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iPSC derived microglia (hMG) culture

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We use this protocol and it's working

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Abstract

iPSC derived microglia (hMG) culture

- 1 Single iPSC colonies are manually passaged onto fresh Matrigel-coated 6 cm² plastic plate.
- 2 After 2-4 days, protocol starts by supplementing mTeSRTM1 medium with 80 ng/mL of Bone Morphogenetic Protein (BMP)-4 every day from day 0 to day 4.
- 3 From day 5, SP34 medium is employed, which consisted of StemProTM-34 SFM (Gibco **TM**) with 1% of P/S and 1% of Ultraglutamine (Glut; Lonza). For two days, SP34 medium is supplemented with 80 ng/ml of VEGF, 100 ng/ml of Stem Cell Factor (SCF) and 25 ng/ml of Fibroblast Growth Factor (FGF)-2.
- 4 After day 7 until day 14, SP34 is supplemented with 50 ng/ml of Fms-like tyrosine kinase 3-Ligand (Flt3-L), 50 ng/ml of IL-3, 50 ng/ml of SCF, 5 ng/ml of Trombopoietin (TPO) and 50 ng/ml of M-CSF, changing the medium at day 10.
- 5 The last step consisted on the addition to SP34 of Flt3-L, M-CSF and 25 ng/ml of GM-CSF from day 14, changing the medium every 3-4 days. At day 35, floating hMG progenitors are collected from the culture's supernatant and passed through a 70 µm Filcon**TM** Syringe-Type nylon mesh (BD Biosciences) to ensure a single-cell suspension and to remove cell clumps.
- 6 Cells are counted, centrifuged at 300xg for 10 minutes and cultured in Roswell Park Memorial Institute (RPMI) 1640 Medium (Gibco**TM**) with 1% of P/S and 1% of Glut, supplemented by M-CSF and 50 ng/ml IL-34.
- 7 Medium is changed with fresh factors every 2-3 days. hMG progenitors are plated onto Matrigel-coated glass coverslips in 24-well plates (25,000 cells/well), on uncoated 96-well plates (10,000 cells/well) or 6-well plates (200,000 cells/well) depending on the experiment.
- 8 hMG mature cells are cultured for one week, before being used for experiments. For treatments, 100 ng/mL of LPS or 5 ug/mL of NM are added for a total of 24 hours.