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Intracellular and extracellular MHC-II immunostaining and microscopy on peritoneal macrophages

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We use this protocol and it's working



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Abstract

Staining of intracellular and extracellular MHCII in peritoneal macrophages in 96 well plate using EVOS fluorescent microscope and analysis on CellProfiler

Troubleshooting



- After harvesting and plating pMacs from mouse, plate in 96-well plate at 50,000 cells/well in 200uL as described in pMac harvesing protocol. Once cells have adhered, aspirate media and replace with fresh growth media containing vehicle or 100U of IFNy for 18
- 2 Retrieve cells from 37 degrees. Aspirate media and wash 3x DPBS+/+
- Incubate cells with 25 ug/mL of APC-MHC-II (Biolegend, RRID:AB_313329) in DPBS+/+ containing 1:100 FcR blocking reagent, final volume 50uL, for 30 minutes at room temperature, protected from light
- 4 Wash cells 3 x DPBS+/+
- fix cells by incubation in 4% PFA for 10 minutes at room temperature, final volume of PFA 50uL, and then washed 3x with DPBS+/+
- Permeabilize cells with 100uL permeabilization buffer (eBiosciences, #88-8824-00) on ice for 15 minutes
- Spike in 25ug/mL of PE-610-MHC-II (Biolegend, RRID:AB_2574618) into permeabilization buffer and incubate cells for 30 minutes at room temperature protected from light.
- 8 Wash cells 3 x DPBS+/+
- Incubate cells in 1 μ g/ml DAPI (Invitrogen, RRID:AB_2629482) for 10 minutes at room temperature in DPBS+/+, final volume 50uL.
- 10 Image cells on an EVOSTM M7000 (Invitrogen) at 20 x magnification.
- Perform image analysis using Cellprofiler 4.2.5 (RRID:SCR_007358). Using the 'IdentifyPrimaryObject' module in Cellprofiler, identify icMHC and exMHC in their respective channels and quantify MFI and calculate Ex:IcMHCII from these quantified MFI values.