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In vivo CAR T cell tumor control assay

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Andrea R Daniel: This protocol is adapted from the M. Brown lab at Duke University.

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We use this protocol and it's working

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Abstract

This protocol describes methods for a HCC1954 cell implanted orthotopic tumor model in mice with delivery of T cells expressing HER2 CAR. Tumor volumes are measured to evaluate tumor control by CAR T cell therapy.

Troubleshooting



HCC1954 orthotopic tumor model

- Six- to 8-week-old female immunodeficient NOD/SCID gamma (NSG) mice were obtained from Jackson Laboratory and then housed in 12-h light/dark cycles, at an ambient temperature (21 ± 3 °C) with relative humidity (50 ± 20 %) and handled in pathogen-free conditions
- 2 HCC1954s were maintained in DMEM/F12 supplemented with 10% FBS, 100 U ml^{-1} penicillin and 100 $\mu g ml^{-1}$ streptomycin.
- Resuspend cells in 2ml PBS (5×10⁷ cells/ml), then diluted 1:1 with Matrigel (2.5e7/ml) thus 2.5e6 per 100ul.
- 4 2.5×10^6 HCC1954 cells are implanted orthotopically into the mammary fat pad of NSG mice in 100µL volume of 50:50 (v:v) PBS:Matrigel.
- Upon detection tumor volumes were calculated based on caliper measurements using the formula volume = $\frac{1}{2}$ (Length × Width²). Measure tumors every 4-6 days.

CAR T cell delivery

- Human donor T cells were expanded for 9-11 days post-transduction with HER2-CAR-2A-GFP and HER2-CAR-2A-BATF3 constructs before delivery to tumor bearing mice.
- 7 Transduction rates were measured on the day of treatment using flow cytometry and transduction rates exceed 80%.
- T cells were resuspended at 50×10^6 CAR+ cells mL⁻¹ in 1X PBS and serially diluted to the appropriate cell concentrations for 200 μ L injections of either 10×10^6 , 2×10^6 , 5×10^5 , 2.5×10^5 , or 1×10^5 HER2 CAR+ T cells.
- 9 21 days after tumor implantation randomize mice into groups for CAR T cell injections. Deliver CAR T cells intravenously by tail vein injection.
- 10 Measure the length and width of tumors every 4-6 days using calipers.



11 Mice are euthanized before reaching a tumor volume of 2,000 mm³, the upper threshold defined by the Duke Institutional Animal Care and Use Committee.