

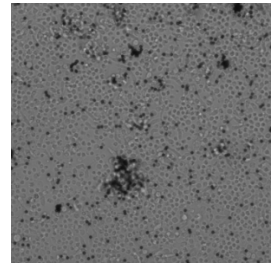
Nov 07, 2018

Version 6

Human Primary T Cells: A Practical Guide V.6

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External link: <https://github.com/hammerlab/t-cell-guide>



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Protocol status: Working

We use this protocol and it's working

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Keywords: T cells, human primary cells, tissue culture, electroporation, optimization, standardization, PBMC, activation, human immune cell, characteristics of the human immune cell, cancer immunotherapy, cell, fresh human blood sample

Abstract

Human primary T cells are invaluable and feasible model systems to study the characteristics of the human immune cells in various contexts, including but not limited to cancer immunotherapy. Following isolation of T cells from fresh human blood samples, it is possible to culture, expand, and manipulate these cells, which allows extensive investigation for research purposes. Techniques for isolation and handling of T cells are well-established but parts of the protocols can highly vary across different labs. These differences in the protocols are, often, there due to historical reasons and are only supported by anecdotal evidence. We systematically modified basic components of the T cell culturing protocols and collected data on how they altered the final yield. Here, based on these data, we provide practical hints and tips on basic cellular and molecular techniques for handling primary human T cells. We hope that this guide will serve as a reference point to allow evaluate, discuss, and improve current practices in T cell culturing and manipulation.

Troubleshooting



Files

Protocol



NAME

CRISPR/Cas9-based knock-out in human primary T cells (24-well setup)

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Resazurin viability assay for human primary T cells in 96-well format

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Preparing primary T cells for fluorescence microscopy

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NAME

PBMC isolation from buffy coat

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Human primary T cell culture media

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