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🌐 Long staining procedure of nuclei in Euplotes crassus using DAPI

🔗 Forked from [Long staining procedure of nuclei in Euplotes crassus using DAPI](#)

📁 In 1 collection

Rachele Cesaroni¹

¹Universität Bern

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Angela Piersanti
University of Camerino

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We use this protocol and it's working

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- 1 Pellet Euplotes crassus cells at 400 rcf for 3 minutes, and remove as much supernatant as possible by pipetting.

Note

Both algae and bacteria are autofluorescent. Better to have a completely starved *Euplotes crassus* culture.

- 2 Add 1 ml of 2% PFA in 1X PHEM or 4% PFA in 1X PBS to the cells, and incubate them for 10 minutes at room temperature.
- 3 Pellet *Euplotes crassus* cells by centrifugation at 400 rcf for 3 minutes, and remove as much supernatant as possible by pipetting.
- 4 Wash cells twice with 1X PBS (400 rcf for 3 minutes each time).
- 5 Add 1 ml of TBSTEM - 3% BSA and 0.5 µl of DAPI (0.1 mg/ml) to the cells, and stain for 10 minutes at room temperature.
- 6 Pellet *Euplotes crassus* cells by centrifugation at 400 rcf for 3 minutes.
- 7 Add 50 µl of Prolong medium.
- 8 Place an approx. 10 µl droplet of *Euplotes crassus* cells on a slide for observation by fluorescence microscopy.