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## iPSC Freezing

 In 1 collection

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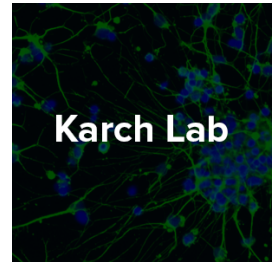
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**Protocol status:** Working

**We use this protocol and it's working**

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**Protocol Integer ID:** 20473

**Keywords:** ipsc freezing, ipsc

## Guidelines

This protocols is part of the [Screening Edited iPSC Clones collection](#).

## Materials

### STEP MATERIALS

 Accutase™ Cell Dissociation Reagent **Gibco - Thermo Fisher Scientific Catalog #A1110501**


## Protocol materials

 Accutase™ Cell Dissociation Reagent **Gibco - Thermo Fisher Scientific Catalog #A1110501**

 Accutase™ Cell Dissociation Reagent **Gibco - Thermo Fisher Scientific Catalog #A1110501**

## Troubleshooting

## Safety warnings

 Please refer to the SDS (Safety Data Sheet) for information about hazards, and to obtain advice on safety precautions.





1 Aspirate media

2 Gently wash cells with 1x PBS.

Note

Use 2-3 mL per well in 6 well plate.

3 Add Accutase (Gibco A11105-01) directly to the cells and incubate at  37 °C for 3-4 minutes.  00:03:00

Note

Individual donor cell lines exhibit variable sensitivity to accutase-mediated dissociation. Thus, monitor cells closely to determine when single cell dissociation is achieved.

Note

For a 6 well plate, add 0.75-1 mL per well.  
For a 10cm<sup>2</sup> dish, add 3 mL.









Accutase™ Cell Dissociation Reagent **Gibco - Thermo Fisher**  
**Scientific Catalog #A1110501**

4 Tap dish to aid in dislocation of cells.

5 Add DMEM/F12 directly to cells.

**Note**

- For a 6 well plate, add  2 mL -  4 mL per well.
- For a 10cm<sup>2</sup> dish, add  9 mL
- If cells remain attached, use a cell scraper to gently dislodge cells (apply gentle pressure and use 1-2 passes to remove cells)

- 6 Collect cells in conical tube (15mL/50mL depending on volume).
- 7 Add  2 mL -  5 mL DMEM/F12 to dish to remove all cells from the dish and add to conical tube.
- 8 Centrifuge cells at 750 rpm for  00:03:00 at room temperature.
- 9 Carefully aspirate supernatant.

**Note**



To avoid aspirating cell pellet, it is OK to leave a small amount of media (0.5-1mL).

- 10 Resuspend cell pellet with mTesR1 (No Rock Inhibitor).
  - Use volume appropriate for freezing.
  - Assume 1 mL per cryovial total and add ½ total volume of mTesR1.
  - Pipet cells 1-2 times only to preserve cell clumps.

**Note**

**Example:** to freeze 10 tubes, you will need 10 mL total and will add 5 mL mTesR1 to cell pellet (and 5 mL of 2x Freezing Media below).



- 11 Add an equal volume of cold 2x Freezing Media (20% DMSO, FBS). Pipet cells 1 time only to preserve cell clumps.
- 12 Transfer cell suspension to pre-labeled cryovials (1 mL per cryovial).
  - Ensure that cryovials are labeled with the following
    - Cell Type
    - Line Name
    - Passage #
    - Date
    - Your Name
- 13 Freeze vials at  -80 °C in foam racks for 48-72 hours.  48:00:00
- 14 Transfer vials to liquid nitrogen for long-term storage.