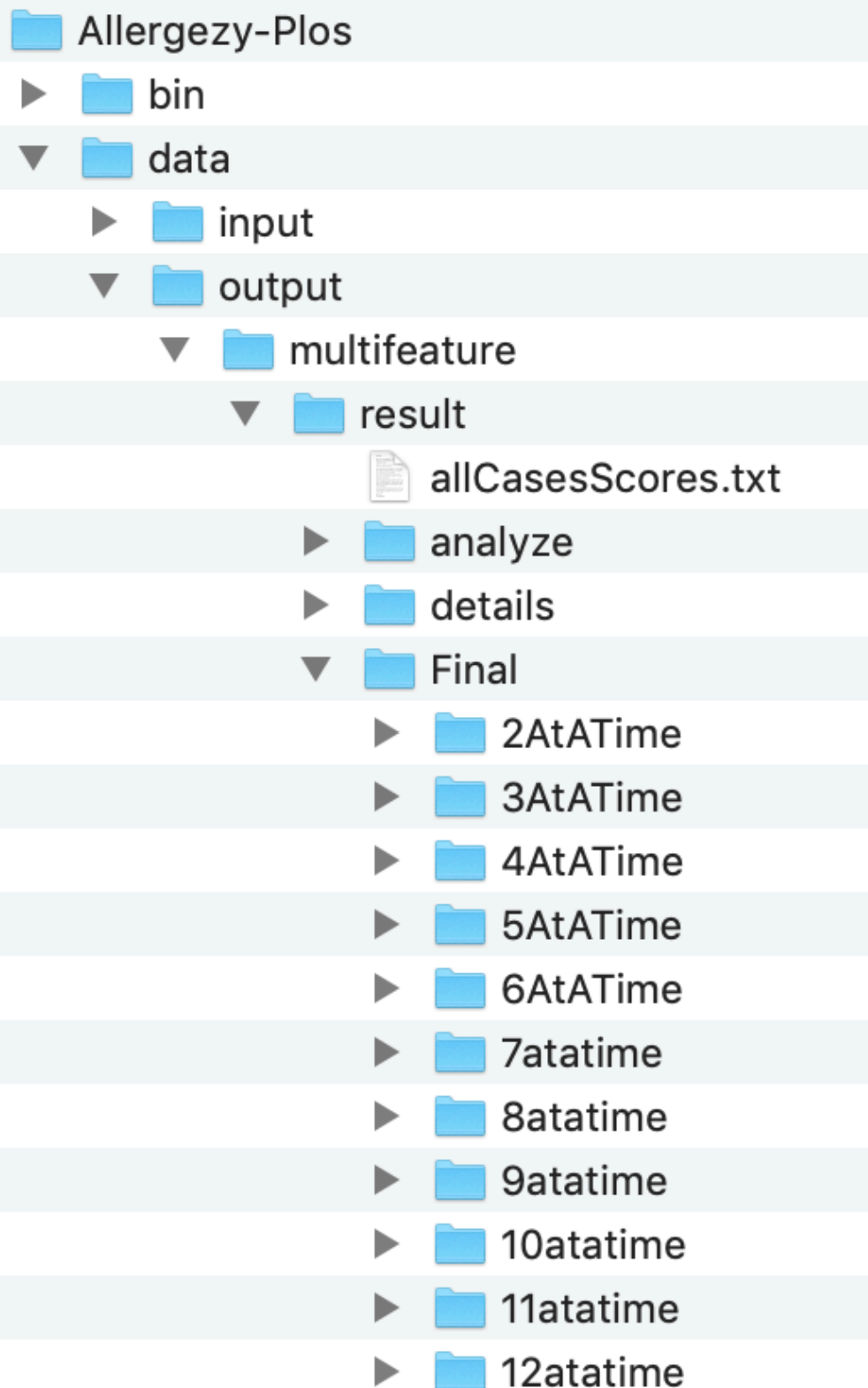
The class `com.allergezy.fa.kfold.verify.VerifyWekaDatasetRunner` runs the set of cases specified in the `data/output/multifeature/setup/summary.txt` file. A screen shot of an example file is shown next.

```
summary.txt
2000000063 MLP 2 1993 100 0 1 96.25 3.75 0.985 100 0 1 cg06410630 cg10461264 cg06116095 cg06628000 cg26963090 cg18988685 cg02788266 cg0306803
2000000084 MLP 2 2665 100 0 1 95 5 0.995 100 0 1 cg06410630 cg10461264 cg06116095 cg06628000 cg26963090 cg18988685 cg02788266 cg03068039 cg1
```

This class has a visitor `BestModelForCasesResultWriterImpl` configured to calculate majority prediction.

```
private List<ResultWriter> getVisitors() {
    List<ResultWriter> resultWriters = new ArrayList<ResultWriter>();
    // Detail of each run
    resultWriters.add(new DetailedRunResultWriterImpl(GSE59999_Verify_DetailCombination_Result, "case"));
    // Best model for each case -- summary majority prediction by voting
    resultWriters.add(new BestModelForCasesResultWriterImpl(GSE59999_Verify_BestAnalyzerOut_CaseResult));
    // Summary for each case
    resultWriters.add(new SummaryRunResultWriterImpl(GSE59999_Verify_FeatureCombination_Result));
}
```

Iterate different combinations of CpG features using the `summary.txt` file as input, run the results, select the best models, and try new combinations adding one feature at a time. The results for various combinations are in the directory `data/output/multifeature/result/Final`.



- 10 The file `com.allergezy.fa.rawdata.RawCpGDataExtractor` can extract raw CpG values for a CpG across the samples and can be used for generating data plots. Here is an example file.

```
rawCpgData.txt
1 1 cg03068039 cg10461264
2 1 0.861645527 0.317720181
3 1 0.837680237 0.216547093
4 1 0.797510971 0.2253189
5 1 0.793506157 0.237929743
6 1 0.839851223 0.184351872
7 1 0.794760701 0.233254063
8 1 0.818285816 0.244467575
9 1 0.788111136 0.247797637
10 1 0.797850347 0.219340641
11 1 0.814959698 0.28031957
```

`com.allergezy.fa.featureanalyzer.CpGFrequencyFeatureAnalyzer`  
Analyzes the frequency of various CpG across the summary file

- 11 Gene set analysis can provide biological context, as well as insights into disease mechanisms and possible treatments. Biological enrichment was performed by applying Illumina's BaseSpace Correlation Engine to the 13-gene list. To understand these genes better we identified tissues where the genes were expressed, found associated

biological pathways, used gene ontology concepts to identify functionally-related gene sets, connected the 13-gene list to the Broad positional gene sets, and connected the gene signature to protein families.


The 13-gene signature was imported into Illumina BaseSpace Correlation Engine

13 features Symbol	EntrezGene ID	Imported ID
RNF213	57674	RNF213
ZNF252P	286101	ZNF252
TMED10P1	286102	TMED10P
SARS	6301	SARS
TIMP2	7077	TIMP2
MAFK	7975	MAFK
CD7	924	CD7
PANX1	24145	PANX1
CTBP2	1488	CTBP2
SLC24A2	25769	SLC24A2
ARID5B	84159	ARID5B
KIF13B	23303	KIF13B
FAM190B	54462	FAM190B

QuickView for Bioset:

 **allergezy-2018**

Bioset from study: 18 CpG 13 gene signature

 Homo sapiens | **RE** RNA Expression | 13 features (mapped to 13 genes)

[View Bioset Details](#)

[Edit](#) [Delete](#)

[NEXTBIO SUMMARY](#)

[BIOSET DATA](#)

Search by genes or keywords



[show all](#)

[Data Filter](#) **NEW**

[Pathway Viewer](#) **NEW**


[Pathway Studio](#) **NEW**

13 features Symbol	EntrezGene ID	Imported ID
RNF213	57674	RNF213
ZNF252P	286101	ZNF252
TMED10P1	286102	TMED10P

## Body Atlas for Bioset:

### **allergezy-2018**

Bioset from study: [18 CpG 13 gene signature](#)












 Homo sapiens | **RE** RNA Expression | 13 features (mapped to 13 genes)

[View Bioset Details](#)

▼ **TISSUES**
▶ CELL TYPES
▶ CELL LINES
▶ STEM CELLS

View by: Body Systems ▼
Organism: Homo sapiens ▼
Source: Array based ▼
Scale:

 Body System locator

Name	Score	Correlation Over
<div>  Urogenital System </div>	<div> <div></div> <div>0246810</div> </div>	<b>RE</b>
<div>  ▶ Uterus </div>	<div> <div></div> </div>	
<div>  ▶ Myometrium </div>	<div> <div></div> </div>	
<div>  ▶ Bladder </div>	<div> <div></div> </div>	
<div>  ▶ Placenta </div>	<div> <div></div> </div>	
<div>  ▶ Endometrium </div>	<div> <div></div> </div>	
<div>Open 13 more</div>		

## Disease Atlas for Bioset:

**allergezy-2018**

Bioset from study: [18 CpG 13 gene signature](#)

Homo sapiens | **RE** RNA Expression | 13 features (mapped to 13 genes)

[View Bioset Details](#)

CORRELATED DISEASES
CORRELATED STUDIES

View by: Categories
View all disease studies

Disease Category locator

Name	Score 0 50 100	Supporting Data Types	# Studies	Correlation with (filter)
<b>Genetic Disorders and Chromosomal Abnormalities</b>				
■ Congenital chromosomal disease	100	CN	1	positive
■ Down syndrome	59	ME RE	7	negative
■ Anomaly of chromosome pair 21	58	ME RE	7	negative
■ Anomaly of chromosome pair 14	51	RE	2	negative
■ Turner syndrome	49	RE	1	positive

Open 5 more

## Pathway Enrichment for Bioset:

**allergezy-2018**

Bioset from study: [18 CpG 13 gene signature](#)

Homo sapiens | **RE** RNA Expression | 13 features (mapped to 13 genes)

Forward

[View Bioset Details](#)

Filter by:
Biogroup Source
Keyword
Biogroup Size:
Min: 10 genes
Max: 500 genes

Select Biogroup to filter by:
☒ Gene Ontology | GO
☐ miRNA targets | TargetScan
☐ Canonical pathways | Broad MSigDB
☐ Positional gene sets | Broad MSigDB
☐ Protein families | InterPro, PMAP
☐ Regulatory motifs | Broad MSigDB

☒ Gene Ontology | GO
Clear Filters
Cancel

Biogroup Biogroup correlation results
1 to 100 of 107 biogroups

Biogroups	Normalized Score	P-Value	Direction (filter)	Co
fat cell differentiation GO	100	0.0002		2
divalent inorganic cation transmembrane transporter activity GO	89	0.0005		2

- 12 GO provides a framework for analyzing a group of co-expressed genes and provides insights into whether particular genes are involved in diseases. Similar to other ontologies, GO consists of terms and relationships represented in a directed acyclic graph. GO focuses on using the terms in the ontology to describe gene functions. In GO, gene function is classified along three categories: molecular functions, cellular components, and biological processes.



The **GO Enrichment Analysis Tool** was used to find GO terms associated with the 13-gene signature. Enter the following gene list as shown

RNF213  
SARS  
ZNF252  
TMED10P  
ABCF2  
TIMP2  
MAFK  
CD7  
PANX1  
CTBP2  
SLC24A2  
ARID5B  
KIF13B  
FAM190B

The screenshot shows the Gene Ontology Consortium website's GO Enrichment Analysis tool. The header includes the logo and navigation links: Home, Documentation, Downloads, Tools, About, and Contact us. The main content area is titled "GO Enrichment Analysis" and includes a sub-header "Tools / GO Enrichment Analysis". On the left, there is a sidebar titled "Enrichment analysis" with a text input field containing the gene list: CTBP2, SLC24A2, ARID5B, KIF13B, and FAM190B. Below the input field are dropdown menus for "biological process" and "Homo sapiens", and a "Submit" button. At the bottom of the sidebar, there is a "Help" link and a note "Powered by PANTHER". The main content area contains a paragraph explaining the tool's purpose, a section titled "Enrichment analysis tool" with a description of the service, and a section titled "Using the GO enrichment analysis tools" with a numbered list of instructions.

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Tools / GO Enrichment Analysis

## GO Enrichment Analysis

One of the main uses of the GO is to perform enrichment analysis on gene sets. For example, given a regulated under certain conditions, an enrichment analysis will find which GO terms are over-represented for that gene set.

### Enrichment analysis tool

Users can perform enrichment analyses directly from the [home page of the GOC website](#). This service from the [PANTHER Classification System](#), which is maintained up to date with GO annotations. The P explained in great detail in [Mi H et al, PMID: 23868073](#). The list of supported gene IDs is available for

### Using the GO enrichment analysis tools

1. Paste or type the names of the genes to be analyzed, one per row or separated by a comma. specific gene names and UniProt IDs (e.g. Rad54 or P38086).

After computation the results are shown below.



Displaying only results for Bonferroni-corrected for  $P < 0.05$ , [click here to display all results](#)

	<a href="#">Homo sapiens</a> (REF)	<a href="#">upload 1</a> (▼ <a href="#">Hierarchy</a> <b>NEW!</b> <a href="#">?</a> )				
<a href="#">GO biological process complete</a>	#	#	<a href="#">expected</a>	<a href="#">Fold Enrichment</a>	<a href="#">+/-</a>	<a href="#">P value</a>
Unclassified	<a href="#">3250</a>	<a href="#">1</a>	1.86	.54	-	0.00E00

Follow the same procedure for the other two options: molecular function and cellular component, as shown below.

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Tools / GO Enrichment Analysis

## Enrichment analysis

RNF213  
SARS  
ZNF252  
TMED10P  
ABCF2  

biological process  
☒ molecular function  
cellular component  
Homo sapiens

Submit

Help  
Powered by [PANTHER](#)

## GO Enrichment Analysis

One of the main uses of the GO is to perform enrichment analysis on gene sets. For example, given a set regulated under certain conditions, an enrichment analysis will find which GO terms are over-represented annotations for that gene set.

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#### Using the GO enrichment analysis tools

1. Paste or type the names of the genes to be analyzed, one per row or separated by a comma. The specific gene names and UniProt IDs (e.g. Rad54 or P38086).
2. Select the GO aspect (molecular function, biological process, cellular component) for your analy:

The results from this analysis is shown below.



[LOGIN](#) [REGISTER](#) [CONTACT US](#)

[Home](#) [About](#) [PANTHER Data](#) [PANTHER Tools](#) [Workspace](#) [Downloads](#) [Help/Tutorial](#)

**UPL14.0 New!** PANTHER14.0 is generated from the 2018\_04 release of [ReferenceProteome dataset](#)

**Analysis Summary:** Please report in publication [?](#)

<b>Analysis Type:</b> PANTHER Overrepresentation Test (Released 20181113)	
<b>Annotation Version and Release Date:</b> GO Ontology database Released 2018-12-01	
<b>Analyzed List:</b>	upload_1 (Homo sapiens) <a href="#">Change</a>
<b>Reference List:</b>	Homo sapiens (all genes in database) <a href="#">Change</a>
<b>Annotation Data Set:</b> <span>GO molecular function complete</span> <a href="#">?</a>	
<b>Test Type:</b> <input checked="" type="radio"/> Fisher's Exact <input type="radio"/> Binomial	
<b>Correction:</b> <input type="radio"/> Calculate False Discovery Rate <input checked="" type="radio"/> Use the Bonferroni correction for multiple testing <a href="#">?</a> <input type="radio"/> No correction	

**Results** [?](#)

	Reference list	upload_1
Mapped IDs:	<a href="#">20996</a> out of 20996	<a href="#">12</a> out of 12
Unmapped IDs:	<a href="#">0</a>	<a href="#">2</a>
Multiple mapping information:	<a href="#">0</a>	<a href="#">0</a>

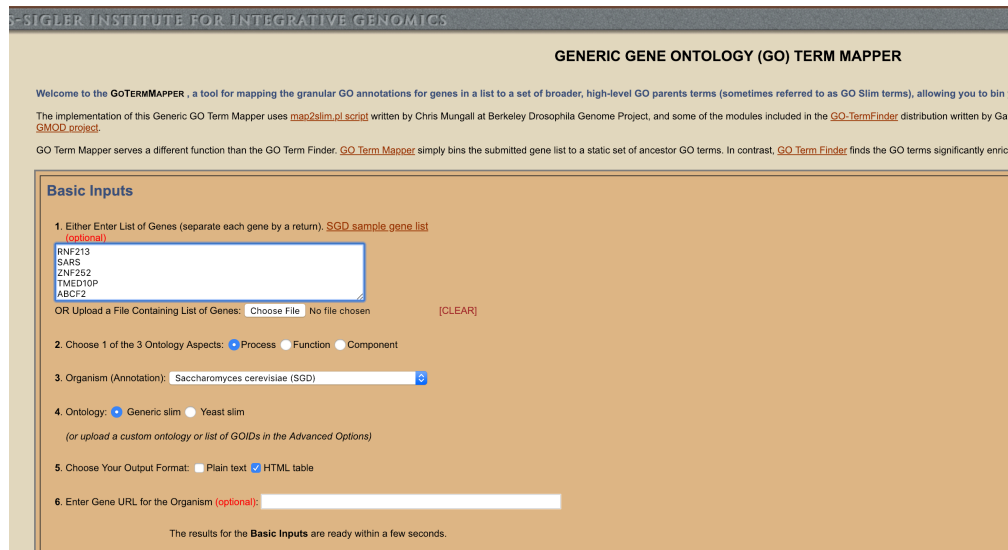
3onferroni count: 2782

[Export results](#)

o statistically significant results. [Click to see all results.](#)

[About](#) | [Relea](#)

Next, the **Generic Gene Ontology (GO) Term Mapper** tool from Princeton University to map granular GO annotations to a higher-level set of terms, thus providing a broad set of categories.



**GENERIC GENE ONTOLOGY (GO) TERM MAPPER**

Welcome to the **GoTERMMapper**, a tool for mapping the granular GO annotations for genes in a list to a set of broader, high-level GO parents terms (sometimes referred to as GO Slim terms), allowing you to bin your gene list.

The implementation of this Generic GO Term Mapper uses [map2slim.pl script](#) written by Chris Mungall at Berkeley Drosophila Genome Project, and some of the modules included in the [GO-TermFinder](#) distribution written by Gavin Sherlock at the [GMOD project](#).

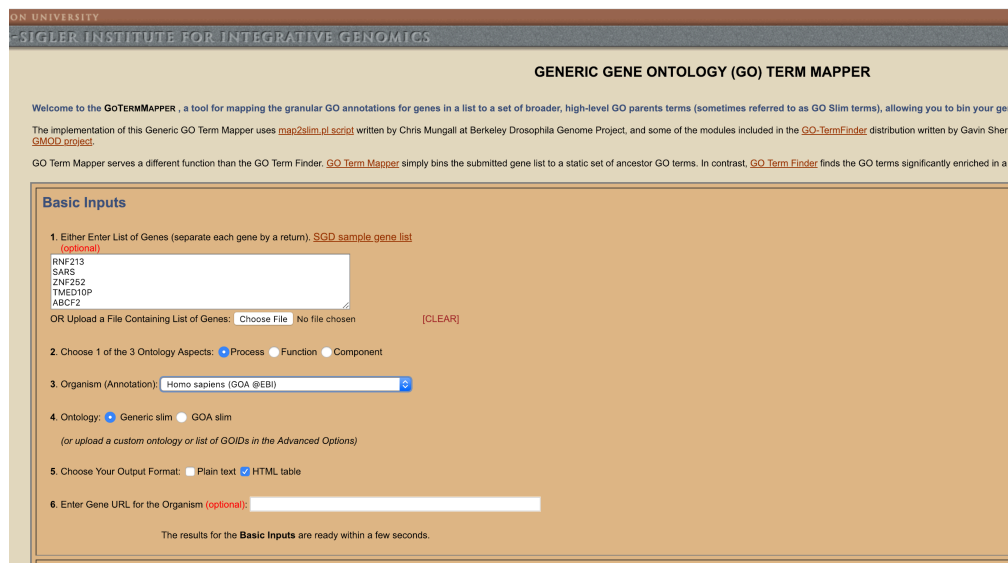
GO Term Mapper serves a different function than the GO Term Finder. [GO Term Mapper](#) simply bins the submitted gene list to a static set of ancestor GO terms. In contrast, [GO Term Finder](#) finds the GO terms significantly enriched in a gene list.

**Basic Inputs**

1. Either Enter List of Genes (separate each gene by a return). [SGD sample gene list](#) (optional)  
  
  
  
  
  
OR Upload a File Containing List of Genes:  No file chosen
2. Choose 1 of the 3 Ontology Aspects: ☒ Process ☐ Function ☐ Component
3. Organism (Annotation):
4. Ontology: ☒ Generic slim ☐ Yeast slim  
(or upload a custom ontology or list of GOIDs in the Advanced Options)
5. Choose Your Output Format: ☐ Plain text ☒ HTML table
6. Enter Gene URL for the Organism (optional):

The results for the **Basic Inputs** are ready within a few seconds.

Make sure to select HomoSapiens for the Organism as shown



**GENERIC GENE ONTOLOGY (GO) TERM MAPPER**

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**Basic Inputs**

1. Either Enter List of Genes (separate each gene by a return). [SGD sample gene list](#) (optional)  
  
  
  
  
  
OR Upload a File Containing List of Genes:  No file chosen
2. Choose 1 of the 3 Ontology Aspects: ☒ Process ☐ Function ☐ Component
3. Organism (Annotation):
4. Ontology: ☒ Generic slim ☐ GOA slim  
(or upload a custom ontology or list of GOIDs in the Advanced Options)
5. Choose Your Output Format: ☐ Plain text ☒ HTML table
6. Enter Gene URL for the Organism (optional):

The results for the **Basic Inputs** are ready within a few seconds.

The results are shown below.

GENERIC GENE ONTOLOGY (GO) TERM MAPPER

SEARCH RESULTS

PLEASE NOTE: You had chosen the GO consortium's and biological\_process for the ontology aspect in the Basic Inputs.

Save Options: [HTML Table](#) | [Plain Text](#) | [Tab-delimited](#) [New](#)  
[Your Input Gene List](#)

Your input list contains 14 genes.

These 2 identifiers were found to be unannotated: ZNF252 TMED10P

GO Terms from the biological_process Ontology			
GO Term (GO ID)	Genes Annotated to the GO Term	GO Term Usage in Gene List	Genome Frequency of Use
signal transduction ( GO:0007165 )	ARID5B, CDZ, CTBP2, KIF13B, RNF213, TIMP2	6 of 12 genes, 50.00%	6032 of 19737 annotated genes, 30.56%
anatomical structure development ( GO:0048856 )	ARID5B, KIF13B, MAPK, RNF213, SARS, TIMP2	6 of 12 genes, 50.00%	8720 of 19737 annotated genes, 28.96%
transport ( GO:0006810 )	CTBP2, KIF13B, PANK1, SLC24A2, TIMP2	5 of 12 genes, 41.67%	5138 of 19737 annotated genes, 26.03%
biosynthetic process ( GO:0009058 )	ARID5B, CTBP2, MAPK, SARS	4 of 12 genes, 33.33%	6194 of 19737 annotated genes, 31.38%
cell differentiation ( GO:0030154 )	ARID5B, CTBP2, KIF13B, TIMP2	4 of 12 genes, 33.33%	4032 of 19737 annotated genes, 20.43%
cellular nitrogen compound metabolic process ( GO:0034641 )	ARID5B, CTBP2, MAPK, SARS	4 of 12 genes, 33.33%	6503 of 19737 annotated genes, 32.95%
cell-cell signaling ( GO:0007267 )	CTBP2, PANK1, RNF213, SLC24A2	4 of 12 genes, 33.33%	1580 of 19737 annotated genes, 8.01%
immune system process ( GO:0002378 )	CDZ, KIF13B, TIMP2	3 of 12 genes, 25.00%	3099 of 19737 annotated genes, 15.70%
cellular protein modification process ( GO:0006464 )	ARID5B, RNF213, TIMP2	3 of 12 genes, 25.00%	4068 of 19737 annotated genes, 20.61%
cellular component assembly ( GO:0022907 )	PANK1, RNF213	2 of 12 genes, 16.67%	2964 of 19737 annotated genes, 15.02%
protein-catalytic complex assembly ( GO:0065003 )	PANK1, RNF213	2 of 12 genes, 16.67%	1863 of 19737 annotated genes, 9.44%
anatomical structure formation involved in morphogenesis ( GO:0048846 )	RNF213, SARS	2 of 12 genes, 16.67%	1987 of 19737 annotated genes, 9.41%
response to stress ( GO:0006950 )	MAPK, PANK1	2 of 12 genes, 16.67%	3875 of 19737 annotated genes, 19.63%
vesicle-mediated transport ( GO:0016192 )	CTBP2, TIMP2	2 of 12 genes, 16.67%	2109 of 19737 annotated genes, 10.69%
cell proliferation ( GO:0008283 )	CTBP2, TIMP2	2 of 12 genes, 16.67%	1989 of 19737 annotated genes, 10.08%
transmembrane transport ( GO:0055085 )	PANK1, SLC24A2	2 of 12 genes, 16.67%	1542 of 19737 annotated genes, 7.81%
catabolic process ( GO:0009056 )	RNF213, TIMP2	2 of 12 genes, 16.67%	2539 of 19737 annotated genes, 12.86%
reproduction ( GO:0000003 )	ARID5B	1 of 12 genes, 8.33%	1419 of 19737 annotated genes, 7.19%
nervous system process ( GO:0050877 )	SLC24A2	1 of 12 genes, 8.33%	1422 of 19737 annotated genes, 7.20%
protein targeting ( GO:0006905 )	KIF13B	1 of 12 genes, 8.33%	423 of 19737 annotated genes, 2.14%
homeostatic process ( GO:0042562 )	SLC24A2	1 of 12 genes, 8.33%	1845 of 19737 annotated genes, 9.35%
cellular amino acid metabolic process ( GO:0006520 )	SARS	1 of 12 genes, 8.33%	335 of 19737 annotated genes, 1.70%
extracellular matrix organization ( GO:0001198 )	TIMP2	1 of 12 genes, 8.33%	355 of 19737 annotated genes, 1.80%
cytoskeleton organization ( GO:007010 )	FAM192B	1 of 12 genes, 8.33%	1304 of 19737 annotated genes, 6.61%
translation ( GO:0006412 )	SARS	1 of 12 genes, 8.33%	640 of 19737 annotated genes, 3.24%
mitotic cell cycle ( GO:0000278 )	TIMP2	1 of 12 genes, 8.33%	920 of 19737 annotated genes, 4.66%
small molecule metabolic process ( GO:0044281 )	SARS	1 of 12 genes, 8.33%	2016 of 19737 annotated genes, 10.21%
synthesis process ( GO:0044403 )	CTBP2	1 of 12 genes, 8.33%	778 of 19737 annotated genes, 3.94%
cell modify ( GO:0048870 )	ARID5B	1 of 12 genes, 8.33%	1594 of 19737 annotated genes, 8.08%
cell morphogenesis ( GO:0009092 )	KIF13B	1 of 12 genes, 8.33%	1010 of 19737 annotated genes, 5.12%
RNA metabolic process ( GO:0006399 )	SARS	1 of 12 genes, 8.33%	194 of 19737 annotated genes, 0.98%
chromosome organization ( GO:0051276 )	ARID5B	1 of 12 genes, 8.33%	1172 of 19737 annotated genes, 5.94%
suicy ( GO:0007558 )	TIMP2	1 of 12 genes, 8.33%	303 of 19737 annotated genes, 1.54%
locomotion ( GO:0040011 )	ARID5B	1 of 12 genes, 8.33%	1820 of 19737 annotated genes, 9.22%

The results are summarized in the following table.



	GO Term	GO ID	Genes Annotated to the GO Term	GO Term Usage in Gene List	Genome Frequency x of 19497 annotated genes
1	signal transduction	GO:0007165	ARID5B, CD7, CTBP2, KIF13B, RNF213, TIMP2	6 of 12 genes, 50.00%	6030
2	anatomical structure development	GO:0048856	ARID5B, CD7, CTBP2, KIF13B, RNF213, TIMP2	6 of 12 genes, 50.00%	5588
3	biosynthetic process	GO:0009058	ARID5B, CTBP2, MAFK, SARS	4 of 12 genes, 33.33%	6416
4	cell differentiation	GO:0030154	ARID5B, CTBP2, KIF13B, TIMP2	4 of 12 genes, 33.33%	3981
5	cellular nitrogen compound metabolic process	GO:0034641	ARID5B, CTBP2, MAFK, SARS	4 of 12 genes, 33.33%	6622
6	transport	GO:0006810	KIF13B, PANX1, SLC24A2, TIMP2	4 of 12 genes, 33.33%	4956
7	immune system process	GO:0002376	CD7, KIF13B, TIMP2	3 of 12 genes, 25.00%	2974
8	cell-cell signaling	GO:0007267	PANX1, RNF213, SLC24A2	3 of 12 genes, 25.00%	1593
9	cellular protein modification process	GO:0006464	ARID5B, RNF213, TIMP2	3 of 12 genes, 25.00%	4113
10	cellular component assembly	GO:0022607	PANX1, RNF213	2 of 12 genes, 16.67%	2713
11	protein-containing complex assembly	GO:0065003	PANX1, RNF213	2 of 12 genes, 16.67%	1686
12	anatomical structure formation involved in morphogenesis	GO:0048646	RNF213, SARS	2 of 12 genes, 16.67%	1020
13	response to stress	GO:0006950	MAFK, PANX1	2 of 12 genes, 16.67%	3830
14	cell proliferation	GO:0008283	CTBP2, TIMP2	2 of 12 genes, 16.67%	1973
15	transmembrane transport	GO:0055085	PANX1, SLC24A2	2 of 12 genes, 16.67%	1534
16	catabolic process	GO:0009056	RNF213, TIMP2	2 of 12 genes, 16.67%	2443
17	reproduction	GO:0000003	ARID5B	1 of 12 genes, 8.33%	1391
18	cytoskeleton-dependent intracellular transport	GO:0030705	KIF13B	1 of 12 genes, 8.33%	157
19	nervous system process	GO:0050877	SLC24A2	1 of 12 genes, 8.33%	1364
20	protein targeting	GO:0006605	KIF13B	1 of 12 genes, 8.33%	397
21	homeostatic process	GO:0042592	SLC24A2	1 of 12 genes, 8.33%	1641
22	cellular amino acid metabolic process	GO:0006520	SARS	1 of 12 genes, 8.33%	367
23	extracellular matrix organization	GO:0030198	TIMP2	1 of 12 genes, 8.33%	335
24	cytoskeleton organization	GO:0007010	FAM190B	1 of 12 genes, 8.33%	1212
25	translation	GO:0006412	SARS	1 of 12 genes, 8.33%	604
26	mitotic cell cycle	GO:0000278	TIMP2	1 of 12 genes, 8.33%	976
27	small molecule metabolic process	GO:0044281	SARS	1 of 12 genes, 8.33%	2081
28	vesicle-mediated transport	GO:0016192	TIMP2	1 of 12 genes, 8.33%	2000
29	symbiont process	GO:0044403	CTBP2	1 of 12 genes, 8.33%	841
30	cell motility	GO:0048870	ARID5B	1 of 12 genes, 8.33%	1488
31	cell morphogenesis	GO:0000902	KIF13B	1 of 12 genes, 8.33%	932
32	tRNA metabolic process	GO:0006399	SARS	1 of 12 genes, 8.33%	188
33	chromosome organization	GO:0051276	ARID5B	1 of 12 genes, 8.33%	1174
34	aging	GO:0007568	TIMP2	1 of 12 genes, 8.33%	284
35	locomotion	GO:0040011	ARID5B	1 of 12 genes, 8.33%	1721
36	growth	GO:0040007	ARID5B	1 of 12 genes, 8.33%	922
37	cell cycle	GO:0007049	TIMP2	1 of 12 genes, 8.33%	1793

The Generic Gene Ontology (GO) Term Mapper tool [41] was used to map granular GO annotations to a broader, higher-level set of terms. The last column should be read as “xxx of 19497 annotated genes”. Note that two genes, ZNF252 and TMED10P, did not map to any GO terms.

The resulting GO terms are

GO:0030705

GO:0007165

GO:0048856

GO:0006810

GO:0009058

GO:0030154

GO:0034641

GO:0007267

GO:0002376

GO:0006464

GO:0022607

GO:0065003



GO:0048646  
GO:0006950  
GO:0016192  
GO:0008283  
GO:0055085  
GO:0009056  
GO:0000003  
GO:0050877  
GO:0006605  
GO:0042592  
GO:0006520  
GO:0030198  
GO:0007010  
GO:0006412  
GO:0000278  
GO:0044281  
GO:0044403  
GO:0048870  
GO:0000902  
GO:0006399  
GO:0051276  
GO:0007568  
GO:0040011  
GO:0040007  
GO:0007049

Next, the **REVIGO**, an online tool that summarizes and visualizes lists of gene ontology terms, to find a representative set of terms using a clustering algorithm.



# REVIGO

reduce + visualize Gene ontology



Rudjer Boskovic Institute,  
Croatia

Like 180 Tweet G+

## Welcome to REVIGO!

REVIGO can take long lists of Gene Ontology terms and summarize them by removing redundant GO terms. The remaining terms can be visualized in semantic similarity-based scatterplots, interactive graphs, or tag clouds. [More about REVIGO...](#)

Please enter a list of Gene Ontology IDs below, each on its own line. The GO IDs may be followed by p-values or another quantity which describes the GO term in a way meaningful to you.

Examples: #1 #2 #3

```
GO:0030705
GO:0007165
GO:0048856
GO:0006810
GO:0009058
GO:0030154
GO:0034641
GO:0007267
GO:0002376
GO:0006464
GO:0022807
GO:0065003
GO:0048646
GO:0006950
GO:0016192
GO:0008283
GO:0055085
GO:0009056
GO:0000003
GO:0050877
```

Allowed similarity: How large would you like the resulting list to be?

☐ Large (allowed similarity=0.9) ☒ Medium (0.7) ☐ Small (0.5) ☐ Tiny (0.4)

If provided, the numbers associated to GO categories are...

☒ p-values  
☐ some other quantity, where higher is better

## Q + A

**Q:** I have a list of interesting genes, but not a list of GO terms.

**A:** You can use one of the following web servers to search for GO terms that are overrepresented in your list of genes:

- [GOtilla](#) (multiple eukaryotes)
- [AgriGO](#) (many plants, several animals)
- [L2L](#) (human, mouse and rat)
- [GO Term Finder](#) (many species)
- [FatiGO](#) (multiple eukaryotes)

After that, return here with the list of GO terms and p-values (or enrichments).

**Q:** I still don't have a list of interesting genes, but I'd like to try out my favourite GO enrichment tool and then bring the output to REVIGO to summarize and visualize.

**A:** Here are the links to two example gene sets, one from [agriGO](#) (click "Example") and another one from [DAVID](#) (click "Demolist\_1").

**Q:** The organism I work on is not listed in the "GO term sizes" box in Advanced options.

**A:** The chosen database is used to find the size of each GO term i.e. the percentage of genes annotated with the term. This quantity determines the size of bubbles in the visualizations, thus indicating a more general GO term (larger) or a more specific one (smaller). The choice of database also has some influence on the GO term clustering/selection process, and on the bubble placement in the visualizations. If your organism is

The frequency and uniqueness of the GO terms can be seen below.

[Hide/show dispensable GO terms](#) [Export results to text table \(CSV\)](#) [Make R script for plotting](#)

term ID	description	frequency	pin?	uniqueness	dispensability
GO:0000003	reproduction	7.871 %		1.00	0.00
GO:0002376	immune system process	16.463 %		0.95	0.00
GO:0006399	tRNA metabolic process	1.016 %		0.90	0.00
GO:0006950	response to stress	21.310 %		0.94	0.00
GO:0007568	aging	1.633 %		0.89	0.00
GO:0030705	cytoskeleton-dependent intracellular transport	0.825 %		0.87	0.00
GO:0040007	growth	5.447 %		0.95	0.00
GO:0040011	locomotion	9.452 %		0.95	0.00
GO:0042592	homeostatic process	9.371 %		0.95	0.00
GO:0044403	symbiosis, encompassing mutualism through parasitism	4.322 %		0.95	0.00
GO:0050877	neurological system process	7.438 %		0.95	0.00
GO:0051276	chromosome organization	3.566 %		0.87	0.00
GO:0008283	cell proliferation	11.321 %		0.94	0.02
GO:0007049	cell cycle	10.000 %		0.92	0.02
GO:0009058	biosynthetic process	36.924 %		0.93	0.05
GO:0000278	mitotic cell cycle	5.822 %		0.92	0.07
GO:0009056	catabolic process	11.471 %		0.92	0.10
GO:0044281	small molecule metabolic process	12.072 %		0.90	0.10
GO:0006412	translation	3.693 %		0.87	0.16
GO:0030198	extracellular matrix organization	1.829 %		0.87	0.18
GO:0022607	cellular component assembly	15.638 %		0.86	0.26
GO:0065003	macromolecular complex assembly	9.977 %		0.86	0.70
GO:0034641	cellular nitrogen compound metabolic process	37.432 %		0.90	0.26
GO:0048856	anatomical structure development	31.558 %		0.88	0.27
GO:0007165	signal transduction	33.618 %		0.91	0.27
GO:0006810	transport	29.215 %		0.86	0.28
GO:0048870	cell motility	8.257 %		0.85	0.35
GO:0048646	anatomical structure formation involved in morphogenesis	5.591 %		0.87	0.35
GO:0007267	cell-cell signaling	9.025 %		0.90	0.37
GO:0006464	cellular protein modification process	22.683 %		0.90	0.38
GO:0007010	cytoskeleton organization	6.861 %		0.87	0.44
GO:0055085	transmembrane transport	8.113 %		0.86	0.45
GO:0006520	cellular amino acid metabolic process	2.152 %		0.85	0.49
GO:0016192	vesicle-mediated transport	11.385 %		0.86	0.50
GO:0006605	protein targeting	4.010 %		0.85	0.52
GO:0030154	cell differentiation	22.095 %		0.84	0.54
GO:0000902	cell morphogenesis	5.297 %		0.78	0.58

Click on the TreeMap tab.

# REVIGO

Biological Process (37)

Tag Clouds

Scatterplot & Table

Interactive Graph

TreeMap

Download the csv file at the bottom of the page. You should see a csv file with the following content.



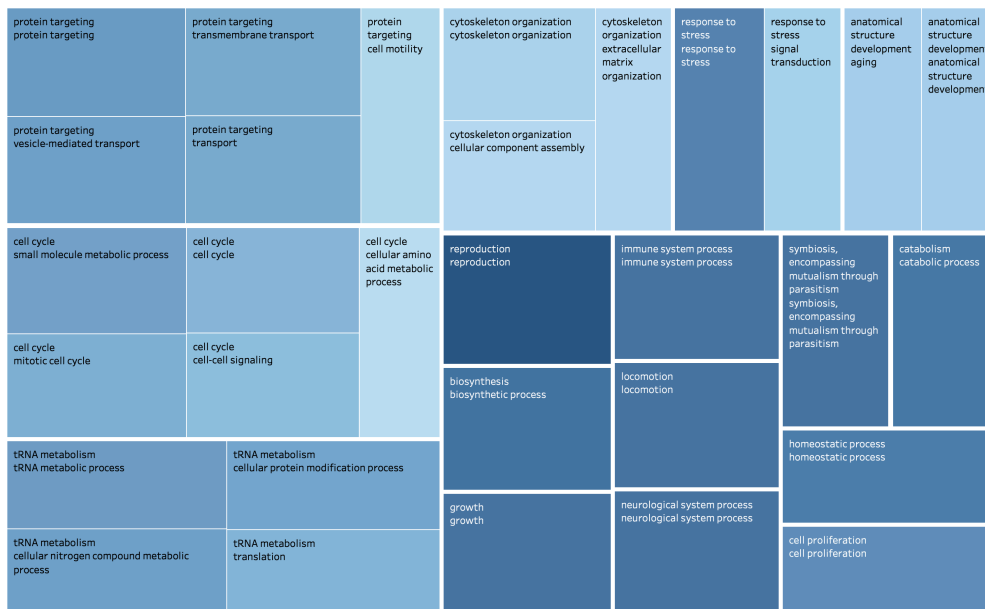


% WARNING - This is exported REVIGO data useful only for the specific purpose of constructing a TreeMap visualization.					
% Do not use this tab as it sets an extremely permissive					
% threshold to detect while normally $c \geq 0.4$ is recommended.					
% To export a reduce go to the Scatterplot & Table tab					
and export to CSV from there.					
term_ID	description	frequency	uniqueness	dispensability	representative
GO:0000003	reproduction	0.77%	1	0	reproduction
GO:0002376	immune system process	0.60%	0.946	0	immune system process
GO:0006950	response to stress	4.58%	0.914	0	response to stress
GO:0007165	signal transduction	6.62%	0.783	0.637	response to stress
GO:0008283	cell proliferation	0.39%	0.897	0	cell proliferation
GO:0009056	catabolic process	4.82%	0.938	0	catabolism
GO:0016192	vesicle-mediated transport	1.09%	0.857	0	vesicle-mediated transport
GO:0006605	protein targeting	0.69%	0.838	0.657	vesicle-mediated transport
GO:0030705	cytoskeleton-dependent intracellular transport	0.06%	0.855	0.238	vesicle-mediated transport
GO:0055085	transmembrane transport	8.92%	0.838	0.684	vesicle-mediated transport
GO:0006810	transport	17.62%	0.835	0.454	vesicle-mediated transport
GO:0048870	cell motility	0.63%	0.762	0.287	vesicle-mediated transport
GO:0040007	growth	0.32%	0.945	0	growth
GO:0040011	locomotion	1.00%	0.946	0	locomotion
GO:0042592	homeostatic process	1.66%	0.926	0	homeostatic process
GO:0044403	symbiosis, encompassing mutualism through	0.17%	0.945	0	symbiosis, encompassing mutualism through parasitism
GO:0048856	anatomical structure development	2.54%	0.787	0	anatomical structure development
GO:0007568	aging	0.09%	0.787	0.648	anatomical structure development
GO:0050877	neurological system process	0.50%	0.945	0	neurological system process
GO:0006399	tRNA metabolic process	2.50%	0.872	0.021	tRNA metabolism
GO:0034641	cellular nitrogen compound metabolic process	34.14%	0.866	0.244	tRNA metabolism
GO:0006412	translation	5.69%	0.831	0.614	tRNA metabolism
GO:0006464	cellular protein modification process	7.73%	0.857	0.225	tRNA metabolism
GO:0009058	biosynthetic process	31.61%	0.947	0.033	biosynthesis
GO:0007010	cytoskeleton organization	0.79%	0.783	0.038	cytoskeleton organization
GO:0030198	extracellular matrix organization	0.06%	0.768	0.397	cytoskeleton organization
GO:0022607	cellular component assembly	2.48%	0.768	0.57	cytoskeleton organization
GO:0007049	cell cycle	1.89%	0.828	0.086	cell cycle
GO:0007267	cell-cell signaling	0.41%	0.818	0.197	cell cycle
GO:0000278	mitotic cell cycle	0.56%	0.841	0.204	cell cycle
GO:0006520	cellular amino acid metabolic process	5.59%	0.763	0.329	cell cycle
GO:0044281	small molecule metabolic process	15.14%	0.861	0.139	cell cycle

This can be summarized by the following table.

	Representative Terms	GO Term (GO ID)	Uniqueness
1	anatomical structure development	aging (GO:0007568)	0.781
		anatomical structure development (GO:0048856)	0.781
2	biosynthesis	biosynthetic process (GO:0009058)	0.946
3	catabolism	catabolic process (GO:0009056)	0.936
4	cell cycle	cellular amino acid metabolic process (GO:0006520)	0.757
		cell-cell signaling (GO:0007267)	0.813
		cell cycle (GO:0007049)	0.813
		mitotic cell cycle (GO:0000278)	0.836
		small molecule metabolic process (GO:0044281)	0.858
5	cell proliferation	cell proliferation (GO:0008283)	0.894
6	cytoskeleton organization	extracellular matrix organization (GO:0030198)	0.762
		cellular component assembly (GO:0022607)	0.762
		cytoskeleton organization (GO:0007010)	0.777
7	growth	growth (GO:0040007)	0.944
8	homeostatic process	homeostatic process (GO:0042592)	0.924
9	<b>immune system process</b>	immune system process (GO:0002376)	0.944
10	locomotion	locomotion (GO:0040011)	0.944
11	neurological system process	neurological system process (GO:0050877)	0.944
12	protein targeting	cell motility (GO:0048870)	0.767
		transport (GO:0006810)	0.847
		transmembrane transport (GO:0055085)	0.848
		vesicle-mediated transport (GO:0016192)	0.865
		protein targeting (GO:0006605)	0.869
13	reproduction	reproduction (GO:0000003)	1
14	<b>response to stress</b>	signal transduction (GO:0007165)	0.778
		response to stress (GO:0006950)	0.911
15	symbiosis, encompassing mutualism through parasitism	symbiosis, encompassing mutualism through parasitism (GO:0044403)	0.944
16	tRNA metabolism	translation (GO:0006412)	0.827
		cellular protein modification process (GO:0006464)	0.853
		cellular nitrogen compound metabolic process (GO:0034641)	0.862
		tRNA metabolic process (GO:0006399)	0.868

This same data can be represented in a treemap.



Representative	Description	term ID	
<b>anatomical structure development</b>	aging	GO:0007568	<a href="#">0.7810</a>
	anatomical structure dev..	GO:0048856	<a href="#">0.7810</a>
<b>biosynthesis</b>	biosynthetic process	GO:0009058	<a href="#">0.9460</a>
<b>catabolism</b>	catabolic process	GO:0009056	<a href="#">0.9360</a>
<b>cell cycle</b>	cell cycle	GO:0007049	<a href="#">0.8230</a>
	cell-cell signaling	GO:0007267	<a href="#">0.8130</a>
	cellular amino acid metab..	GO:0006520	<a href="#">0.7570</a>
	mitotic cell cycle	GO:0000278	<a href="#">0.8360</a>
	small molecule metabolic ..	GO:0044281	<a href="#">0.8580</a>
<b>cell proliferation</b>	cell proliferation	GO:0008283	<a href="#">0.8940</a>
<b>cytoskeleton organization</b>	cellular component assem..	GO:0022607	<a href="#">0.7620</a>
	cytoskeleton organization	GO:0007010	<a href="#">0.7770</a>
	extracellular matrix orga..	GO:0030198	<a href="#">0.7620</a>
<b>growth</b>	growth	GO:0040007	<a href="#">0.9440</a>
<b>homeostatic process</b>	homeostatic process	GO:0042592	<a href="#">0.9240</a>
<b>immune system process</b>	immune system process	GO:0002376	<a href="#">0.9440</a>
<b>locomotion</b>	locomotion	GO:0040011	<a href="#">0.9440</a>
<b>neurological system proc..</b>	neurological system proc..	GO:0050877	<a href="#">0.9440</a>
<b>protein targeting</b>	cell motility	GO:0048870	<a href="#">0.7670</a>
	protein targeting	GO:0006605	<a href="#">0.8690</a>
	transmembrane transport	GO:0055085	<a href="#">0.8480</a>
	transport	GO:0006810	<a href="#">0.8470</a>
	vesicle-mediated transport	GO:0016192	<a href="#">0.8650</a>
<b>reproduction</b>	reproduction	GO:0000003	<a href="#">1.0000</a>
<b>response to stress</b>	response to stress	GO:0006950	<a href="#">0.9110</a>
	signal transduction	GO:0007165	<a href="#">0.7780</a>
<b>symbiosis, encompassing ..</b>	symbiosis, encompassing ..	GO:0044403	<a href="#">0.9440</a>
<b>tRNA metabolism</b>	cellular nitrogen compoun..	GO:0034641	<a href="#">0.8620</a>
	cellular protein modificati..	GO:0006464	<a href="#">0.8530</a>
	translation	GO:0006412	<a href="#">0.8270</a>
	tRNA metabolic process	GO:0006399	<a href="#">0.8680</a>

The GO terms can be visualized using [NaviGo](#).



An analytic tool for Gene Ontology Visualization and Similarity

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Computes GO Term Similarity and Association Scores

For an input set of GO terms, 3 similarity scores and 3 associations scores are computed for every GO term pairs. Results are provided in a table and also visualized as a network and a bubble map.

Upload a file:

[Choose File](#) No file chosen

Notice the uploaded file format should be:

GO:0033301,GO:0008283,GO:0060718,GO:0006956,GO:0006958,

Please enter the go terms that belong to the **same** category  
i.e: Biological Process, Molecular Function and Cellular Component

[Load Sample](#)

[Reset](#)

Input GO Terms:

GO:0030705 x GO:0007165 x GO:0048856 x GO:0006810 x GO:0009058 x GO:0030154 x GO:0034641 x  
GO:0007267 x GO:0002376 x GO:0006464 x GO:0022607 x GO:0065003 x GO:0048646 x GO:0006950 x  
GO:0016192 x GO:0008283 x GO:0055085 x GO:0009056 x GO:0000003 x GO:0050877 x GO:0006605 x  
GO:0042592 x GO:0006520 x GO:0030198 x GO:0007010 x GO:0006412 x GO:0000278 x GO:0044281 x  
GO:0044403 x GO:0048870 x GO:0000902 x GO:0006399 x GO:0051276 x GO:0007568 x GO:0040011 x  
GO:0040007 x GO:0007049 x

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Results are shown below.

## NaviGO Results

[Home](#) [GO Set Result](#) [Network Visualization](#) [Multidimensional Scaling Visualization](#)

BP: ● MF: ● CC: ●

GO terms input by the user:

```
GO:0030705 1
GO:0007165 1
GO:0048856 1
GO:0006810 1
GO:0009058 1
GO:0030154 1
GO:0034641 1
GO:0007267 1
GO:0002276 1
```

[\[?\] Open BP Visualizer](#)

### GO term Pairwise Scores Results

Go term pairwise scores are listed in the table below and also visualized as a network and with a bubble map with the multi-dimensional scaling from the tabs above.

#### GO Term Pair Scores

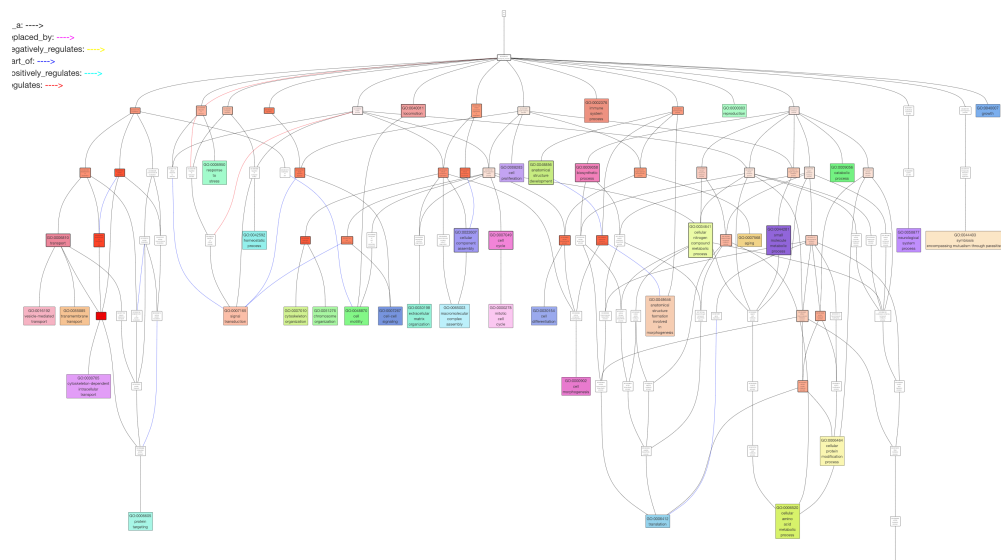
For all the input GO term pairs, 3 GO semantic similarity scores, Resnik, Lin's (LSS), Relevance (RSS), and 3 GO associations scores, GO Co-occurrence (CAS), Pubmed (PAS), protein Interaction (IAS), are computed. For the definition of the scores, see [here](#) . Results can be downloaded in a [CSV file](#) .

**B** :Biological Process, **M** :Molecular Function, **C** :Cellular Component

[\[?\] High](#)    [Low](#)

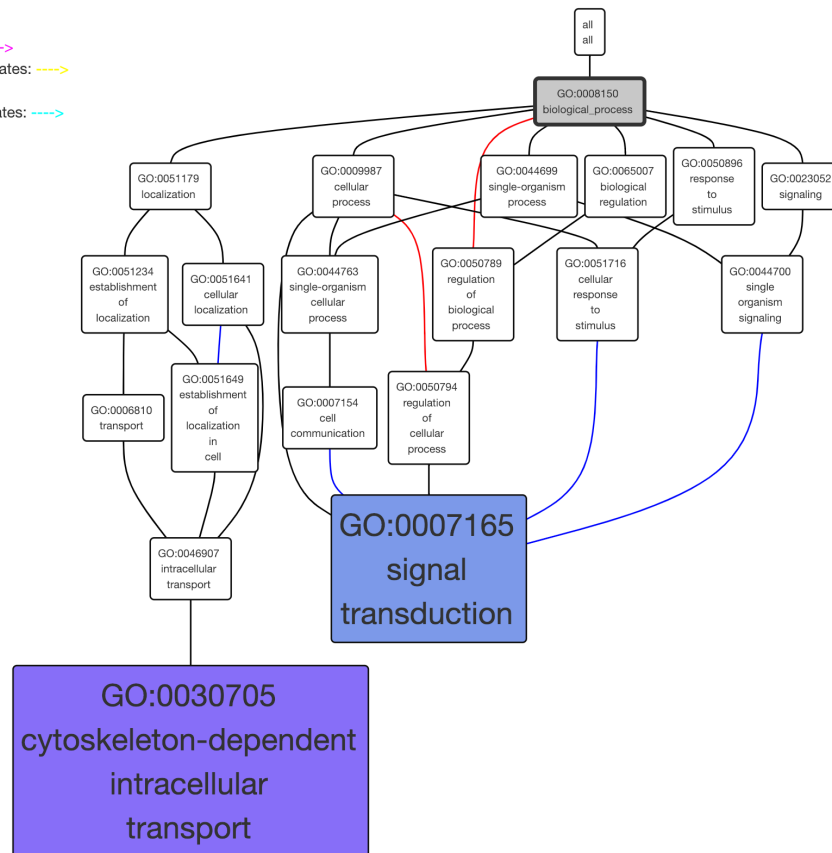
GO term1	GO term2	Resnik	LSS	RSS	CAS	PAS	IAS	Common Parents
<b>B</b> GO:0030705	<b>B</b> GO:0007165	-0.082	-0.039	0.008	0.020	0.001	21.044	<a href="#">Show parents[+]</a> <a href="#">vis</a>
<b>B</b> GO:0030705	<b>B</b> GO:0048856	-0.082	-0.041	0.009	0.032	n/a	n/a	<a href="#">Show parents[+]</a> <a href="#">vis</a>
<b>B</b> GO:0030705	<b>B</b> GO:0006810	1.050	0.519	0.473	0.149	0.003	12.565	<a href="#">Show parents[+]</a> <a href="#">vis</a>
<b>B</b> GO:0030705	<b>B</b> GO:0009058	-0.082	-0.048	0.010	0.008	0.000	n/a	<a href="#">Show parents[+]</a> <a href="#">vis</a>
<b>B</b> GO:0030705	<b>B</b> GO:0030154	-0.082	-0.037	0.008	0.050	0.000	12.206	<a href="#">Show parents[+]</a> <a href="#">vis</a>
<b>B</b> GO:0030705	<b>B</b> GO:0034641	-0.082	-0.048	0.010	0.004	n/a	26.175	<a href="#">Show parents[+]</a> <a href="#">vis</a>
<b>B</b> GO:0030705	<b>B</b> GO:0007267	-0.082	-0.033	0.007	0.054	n/a	10.701	<a href="#">Show parents[+]</a> <a href="#">vis</a>

Click on Open BP Visualizer to see the network of terms.



Single GO terms can also be visualized, as shown below.

is\_a: ---->  
replaced\_by: ---->  
negatively\_regulates: ---->  
part\_of: ---->  
positively\_regulates: ---->  
regulates: ---->



See [https://link.springer.com/protocol/10.1007/978-1-4939-3743-1\\_15](https://link.springer.com/protocol/10.1007/978-1-4939-3743-1_15) for a list of GO visualization tools.