



Mar 19, 2024

Immunoblotting of I3 Neurons and dopaminergic neurons

DOI

<https://dx.doi.org/10.17504/protocols.io.3byl49eqjgo5/v1>

Nisha Mohd Rafiq¹, Pietro De Camilli^{2,3,4,5}

¹University of Tuebingen;

²Department of Neuroscience, Yale University School of Medicine, New Haven, Connecticut 06510, USA.;

³Department of Cell biology, Yale University School of Medicine, New Haven, Connecticut 06510, USA.;

⁴Program in Cellular Neuroscience, Neurodegeneration and Repair.;

⁵Aligning Science Across Parkinson's (ASAP) Collaborative Research Network, Chevy Chase, MD, 20815, USA.



Berrak Ugur

Yale University

Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

OPEN  ACCESS



DOI: <https://dx.doi.org/10.17504/protocols.io.3byl49eqjgo5/v1>

Protocol Citation: Nisha Mohd Rafiq, Pietro De Camilli 2024. Immunoblotting of I3 Neurons and dopaminergic neurons. protocols.io <https://dx.doi.org/10.17504/protocols.io.3byl49eqjgo5/v1>

License: This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited



Protocol status: Working

We use this protocol and it's working

Created: March 12, 2024

Last Modified: May 31, 2024

Protocol Integer ID: 96917

Keywords: ASAPCRN, i3 neuron, dopaminergic neurons this protocol, preparation of cell lysate, immunoblotting procedure, cell lysate, dopaminergic neuron, derived neuron, neuron, dopaminergic

Funders Acknowledgements:

ASAP

Grant ID: ASAP-000580

Abstract

This protocol describes the preparation of cell lysate from and iPSC-derived neurons (i³ Neurons, dopaminergic) and the immunoblotting procedure.

Attachments



Rafiq_Immunoblotting...

17KB

Materials

- **TBS:** [M] 50 millimolar (mM) Tris-Cl, [M] 150 millimolar (mM) NaCl, adjust  07.5 .
- **TBST:** TBS with 0.1% TWEEN-20 (Sigma-Aldrich).



Cell culture and treatments

10m

- 1 Grow I³ Neurons and dopaminergic (DA) neurons on six-well plates ($3-5 \times 10^5$ cells/well).
- 2 After differentiation in their respective maturation media, wash the neurons with  1 mL ice-cold PBS. 
- 3 Lyse the cells in  200 μ L 1xRIPA lysis buffer (10X RIPA lysis buffer, Sigma-Aldrich) supplemented with complete™ EDTA-free protease inhibitor cocktail (Roche) and PhosSTOP phosphatase inhibitor cocktail (Roche).
- 4 Centrifuge the cells at  13000 x g for  00:10:00 . 
10m
- 5 Collect the supernatant and store at  -20 °C .
- 6 Determine the protein concentration in sample using Pierce BCA assay (ThermoFisher).

Gel electrophoresis and immunoblotting (Tris-glycine buffer system)

5h 10m

- 7 Incubate appropriate volume of cell lysate solution at  95 °C for  00:05:00 in SDS sample buffer containing 1% 2-mercaptoethanol (Sigma). 
5m
- 8 Separate the extracted proteins by SDS-PAGE in Mini-PROTEAN TGX precast polyacrylamide gels (Bio-Rad) at 250V and transfer to nitrocellulose membranes (Bio-Rad) at 100 V for  01:00:00 or 75 V for  02:00:00 (for high molecular weight proteins: >150 kDa). 
3h
- 9 Block the nitrocellulose membranes for  01:00:00 with 5% non-fat milk (AmericanBIO) in TBST (tris-buffered saline [TBS] + 0.1% tween 20), then incubate  Overnight at  4 °C with primary antibodies.  
1h

Note

Antibody dilutions can be found in Table S1.



- 10 Wash the blots with TBST, thrice, each  00:05:00 .
- 11 Incubate the blots with IRDye 680RD or 800CW (LI-COR) secondary antibodies (1:8000) for  01:00:00 at  Room temperature in TBST.
- 12 Image blots using the Gel Doc imaging system (Bio-Rad) using manufacturer's protocols.

5m



1h

