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iPSC PCR: For Screening Edited Clones



In 1 collection

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Protocol status: Working

We use this protocol and it's working

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Attachments



Guidelines

This protocols is part of the **Screening Edited iPSC Clones collection**.

Safety warnings



Please refer to the SDS (Safety Data Sheet) for information about hazards, and to obtain advice on safety precautions.

Before start

Using the extracted gDNA, set up the following PCR and run using the following conditions.

Each cell line/mutation will require its own unique set of primers. Ensure that you have the appropriate primers for the cell line/mutation before setting up the PCR.

Note: Design primers to amplify approximately a 300-400 bp region for best results.



PCR

1 This is the set up for a 50 μ L reaction.

	Volume	x# rxns
5x Green GoTaq Flexi Buffer	10 μΙ	
25mM MgCl2	6 μΙ	
25mM dNTPs	0.8 μΙ	
Forward Primer (10uM)	2 μΙ	
Reverse Primer (10uM)	2 μΙ	
GoTaq DNA Polymerase (5U/uL)	0.25 μΙ	
Milli-Q H20	26.95 μΙ	
Quick Extract gDNA	2 μΙ	
Total	50 μΙ	

2

3 After PCR has been set up, run in thermocycler using the following conditions.

Segme nt	Cycle s	Temperature	Time
1	1	95°C	2 minutes
		95°C	30 seconds
2	40	65°C - 1°C/cycle	30 seconds
		72°C	1 minute
3	1	72°C	5 minutes
4	1	4°C	Forever

4 After PCR has been run in thermocycler, it is best to run the sample on a 2% gel to confirm the presence of PCR product.

Gel preparation

5 To make the gel, combine an appropriate amount of Agarose, TBE and Ethidium Bromide using the following guidelines.



a. The 2% gel will be cast in one of the following ways:

	15×15 cast	15×25 cast
Agarose	1.5 g	3.0 g
TBE	75 mL	150 mL
Ethidium Bromide	3.75 uL	7.5 uL

- b. Combine Agarose and TBE in an appropriately sized flask and microwave until Agarose is completely dissolved. Swirl intermittently during heating.
- c. Once completely dissolved add appropriate amount of Ethidium Bromide to flask and swirl until dispersed evenly.
- Pour gel from flask into casting tray (be sure to add appropriate amount of combs to casting tray).
- 7 Let sit for 30-40 minutes, or until firm. 👏 00:30:00
- 8 Place gel cast into the gel rig apparatus.
- 9 Load samples.

Note

Be sure to include positive and negative controls when possible.

- 10 Load 50 bp ladder.
- 11 Place lid on gel rig apparatus.
- Run gel at 150 volts for 01:30:00 (checking at 01:00:00 to ensure samples have not run too far or off the gel).
- 13 Turn off gel rig apparatus and remove cast.



- 14 Blot off excess TBE from cast.
- 15 Visualize gel
- 16 If PCR product is the approrpiate size and concentration as indicated by the gel, use the remaining PCR product to perform restriction digest