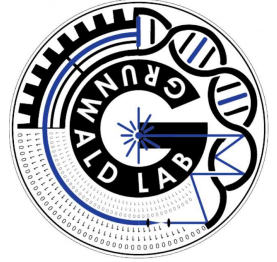


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Liquid Growth Medium - Yeast

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<https://dx.doi.org/10.17504/protocols.io.5qpvokr2bl4o/v1>



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Protocol status: Working

We use this protocol and it's working

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Abstract

This protocol describes the steps to prepare liquid culture medium for *Saccharomyces cerevisiae*.



Materials

SC-Ura Powder

Sunrise Science Products

Cat#: 1306-030

Lot#: 23K3083

Exp: 10/2027

Yeast Nitrogen Base Without Amino Acids

Sigma Life Science

Cat#: Y0626-250G

Lot#: SLBG0555V

Glucose

Sunrise Science Products

Cat#: 1907-1kg

Lot#: 3A0036

Deionized Water

Equipment:

500 ml laboratory bottle with screw cap

1ml pipette

50 ml pipette

stirring hot plate

magnetic stirring bar

micro scales

autoclave



Before start

Have the following solutions premixed:

Glucose 20% 500 ml solution:

Concentration: 200 g/l

mix 100 g Glucose in 500 ml deionized water (ddH₂O)

Optional:

SC-xx 10x 100ml solution:

Concentration: 19.2 g/l

mix 1.92 g into 100 ml ddH₂O

YNB 20x 100ml solution:

Concentration: 134.4 g/l

mix 13.44 g into 100 ml ddH₂O



1 Compound medium for autoclave

STEP CASE

Medium preparation with pre-resolved components

6 steps

This version of the protocol shows the preparation of the medium from SC-XX 10x and YNB 20x solutions.

- 1.1 Fill a 500 ml flask with  325 mL ddH₂O.

Add a magnetic stirring bar and place the flask on a stirring hot plate.

- 1.2 Add  25 mL YNB 20x solution (Yeast Nitrogen Base with Ammonium Sulfate without Amino Acids).

- 1.3 Add  100 mL SC-XX 10x solution.

Note

In regard to cover all optional dropout media the amino acid base holds the notification - xx, where xx stand for the amino acid(s) that is as selection factor, missing in the medium.

- 2 Autoclave for  00:15:00 at  121 °C .

Note

Remove the stirring bar before going to autoclave.

- 3 When the medium cooled down to around  80 °C add  50 mL sterile Glucose 20%.

4

Note

The medium can be store at the bench for 2 to 3 months.