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LR Clonase

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Invitrogen¹

¹Thermofisher



Robert Roden

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Protocol status: Working

We use this protocol and it's working

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- 1 Add the following components to a 1.5-mL microcentrifuge tube at room temperature and mix: • 1–7 μL entry clone (50–150 ng) • 1 μL destination vector (150 ng/ μL) • TE buffer pH 8.0, to 8 μL
- 2 Thaw on ice the LR Clonase™ II enzyme mix for about 2 minutes. Vortex the LR Clonase™ II enzyme mix briefly twice (2 seconds each time).
- 3 To each sample (step 1), add 2 μL of LR Clonase™ II enzyme mix to the reaction and mix well by vortexing briefly twice. Microcentrifuge briefly.
- 4 Return LR Clonase™ II enzyme mix to -20°C or -80°C storage.
- 5 Incubate reactions at 25°C for 1 hour. (overnight)
- 6 Add 1 μL of the Proteinase K solution to each sample to terminate the reaction. Vortex briefly. Incubate samples at 37°C for 10 minutes. Transform
- 7 Transform 1 μL of each LR reaction into 50 μL of One Shot™ OmniMAX™ 2 T1 Phage-Resistant Cells (Cat. no. C8540-03) (5 alpha e. coli). Incubate on ice for 30 minutes.

Follow electrotransformation protocol.
- 8
- 9 T1 Phage-Resistant Cells as described above. Plate 20 μL and 100 μL on LB plates containing 100 $\mu\text{g}/\text{mL}$ ampicillin. Expected results An efficient LR recombination reaction will produce >5000 colonies if the entire LR reaction is transformed and plated.