

Apr 24, 2019

Long staining procedure of nuclei in Euplotes crassus using DAPI

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DOI

<https://dx.doi.org/10.17504/protocols.io.2akgacw>

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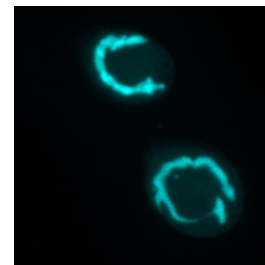
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Protocol Citation: Rachele Cesaroni 2019. Long staining procedure of nuclei in Euplotes crassus using DAPI. **protocols.io** <https://dx.doi.org/10.17504/protocols.io.2akgacw>



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Protocol status: Working

We use this protocol and it's working

Created: April 24, 2019

Last Modified: April 24, 2019

Protocol Integer ID: 22572

Keywords: staining procedure of nuclei, euplotes crassus, euplote, staining procedure, nuclei, staining, using dapi



- 1 Pellet *Euplotes crassus* cells at 400 rcf for 3 minutes, and remove as much supernatant as possible by pipetting.

Note

Both algae and bacteria are autofluorescent. Better to have a completely starved *Euplotes crassus* culture.

- 2 Add 1 ml of 2% PFA in 1X PHEM or 4% PFA in 1X PBS to the cells, and incubate them for 10 minutes at room temperature.
- 3 Pellet *Euplotes crassus* cells by centrifugation at 400 rcf for 3 minutes, and remove as much supernatant as possible by pipetting.
- 4 Wash cells twice with 1X PBS (400 rcf for 3 minutes each time).
- 5 Add 1 ml of TBSTEM - 3% BSA and 0.5 μ l of DAPI (0.1 mg/ml) to the cells, and stain for 10 minutes at room temperature.
- 6 Pellet *Euplotes crassus* cells by centrifugation at 400 rcf for 3 minutes.
- 7 Add 50 μ l of Prolong medium.
- 8 Place an approx. 10 μ l droplet of *Euplotes crassus* cells on a slide for observation by fluorescence microscopy.