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# Immunohistochemical labelling of spinal cord neurons involved in bladder activity



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Protocol status: Working

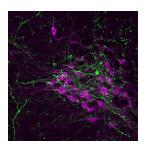
We use this protocol and it's working

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**Keywords:** activity-mapping, neuroanatomy, immunohistochemistry





## **Abstract**

This protocol is used for immunohistochemical visualisation of immediate early gene expression (c-Fos or Egr-1) in cryosections of rat lumbosacral spinal cord. Free-floating sections are processed in a double labelling protocol to distinguish immediate early gene expression in different neurochemical classes of spinal cord neurons:

- ChAT [choline acetyltransferase]: preganglionic neurons
- TH [tyrosine hydroxyls]: dopaminergic neurons
- Pax2: inhibitory interneurons



## **Materials**

#### **MATERIALS**

- Horse serum Merck MilliporeSigma (Sigma-Aldrich) Catalog #12449C
- OCT (Optimal Cutting Temperature compound) Sakura Finetek Catalog #4583
- X Cy3 Donkey anti-goat IgG Jackson ImmunoResearch Laboratories, Inc. Catalog #705-165-147
- X AF488 Donkey anti-rabbit IgG Jackson ImmunoResearch Laboratories, Inc. Catalog #711-545-152
- NeuroTrace™ 640/660 Deep-Red Fluorescent Nissl Stain Solution in DMSO Thermo Fisher Catalog #N21483
- Mouse anti-cFos antibody Santa Cruz Biotechnology Catalog #sc166940
- Rabbit anti-Pax2 antibody Invitrogen Thermo Fisher Catalog #71-6000
- Rabbit anti-Egr-1 (588) antibody Santa Cruz Biotechnology Catalog #sc-110
- Rabbit anti-TH antibody Merck Millipore (EMD Millipore) Catalog #AB152
- X Cy3 Donkey anti-mouse IgG Jackson ImmunoResearch Laboratories, Inc. Catalog #715-165-150
- Soat anti-ChAT antibody Merck Millipore (EMD Millipore) Catalog #AB144P

#### Solutions:

- PBS: phosphate-buffered saline, 0.1 M, pH 7,2
- PBS containing 0.1% sodium azide
- PBS containing 30% sucrose (w/v)
- Blocking solution: PBS containing 10% normal horse serum and 0.5% triton X-100
- PBS containing 0.1% sodium azide, 2% normal horse serum and 0.5% triton X-100

## **Primary Antibodies:**

Abbr eviati on	Gene name	Syno nym	RRID	Host Speci es	Diluti on
cFos	Fos	cFos	AB_10 6096 34	Mous e	1:100
ChAT	Chat	Choli ne acety Itrans feras e	AB_2 0797 51	Goat	1:500



Egr-1/ Zif26 8	Egr1/ Zif26 8	Early growt h recep tor 1	AB_2 09717 4	Rabbi t	1:500 0
Pax2	Pax2	Paire d box gene 2	AB_2 5339 90	Rabbi t	1:100 0
тн	Th	Tyros ine hydro xylas e	AB_3 9020 4	Rabbi t	1:200

# Secondary Antibodies:

Tag- antib ody	Host speci es	Diluti on
Cy3 anti- mous e	Donk ey	1:200 0
AF48 8 anti- rabbit	Donk ey	1:100 0
Cy3 anti- goat	Donk ey	1:100 0



## **Preparation of cryosections**

- 1 Cryoprotect fixed tissue (L5-S2 spinal cord) in phosphate-buffered saline (PBS; 0.1 M, pH7.2) containing 30% sucrose. This should be performed at 4C, 24-72h prior to cutting.
- 2 Embed tissue in cryomold using OCT, freeze in cryostat and cut sections (40 μm), collecting sections progressively across sets of 4 wells to collect 160 μm spaced series.

## **Immunostaining**

- 3 Wash sections in PBS ( $3 \times 10 \text{ min}$ )
- 4 Incubate sections in blocking solution at room temperature for 2 h
- Incubate sections in appropriate dilutions of primary antibodies (or combinations of primary antibodies) for 48-72h. Antibodies are diluted in PBS containing 0.1% sodium azide, 2% horse serum, and 0.5% triton-X.
- 6 Wash sections in PBS ( $3 \times 10 \text{ min}$ )
- Incubate sections in appropriate dilutions of secondary antibodies (or combinations of secondary antibodies) 4 h in the dark. Antibodies are diluted in PBS containing 2% horse serum, and 0.5% triton-X.

#### Note

A useful counterstain to visualise spinal regions can be included here, by adding NeuroTrace (fluorescent Nissl stain; 1:100) to the secondary antibody incubation.

- 8 Wash sections in PBS  $(3 \times 10 \text{ min})$
- 9 Mount sections onto glass slides and coverslip in preferred anti-fade mountant.



# Microscope

10 Labeled neurons are counted and classified according to their immunoreactivity, including only nucleated neuronal profiles in the analysis.

## Note

For digital analysis, tile-scanning of complete spinal cord sections is recommended, ensuring that the order of sections (rostral to caudal) is noted.