

Oct 21, 2019

LAMP assay

DOI

dx.doi.org/10.17504/protocols.io.8jahuie

Niels Appelman¹

¹Wageningen University

iGEM Wageningen 2019



Niels Appelman

Wageningen University

OPEN  ACCESS



DOI: dx.doi.org/10.17504/protocols.io.8jahuie

Protocol Citation: Niels Appelman 2019. LAMP assay. [protocols.io https://dx.doi.org/10.17504/protocols.io.8jahuie](https://dx.doi.org/10.17504/protocols.io.8jahuie)

License: This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: October 21, 2019

Last Modified: October 21, 2019

Protocol Integer ID: 28994

Abstract

This is a protocol used to perform LAMP assays. It was adapted to

<https://international.neb.com/protocols/2016/08/15/warmstart-colorimetric-lamp-2x-master-mix-typical-lamp-protocol-m1800>

- 1 Prepare a 10X primermix according to the following scheme:

Primer	Concentration
FIP	16 uM
BIP	16 uM
F3	2 uM
B3	2 uM
LOOP F	8 uM
LOOP B	8 uM

- 2 Add the following components to a microcentrifuge tube

Component	Volume added (25 uL reaction)	Volume added (10 uL reaction)
Mast ermix	12.5 uL	5 uL
Lamp primers (10X)	2.5 uL	1 uL
Temp late	x uL	x uL
MilliQ	10 – x uL	4 – x uL

Add a no-template control to ensure template-specificity

- 3 Incubate at 65 C for 30 minutes.
- 4 Record the color of all samples. Yellowing of a sample indicates a positive result.

- 5 In case of doubt regarding the result of a sample, run it on an agarose gel. The presence of many bands, often not individually recognizable, indicates a positive result.