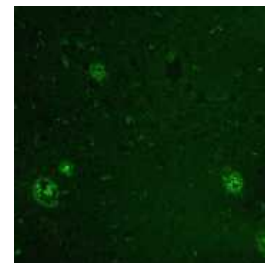


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🌐 Luminescent Conjugated Oligothiophenes (LCO) Staining

DOI

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Protocol status: Working

We use this protocol and it's working

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Abstract

The protocol describes Luminescent Conjugated Oligothiophenes staining on brain sections.

Materials

- 100 % ethanol
- Coverslips
- DEPC H₂O
- Epifluorescence microscope (Leica) using FITC filters
- Humidified chamber for 30 mins
- HydromountTM aqueous non-fluorescing mounting media (National Diagnostics)
- Hydrophobic ImmEdgeTM PAP pen (Vector laboratories)
- PBS
- pFTAA solution (diluted in PBS).
- Xylene



Safety warnings

- ⚠ Breathing in xylene vapours in the air can cause irritation to the eyes, nose and throat and it can cause irritation, redness and swelling to the skin or eyes; and headaches, dizziness, vertigo and drowsiness. Please work to your institution approved health & safety policies, risk assessments and local procedures for the safe use, storage and disposal of xylene. Refer to the data safety sheet.

Preparation

1 Generate tissue sections using standard microtome sectioning protocols.

2 Heat tissue dry tissue sections for  01:00:00 at  60 °C

1h

Deparaffinisation and LCO Staining

45m

3

15m

Safety information

Please refer to the Xylene Data Safety Sheet and follow local approved risk assessments and protocols for safe handling and disposal of xylene.

Hazard pictograms



De-paraffinise sections by  00:05:00 washes in xylene (x2),  00:05:00 100 % ethanol (x2) and  00:05:00 in DEPC H₂O (x1).

4 Circle tissue on slide using a hydrophobic ImmEdge™ PAP pen (Vector laboratories) to contain solution.



- 5 Prepare 1:500 pFTAA solution (diluted in PBS). **All steps from here should be performed away from light.**
- 6 Pipette pFTAA solution onto tissue and incubate at room temperature in a humidified chamber for  00:30:00 mins.
- 7 Wash slides in PBS 3x.
- 8 Coverslip slides using Hydromount™ aqueous non-fluorescing mounting media (National Diagnostics).

30m