

Mar 01, 2022

Immunofluorescent Staining

DOI

dx.doi.org/10.17504/protocols.io.b5s5q6g6

Haley Geertsma¹

¹University of Ottawa



Haley Geertsma

University of Ottawa

OPEN  ACCESS



DOI: dx.doi.org/10.17504/protocols.io.b5s5q6g6

Protocol Citation: Haley Geertsma 2022. Immunofluorescent Staining. **protocols.io**

<https://dx.doi.org/10.17504/protocols.io.b5s5q6g6>

License: This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: March 01, 2022

Last Modified: March 01, 2022

Protocol Integer ID: 58941

Abstract

This protocol is used to stain cryosectioned mouse brain tissue.



- 1 To cryo-sectioned brain tissue, wash with 1X phosphate buffered saline (PBS) for 3× 5-minute washes. 15m
- 2 Incubate in blocking buffer for 1 hour at room temperature.
Blocking buffer: 10% serum + 0.5% Triton X-100 in 1X PBS 1h
- 3 Wash tissue with 1X PBS. 5m
- 4 Incubate in primary antibody diluted in blocking buffer overnight at 4°C. 1d
- 5 Wash tissue with 1X PBS for 5× 5-minute washes. 30m
- 6 Incubate in secondary antibody diluted in blocking buffer for 1 hour at room temperature. 1h
- 7 Wash tissue with 1X PBS for 5× 5-minute washes. 30m
- 8 If tissue wasn't previously mounted on a slide, mount on a superfrost plus slide and let dry at room temperature for at least 10 minutes. 15m
- 9 Coverslip with fluorescent mounting medium and a #1.5 coverslip. Outline the coverslip with clear nailpolish. 1m