

Immunohistochemistry Protocol for Sternberger Monoclonal Antibodies V.2

DOI

dx.doi.org/10.17504/protocols.io.tkkekuw



Kelsey Miller¹

¹BioLegend

BioLegend

Tech. support email: tech@biolegend.com



Sam Li

BioLegend



DOI: dx.doi.org/10.17504/protocols.io.tkkekuw

External link: https://www.biolegend.com/protocols/immunohistochemistry-protocol-for-sternberger-monoclonal-antibodies/4253/

Protocol Citation: Kelsey Miller . Immunohistochemistry Protocol for Sternberger Monoclonal Antibodies. **protocols.io** https://dx.doi.org/10.17504/protocols.io.tkkekuw

License: This is an open access protocol distributed under the terms of the **Creative Commons Attribution License**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Created: September 14, 2018

Last Modified: August 29, 2019

Protocol Integer ID: 15724

Keywords: IHC

Guidelines

Use with Ultra Streptavidin Detection Kit (SIG-32250) or (SIG-32248)

Positive control: Normal human cerebellum (except SMI-71, which should be rat brain)



Clear Slides

1 Clear Slides: Removes paraffin and hydrates the tissue.

Xylene	5 minutes in each of (3) different 250mL containers
100% alcohol	5 minutes in each of (3) different 250mL containers
95% alcohol	3 minutes in (1) 250mL container
70% alcohol	3 minutes in (1) 250mL container
water	1 minutes in each of (3) different 250mL containers
H2O2 (3%)	15 minutes in (1) 250mL container

Rinse slides

2 Rinse slides with lab grade water.

> Note: Lab grade filtered water such as injection grade, cell culture grade, Reverse Osmosis De-Ionization (RODI)

Antigen Retrieval

- 3 Heat slides in 1X Retrieve ALL3 solution for 1 minute 40 seconds on high power in microwave
 - **(:)** 00:01:40
- 4 Reduce to low power and simmer 10 minutes in microwave.
 - **(:)** 00:10:00
- 5 Remove from microwave and allow slides to cool on the bench top for 10 minutes.
 - **(5)** 00:10:00
- 6 Rinse slides with lab grade water.
- 7 Apply serum block for at least 5 minutes. Do NOT wash after this step.
 - **(5)** 00:05:00
- 8 Blot off serum block.

- 9 Apply primary antibody (see recommended dilution from datasheet).
- 10 Incubate primary antibody 60 minutes at room temperature.

(:) 01:00:00

- 11 Rinse slides with 1X PBS.
- 12 Apply USA Linking reagent - 20 minutes incubation. (If using a biotinylated product, omit this step)

00:20:00

- 13 Rinse slides with 1X PBS.
- 14 Apply Labeling Reagent - 20 minutes incubation. 00:20:00
- 15 Rinse with 1X PBS.
- 16 Apply chromogen - 5 minutes incubation. Dilute according to manufacturer's instructions.

(?) 00:05:00

- 1. AEC Chromogen: 20µL AEC chromogen + 1mL AEC substrate buffer
- 2. DAB Chromogen: 40µL DAB chromogen + 1mL DAB substrate buffer

Counterstain

17 Submerge slides in Mayer's Hematoxylin for 30 seconds.

(:) 00:00:30

18 Rinse under running lab grade water for 1 minute or until water is clear.

00:01:00

19 Submerge slides in Bluing Reagent for 1 minute.

00:01:00

20 Rinse under running lab grade water for 1 minute.

(5) 00:01:00



Clear slides

- 21 Clear slides: Dehydrate the tissue.
 - 1. 95% alcohol 3 minutes in (1) 250mL container
 - 2. 100% alcohol 5 minutes in each of (3) different 250mL container
 - 3. Xylene 5 minutes in each of (3) different 250mL container

Coverslip

22 Cover slip slide using Permanent Aqueous Mounting Medium (SIG-31010).

Note: Do not use xylene based mount with AEC Chromogen as it will dissolve the chromogen