

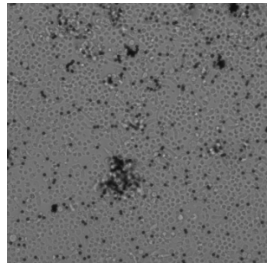
Nov 06, 2018

Version 4

Human Primary T Cells: A Practical Guide V.4

DOI

dx.doi.org/10.17504/protocols.io.vdde226



Bulent Arman Aksoy¹, Pinar Aksoy¹, Megan Wyatt¹, Chrystal M. Paulos¹, Jeff Hammerbacher¹

¹Medical University of South Carolina



Bulent Arman Aksoy

Medical University of South Carolina

Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

OPEN  ACCESS



DOI: <https://dx.doi.org/10.17504/protocols.io.vdde226>

External link: <https://github.com/hammerlab/t-cell-guide>

Collection Citation: Bulent Arman Aksoy, Pinar Aksoy, Megan Wyatt, Chrystal M. Paulos, Jeff Hammerbacher 2018. Human Primary T Cells: A Practical Guide. **protocols.io** <https://dx.doi.org/10.17504/protocols.io.vdde226>

License: This is an open access collection distributed under the terms of the **Creative Commons Attribution License**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited



Protocol status: Working

We use this protocol and it's working

Created: November 06, 2018

Last Modified: November 06, 2018

Collection Integer ID: 17541

Keywords: T cells, human primary cells, tissue culture, electroporation, optimization, standardization, PBMC, activation, human immune cell, characteristics of the human immune cell, cancer immunotherapy, cell, fresh human blood sample

Abstract

Human primary T cells are invaluable and feasible model systems to study the characteristics of the human immune cells in various contexts, including but not limited to cancer immunotherapy. Following isolation of T cells from fresh human blood samples, it is possible to culture, expand, and manipulate these cells, which allows extensive investigation for research purposes. Techniques for isolation and handling of T cells are well-established but parts of the protocols can highly vary across different labs. These differences in the protocols are, often, there due to historical reasons and are only supported by anecdotal evidence. We systematically modified basic components of the T cell culturing protocols and collected data on how they altered the final yield. Here, based on these data, we provide practical hints and tips on basic cellular and molecular techniques for handling primary human T cells. We hope that this guide will serve as a reference point to allow evaluate, discuss, and improve current practices in T cell culturing and manipulation.

Troubleshooting

Files

 SEARCH

Protocol

NAME

Human primary T cell culture media

VERSION 1

CREATED BY



Bulent Arman Aksoy
 Medical University of South Carolina

OPEN →

Protocol



NAME

Resazurin viability assay for human primary T cells in 96-well format

VERSION 1

CREATED BY



Bulent Arman Aksoy
 Medical University of South Carolina

OPEN →

Protocol



NAME

CRISPR/Cas9-based knock-out in human primary T cells (24-well setup)

VERSION 1

CREATED BY



Bulent Arman Aksoy
 Medical University of South Carolina

OPEN →

Protocol



NAME

PBMC isolation from buffy coat

VERSION 2

CREATED BY



Bulent Arman Aksoy
Medical University of South Carolina

OPEN →