

Sep 22, 2021

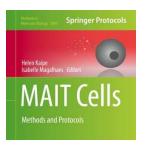
# C Lung Homogenization



In 1 collection

DOI

dx.doi.org/10.17504/protocols.io.bmgzk3×6



Timothy S C Hinks<sup>1</sup>, Bonnie van Wilgenburg<sup>2</sup>, Huimeng Wang<sup>3</sup>, Liyen Loh<sup>3</sup>, Marios Koutsakos<sup>3</sup>, Katherine Kedzierska<sup>3</sup>, Alexandra J. Corbett<sup>3</sup>, Zhenjun Chen<sup>3</sup>

<sup>1</sup>Respiratory Medicine Unit, Nuffield Department of Medicine Experimental Medicine, University of Oxford, Oxfordshire, UK;

<sup>2</sup>Peter Medawar Building for Pathogen Research and Translational Gastroenterology Unit, Nuffield Department of Clinical Medicine, University of Oxford, Oxford, UK;

<sup>3</sup>Department of Microbiology and Immunology, The Peter Doherty Institute for Infection and Immunity, The University of Melbourne, Parkville, Australia

Springer Nature Books



#### Satyavati Kharde

Springer Nature

### Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

OPEN ACCESS





DOI: https://dx.doi.org/10.17504/protocols.io.bmgzk3x6

External link: https://link.springer.com/protocol/10.1007/978-1-0716-0207-2\_17

**Protocol Citation:** Timothy S C Hinks, Bonnie van Wilgenburg, Huimeng Wang, Liyen Loh, Marios Koutsakos, Katherine Kedzierska, Alexandra J. Corbett, Zhenjun Chen 2021. Lung Homogenization. **protocols.io** 

https://dx.doi.org/10.17504/protocols.io.bmgzk3x6

**License:** This is an open access protocol distributed under the terms of the **Creative Commons Attribution License**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

Created: September 18, 2020

Last Modified: September 22, 2021

Protocol Integer ID: 42233

**Keywords:** Virus, MAIT cell, Flow cytometry, MR1-tetramer, Infection, Mouse, pulmonary mait cell, synthetic mait cell antigen, combination of synthetic mait cell antigen, mait cell activation, study of mait cell activation, viral infections in vivo, mait cell, intracellular cytokine, cytokine, methods for lung homogenization, lung homogenization, viral infection, range of viral infection, independent manner via cytokine, specific pathogen, like lymphocyte, lung, infection, murine model,

#### Abstract

This is part 3.4 of the "Study of MAIT Cell Activation in Viral Infections In Vivo" collection of protocols.

Collection Abstract: MAIT cells are abundant, highly evolutionarily conserved innate-like lymphocytes expressing a semi-invariant T cell receptor (TCR), which recognizes microbially derived small intermediate molecules from the riboflavin biosynthetic pathway. However, in addition to their TCR-mediated functions they can also be activated in a TCR-independent manner via cytokines including IL-12, -15, -18, and type I interferon. Emerging data suggest that they are expanded and activated by a range of viral infections, and significantly that they can contribute to a protective anti-viral response. Here we describe methods used to investigate these anti-viral functions in vivo in murine models. To overcome the technical challenge that MAIT cells are rare in specific pathogen-free laboratory mice, we describe how pulmonary MAIT cells can be expanded using intranasal bacterial infection or a combination of synthetic MAIT cell antigen and TLR agonists. We also describe protocols for adoptive transfer of MAIT cells, methods for lung homogenization for plaque assays, and surface and intracellular cytokine staining to determine MAIT cell activation.



#### **Attachments**



Study of MAIT Cell A...

386KB

#### **Materials**

For materials, please refer to the Guidelines section of the "Study of MAIT Cell Activation in Viral Infections In Vivo" collection.

## **Troubleshooting**

## Safety warnings



• Personal protective equipment (PPE) should be worn at all times (gloves, lab coat, & eye protection) (see Notes 3 and 4).

For hazard information and safety warnings, please refer to the SDS (Safety Data Sheet).



- 1 Collect the lungs into 4 2 mL RPMI supplemented with penicillin/streptomycin.
- For homogenization, place the lung and the 2 mL media into 10 mL falcon tubes with lids (see **Note 16**).
- Prepare 10 or 15 mL Falcon tubes with 2 × \$\frac{\pi}{\pi}\$ 5 mL 80%w/v EtOH for cleaning the homogenization probe initially and 1 tube containing HBSS. For each group of samples, prepare further 1 × \$\frac{\pi}{\pi}\$ 5 mL EtOH and 1 × \$\frac{\pi}{\pi}\$ 5 mL HBSS , and for the final probe clean set up 2 × \$\frac{\pi}{\pi}\$ 5 mL EtOH .
- Homogenize the sample using a homogenizer, mounted on a retort stand with the probe set to medium for 00:00:30 per sample. Keep samples on ice (see Note 17).
- Centrifuge the samples at  $\bigcirc$  1000 x g, 00:07:00.
- Using a 1 mL pipette, carefully draw up approximately 1 mL supernatant (a little bit more is good), avoiding the pellet and fatty residue on top. Divide this volume into two 1.5 mL Eppendorf tubes. Store at -80 °C for subsequent plaque assays.