

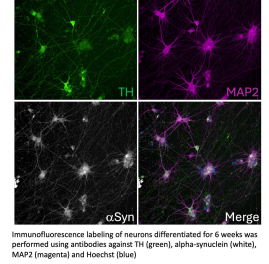


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Immunofluorescence Labeling of TH, Alpha-Synuclein, and MAP2 in iPSC-Derived Dopaminergic Neurons

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Protocol status: Working

We use this protocol and it is working

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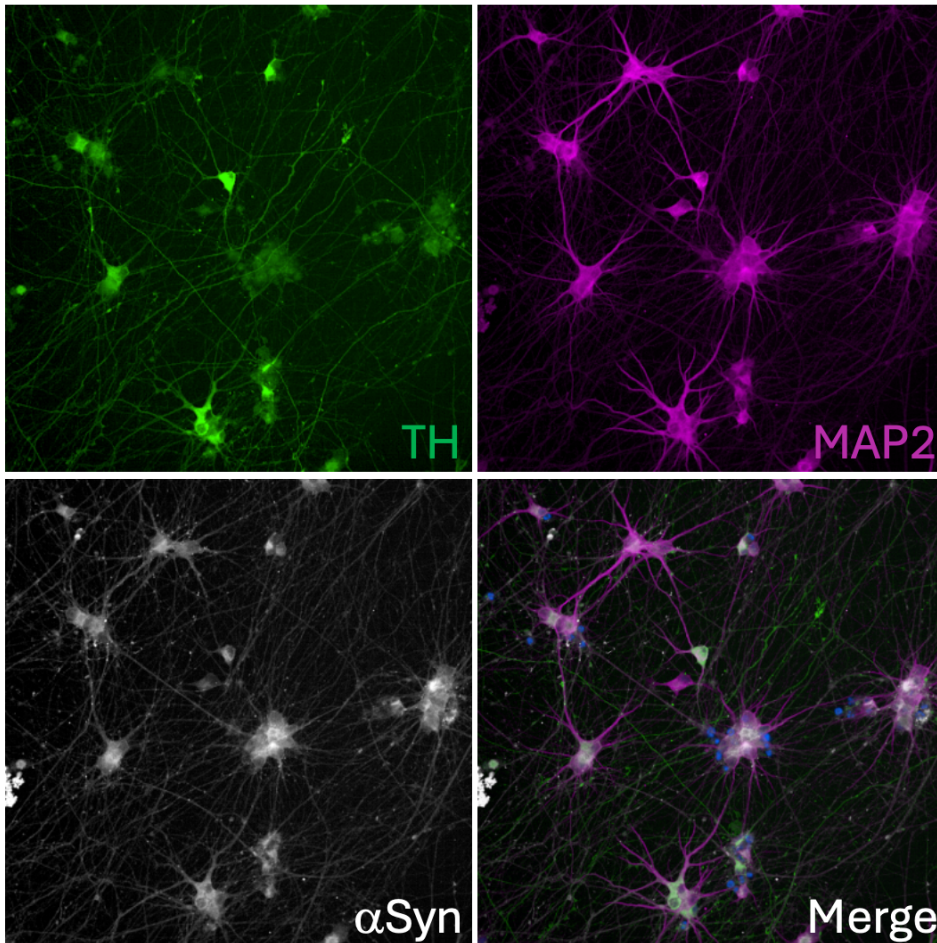
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Abstract

This protocol outlines the procedure for labeling tyrosine hydroxylase (TH), alpha-synuclein (α -syn), and microtubule-associated protein 2 (MAP2) in induced pluripotent stem cell (iPSC)-derived dopaminergic neurons using immunofluorescence.



Immunofluorescence labeling of neurons differentiated for 6 weeks was performed using antibodies against TH (green), alpha-synuclein (white), MAP2 (magenta) and Hoechst (blue)



Materials

Reagents	Company	Cat. Number
Rabbit Tyrosine hydroxylase antibody	Pel-Freez	P40101
Mouse alpha-synuclein antibody	BD Biosciences	610787
Chicken MAP2 antibody	Encor Biotech	CPCA-MAP2
Goat anti-Chicken IgY (H+L) Secondary Antibody, Alexa Fluor 555	ThermoFisher Scientific	A21437
Goat anti-Mouse IgG (H+L) Cross-Adsorbed Secondary Antibody, Alexa Fluor 647	ThermoFisher Scientific	A21235
Goat anti-Rabbit IgG (H+L) Cross-Adsorbed Secondary Antibody, Alexa Fluor 488	ThermoFisher Scientific	A11008
Hoechst 33342	SIGMA	H3570
16% Formaldehyde solution (w/v) Methanol-free	ThermoScientific	28908
Normal goat serum	SIGMA	G9023
Triton X-100	SIGMA	X-100

Before start

iPSC-derived dopaminergic neurons are cultured on 96-well plates for up to 4 to 6 weeks of differentiation. We use black Costar 96-well assay plates (Corning cat. no. 3904), which are compatible with imaging using the Opera Phenix high-content screening system.

Be extremely gentle when dispensing into the wells, as dopaminergic neurons are prone to detaching with forceful pipetting.



Fixation of dopaminergic neurons in 96 well plates

40m

- 1
 - Leave  50 μL of media and gently add  50 μL of 8% PFA per well.
 - Incubate at  Room temperature  00:20:00 .
 - Remove PFA and wash 3 times with 1X PBS,  75 μL per well,  00:05:00 each

40m

Permeabilization

25m

- 2
 - Prepare permeabilization solution: 0.2% TX-100; 1X PBS.
 - Incubate in permeabilization solution  00:10:00 ,  60 μL per well
 - Remove permeabilization solution and wash 3 times with 1X PBS,  75 μL per well,  00:05:00 each

25m

Blocking

1h

- 3
 - Prepare blocking buffer: 5% normal goat serum; 0.02% TX-100; 1X PBS
 - Incubate in blocking solution  01:00:00 ,  60 μL per well

1h

Incubation with primary antibodies

16h

- 4
 - Prepare primary antibody mix in blocking buffer,  60 μL per well
 - rabbit TH antibody 1:500
 - mouse alpha-synuclein antibody 1:500
 - chicken MAP2 antibody 1:1500
 - Incubate  Overnight at  4 $^{\circ}\text{C}$ with gentle shaking

16h

Washes

20m

- 5
 - Wash 4 times with  75 μL 1X PBS,  00:05:00 each

20m



Incubation with secondary antibodies

2h

- 6 - Prepare secondary antibody mix in 1% normal goat serum; 1X PBS. Dispense  60 μL per well.

2h

- anti-Rabbit Alexa Fluor 488 1:1000
- anti-mouse Alexa Fluor 647 1:1000
- anti-Chicken Alexa Fluor 555 1:1000
- Hoechst 33342 1:5000

- Incubate  02:00:00 at  Room temperature with gentle shaking

Washes

20m

- 7 - Wash 4 times with  75 μL 1X PBS,  00:05:00 each
- Leave  100 μL of 1X PBS into wells

20m

Imaging

- 8 - Samples are ready for imaging