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Immunoblotting of macrophages and microglia

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Protocol status: Working

We use this protocol and it's working

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Abstract

This protocol describes the preparation from cell lysate from cultured cells and immunoblotting procedure.

Attachments



[mqjwbj3up.docx](#)

17KB



Materials

Solutions to prepare

RIPA buffer:

	A	B
	NaCl	150 mM
	Tris	10 mM
	EDTA	0.5 mM
	0.5% NP40	

supplemented immediately before use with Protease Inhibitor Cocktail (Roche) and PhosStop phosphatase inhibitor (Roche).

TBS:

	A	B
	Tris-Cl	50 mM
	NaCl	150 mM

adjust pH to 7.5

TBST: TBS with 0.1% TWEEN-20 (Sigma-Aldrich)

4x Laemmli buffer:

	A	B
	Tris-Cl	188 mM
	3% SDS	
	30% glycerol	
	0.01% bromophenol blue	
	15% β -mercaptoethanol	

Tris-glycine running buffer:

	A	B
	tris base	25 mM
	glycine	192 mM
	0.1% SDS in milliQ milliQ water	

Tris-glycine transfer buffer:

	A	B
	Tris	25 mM
	glycine	192 mM
	20% methanol in milliQ water	

Chill to 4 °C .

Antibody dilutions:

Primary antibodies:

GAPDH, 1:10000 dilution (EnCor biotechnology Inc, # MCA-1D4), human specific cathepsin D, 1:4000 dilution (R&D systems #AF1014), mouse specific cathepsin D, 1:5000 dilution (R&D systems #AF1061), human cathepsin B, 1:1000 dilution (R&D systems #AF953), human cathepsin L, 1:10000 dilution (R&D systems # AF952), mouse cathepsin L, 1:2000 dilution (R&D systems # AF1515), human cathepsin C, 1:1000 dilution (R&D systems #AF1071), human GBA, 1:1000 dilution (R&D systems #MAB7410), hLAMP1, 1:2000 dilution (Cell signaling Technology # 9091), mouse LAMP1, 1:2000 dilution (DSHB #ID4B), MITF, 1:1000 dilution (Cell signaling Technology # 125903), MITF, 1:1000 dilution (Abcam #Ab303530), S6K, 1:1000 dilution (Cell signaling Technology #9202), S6K-Phospho-T389, 1:1000 dilution (Cell signaling Technology #9205), human TFEB, 1:1000 dilution (Cell signaling Technology #37785), mouse TFEB, 1:1000 dilution (Proteintech #13372-1-AP), TFE3, 1:1000 dilution (Sigma #HPA023881).

Secondary antibodies:

All the HRP tagged secondary antibodies are purchased from Cell signaling Technology and diluted at 1:2000 ratio.

Human Cathepsin D Antibody **R&D Systems Catalog #AF1014**

Mouse Monoclonal Antibody to GAPDH **Encor Biotechnology Catalog #MCA-1D4**



⊗ Human Cathepsin B Antibody **R&D Systems Catalog #AF953**

⊗ Human Cathepsin L Antibody **R&D Systems Catalog #AF952**

⊗ Mouse/Rat Cathepsin L Antibody **R&D Systems Catalog #AF1515**

⊗ Human Cathepsin C/DPPI Antibody **R&D Systems Catalog #AF1071**

⊗ Human Glucosylceramidase/GBA Antibody **R&D Systems Catalog #MAB7410**

⊗ MITF (D5G7V) Rabbit mAb **Cell Signaling Technology Catalog #12590**

⊗ Recombinant Anti-MiTF antibody **Abcam Catalog #ab303530**

⊗ p70 S6 Kinase Antibody **Cell Signaling Technology Catalog #9202**

⊗ Phospho-p70 S6 Kinase (Thr389) Antibody **Cell Signaling Technology Catalog #9205**

⊗ TFEB (D2O7D) Rabbit mAb **Cell Signaling Technology Catalog #37785**









⊗ TFEB Polyclonal antibody **Proteintech Catalog #13372-1-AP**

⊗ Anti-TFE3 antibody produced in rabbit **Merck MilliporeSigma (Sigma-Aldrich) Catalog #HPA023881**

Troubleshooting







Cell culture and treatments










- 1 Supplement RIPA buffer with Protease Inhibitor Cocktail (Roche) and PhosStop phosphatase inhibitor (Roche) and chill  On ice .
- 2 Aspirate media from cells and rinse cells with PBS  On ice . Aspirate PBS thoroughly. 
- 3 Pipette RIPA lysis buffer onto cells and scrape cells using a cell lifter (Corning).
- 4 Pipette lysis buffer containing cell mass into Eppendorf tube.
- 5 Treat with 25 units of benzinase for  00:05:00 . 
- 6 Spin down at  14000 rpm, 4°C, 00:05:00 . 

- 7 Collect supernatant.
- 8 Determine protein concentration in sample using Pierce BCA assay (ThermoFisher).
- 9 Prepare samples at desired concentration and add 4x Laemmli buffer.

Gel electrophoresis and immunoblotting (Tris-glycine buffer system)


4h 48m

- 10 Incubate samples at  95 °C for  00:03:00 . 

- 11 During this incubation, prepare gel apparatus with Mini PROTEAN TGX 4-15% tris-glycine gels (Bio-Rad) and running buffer.



- 12 Load samples into gel and run until dye front reaches bottom (250 V).
- 13 Remove gel and set up transfer cassette with nitrocellulose membrane.
- 14 Transfer at 100 V for  01:00:00 in tris-glycine transfer buffer. 1h
- 15 Remove nitrocellulose membrane and stain for total protein with ponceau stain.
- 16 Wash with milliQ water.
- 17 Block membrane with 5% milk in TBST for  01:00:00 at  Room temperature . 1h
- 18 Incubate membrane with primary antibodies in 2.5% milk in TBST  Overnight at  4 °C . 1h
- Note**
- NOTE:** Optimal primary antibody incubation time and temperature can be determined empirically for a given primary antibody.
- 19 Wash membrane. 
- 19.1 Wash membrane for  00:10:00 with TBST. (1/3) 10m
- 19.2 Wash membrane for  00:10:00 with TBST. (2/3) 10m
- 19.3 Wash membrane for  00:10:00 with TBST. (3/3) 10m



20 Incubate membrane with secondary antibodies conjugated HRP for  01:00:00 .

1h



21 Wash membrane.



21.1 Wash membrane for  00:05:00 with TBST. (1/3)

5m

21.2 Wash membrane for  00:05:00 with TBST. (2/3)

5m

21.3 Wash membrane for  00:05:00 with TBST. (3/3)

5m

22 Image membranes using a Biorad Chemidco.