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LRRK2 expression and purification

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Protocol status: Working

We use this protocol and it's working

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Abstract

This protocol details methods for the expression of human LRRK2 in Expi293F cells and its *in vitro* purification.

Attachments



[iut8bvk9p.docx](#)

20KB

Materials

⊗ ExpiFectamine[®]; 293 Transfection Kit **Thermo Fisher Catalog #A14525**

⊗ Prescission Protease **Genscript Catalog #Z02799**

⊗ 3xFLAG Peptide **Merck MilliporeSigma (Sigma-Aldrich) Catalog #F4799**

⊗ cOmplete[™], EDTA-free Protease Inhibitor Cocktail **Merck MilliporeSigma (Sigma-Aldrich) Catalog #05056489001**

Glutathione Sepharose (GE Healthcare, 17075601)

⊗ Slide-A-Lyzer[®]; MINI Dialysis Device, 10K MWCO, 0.1 mL **Thermo Fisher Catalog #69572**

⊗ Amicon Ultra-15 Centrifugal Filter Unit **Merck MilliporeSigma (Sigma-Aldrich) Catalog #UFC901024 & UFC903024**

⊗ Monoclonal ANTI-FLAG M2 resin **Merck MilliporeSigma (Sigma-Aldrich) Catalog #F3165**

EDTA-free protease inhibitor cocktail (Roche).

Solutions to prepare:

Lysis salt buffer:

	A	B
	HEPES (7.4)	20 mM
	NaCl	500 mM
	Glycerol	10 %
	DTT	2 mM
	1xcomplete EDTA-free protease inhibitor	

Dialysis buffer:

	A	B
	HEPES (7.4)	20 mM
	NaCl	150 mM
	Glycerol	5 %
	MgCl ₂	2.5 mM
	DTT	2 mM
	GDP	20 μM



LRRK2 expression and purification

3h 14m

1 Transfect the constructs encoding 3xFlag-LRRK2, 3xFlag-LRRK2(I2020T), 3xFlag-RCKW or 3xFlag-GFP-LRRK2 into Expi293F cells according to manufacturer instructions.

2 Express the proteins for  72:00:00 following induction according to manufacturer instructions.

3d

3 Harvest the cells by centrifugation ( 400 x g ,  00:04:00) and lyse by 3 freeze-thaw cycles in lysis buffer.

4m

Note

Note: For  60 mL of cell suspension, we used  15 mL lysis buffer.

4 Remove the cellular debris by centrifugation at  15000 x g for  01:00:00 at  4 °C .

1h

5 Mix the clarified lysate with anti-FLAG M2 resin for  02:00:00 while rotating at  4 °C .

2h

Note

Note: For  60 mL of cell suspension, we used  180 µL of Anti-FLAG resin.

6 Wash the resin with 3×10 bed volumes of lysis buffer.



7 Elute the proteins with lysis buffer supplemented with [M] 0.2 mg/mL 3xFlag peptides.

Note

Note: For  60 mL of cell suspension, we used  800 µL elution buffer (without protease inhibitor).



- 8 Remove the N-terminal 3xFlag tag by incubation with the GST tagged Prescission Protease ( 0.01 U/ μ L)  Overnight while rotating at  4 °C . 
- 9 Remove the GST tagged Prescission Protease subsequently by Glutathione Sepharose.
- 10 Assess the purity of the proteins by SDS-PAGE and Western blotting.
- 11 Dialyze the purified proteins  Overnight at  4 °C against the dialysis buffer. 
- 12 After dialysis, clarify the proteins by centrifugation at  17000 x g for  00:10:00 at  4 °C . 10m 
- 13 Determine the protein concentration by SDS-PAGE using Bovine Serum Albumin (BSA) as standard and used without freezing in liposome binding and tubulation experiments.