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## Immunofluorescence in PFA-fixed grafted mouse brain slices

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SOX6 mDA differentiation



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**Protocol status:** Working

**We use this protocol and it's working**

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**Keywords:** immunohistochemistry, immunofluorescence, animal tissue, immunofluorescence in pfa, grafted mouse brain slice, fixed grafted mouse brain slice,  $\mu\text{m}$  mouse brain slice, conjugated antibody, immunofluorescence, double labeling of protein, fixed brain slice, protein expression, protein, brain slice, pfa, succesful on pfa, using fluorophore

## Abstract

This protocol was used to tag protein expression using fluorophore-conjugated antibodies on 20  $\mu\text{m}$  mouse brain slices. Double labeling of proteins and nuclei was succesful on PFA-fixed brain slices.

## Materials

PBS1X

PBTx0.3% [Triton-X 0.3% in PBS]

PBTx0.1% [Triton-X 0.1% in PBS]

Blocking Serum Solution (BSS) [Donkey Serum 5%, BSA 1 mg/ml, PBTx0.1%]

Primary antibodies

Secondary antibodies

ReadyProbes Autofluorescence quenching agent

DAPI

Mounting medium

## Protocol materials

 Fluoromount **Merck MilliporeSigma (Sigma-Aldrich) Catalog #F4680**

## Troubleshooting



## Day one: Primary antibody

19h 35m

- 1 Place up to eight  $\rightarrow$   $\pm$  20-30  $\mu\text{m}$  slices in wells filled with PBS1X.  
*For free-floating, 12 well plates were used and slow constant shaking to ensure coating without compromising sample structural integrity.*
- 2 Wash tissue with PBS1X.  
PBS1X can be commercial or self-made, but make sure that it is  $\text{pH } 7.4$   
  

- 3 Wash tissue with PBS1X  
  

- 4 Wash with PBTx0.3%  
  

- 5 Wash with PBTx0.1%  
  

- 6 Wash with PBTx0.1%  
  

- 7 Incubate samples in BSS [Donkey Serum 5%, BSA 1 mg/ml, PBTx0.1%], preferably in plates of smaller wells (e.g 24 well) to optimize volume used.  
*Blocking solution was prepared for secondary (2ndary) antibodies hosted in donkey. If 2ndary antibody is hosted in goat, add Goat serum 5%. Remember to not use donkey or goat serum if the primary antibody is hosted on any of those serums.*  
  

- 8 Incubate samples in primary antibody solution (in BSS) at  $4\text{ }^{\circ}\text{C}$   
*Always remember to check antibody host. Use recommended dilution, if no recommendations are there for immunohistochemistry, rule of thumb is 5 x recommendation of immunocytochemistry.*  
  
  
  


## Day two: Secondary antibody

2h 50m

- 9 Recover slices to 12 well plates. Wash with PBTx0.1%  
  


- 10 Wash with PBTx0.1% 5m  

- 11 Incubate in secondary solution (in BSS) 🌡️ Room temperature 2h  

- 12 Wash tissue with PBS1X 5m
- 13 Wash tissue with PBS1X 5m
- 14 Optional: Incubate samples in ReadyProbes Tissue Autofluorescence quencher (Thermo Scientific, R37630). 10m  
\*
- 15 Wash with PBS1X 5m  
\*
- 16 Incubate in DAPI:PBS 1:1000 solution 10m
- 17 Wash with PBS1X 5m
- 18 Mount on glass slides with Fluorescent Mounting Media  
🧪 Fluoromount Merck MilliporeSigma (Sigma-Aldrich) Catalog #F4680