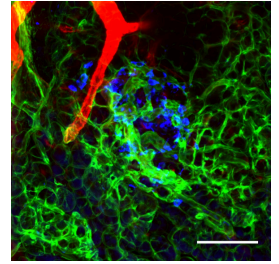


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# Human Islet Microvasculature Immunofluorescence in Optically Cleared Samples

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**Protocol status:** Working

We use this protocol and it's working

**Created:** March 12, 2019

**Last Modified:** August 14, 2020



Protocol Integer ID: 21331

**Keywords:** human islet, glucagon, optical clearing, confocal microscopy, 3D, PACT, collagen IV, SMA, Vesselucida360, alpha-cells, pancreas, neuro-insular network, passive CLARITY

## Disclaimer

Different primary and secondary antibody lots may differ in affinities and should be independently optimized.

## Abstract

This protocol describes the immunostaining performed on five human control pancreas samples, cleared using a modified passive CLARITY (PACT) method according to our published methods. Islets were identified based on alpha-cells stained with glucagon while collagen-containing basement membranes in the extracellular matrix were stained using anti-collagen IV. Smooth muscle cells surrounding arteries and arterioles were identified using anti-SMA primary antibody directly conjugated with Cy3 fluorophore. An accompanying protocol describes using Vesselucida360 to contour islets and determine islet basement membrane and SMA densities and morphometric variables (<https://dx.doi.org/10.17504/protocols.io.bjfzkjp6>).

## Attachments



[jove-protocol-56859-...](#)


786KB

## Image Attribution

An islet is shown from case 6232 with all 3 fluorescent channels visible.

## Guidelines

This protocol is for PACT-cleared human pancreas samples, fixed with 4% paraformaldehyde for at minimum

 48:00:00 , 4% agarose embedded and sectioned at 400 um or greater thickness. The Triton-X concentration in the antibody diluent was increased from 0.1% to 0.5% based on experiments that showed antibody penetration into the middle of the sample was improved without loss of signal.

## Materials

### MATERIALS

- ⊗ Glucagon (GCG) Mouse anti-human **Abcam Catalog #ab10988** (Lot# GR290488-2)
- ⊗ Goat anti-ms Dylight 405 hi-cross **Thermo Fisher Scientific Catalog #35500BID**
- ⊗ PBS Phosphate Buffered Saline 10X Solution **Fisher Scientific Catalog #BP399-1**
- ⊗ Goat normal serum **Vector Laboratories Catalog #S-1000**
- ⊗ Triton X-100 **Merck MilliporeSigma (Sigma-Aldrich) Catalog #T8787-100**
- ⊗ Collagen IV (Col14) Rabbit anti-human **Abcam Catalog #ab6586**
- ⊗ Smooth muscle actin- alpha (a-SMA) Mouse anti-human-Cy3 [clone 1A4] **Merck MilliporeSigma (Sigma-Aldrich) Catalog #C6198**
- ⊗ Goat anti-rb AF488 hi cross **Thermo Fisher Scientific Catalog #A-11034**

### STEP MATERIALS

- ⊗ Triton X-100 **Merck MilliporeSigma (Sigma-Aldrich) Catalog #T8787-100**
- ⊗ PBS Phosphate Buffered Saline 10X Solution **Fisher Scientific Catalog #BP399-1**

#### Software

**Vesselucida 360**

NAME

MBF Bioscience

DEVELOPER

#### Software

**Vesselucida Explorer**

NAME

MBF Bioscience

DEVELOPER




## Protocol materials




- Goat anti-ms Dylight 405 hi-cross **Thermo Fisher Scientific Catalog #35500BID**
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## Safety warnings

- ⚠ Follow all laboratory safety procedures when handling hazardous chemicals and materials.


## Before start

Ensure samples are cleared and the SDS is well-removed by extensive washing in PBS at  Room temperature on a rocker plate.


- 1 For these samples, optical clearing was performed using 4% SDS at  60 °C with rocking for up to 3 weeks until the samples reached transparency. Our original protocol used 8% SDS. We have subsequently tested both 4% and 8% SDS at  Room temperature and  60 °C . Different tissues should be tested independently for optimal results.

## Prepare the Antibodies

- 2 Make antibody dilution buffer (ABD): 1x PBS containing 2% normal goat serum (NGS) and 0.5% Triton-X 100.



 PBS Phosphate Buffered Saline 10X Solution **Fisher Scientific Catalog #BP399-1**



 Goat normal serum **Vector Laboratories Catalog #S-1000**


 Triton X-100 **Merck MilliporeSigma (Sigma-Aldrich) Catalog #T8787-100**

- 3 Dilute the primary antibodies to 1:200 in ABD and prepare at least  1 mL per sample.




## Stain the Tissue

- 4 Incubate the samples with primary antibodies on a rocker plate for  48:00:00 1-5 days at  Room temperature .


- 5 Wash the tissue sections with 1xPBS for five times, minimum  01:00:00 at  Room temperature .

 PBS Phosphate Buffered Saline 10X Solution **Fisher Scientific Catalog #BP399-1**



6 Dilute the secondary antibodies to 1:500 and prepare at least  1 mL per sample. Incubate for  48:00:00 or more on a rocker at  Room temperature .

7 Wash the tissue with 1 x PBS 5 times  01:00:00 at  Room temperature .

8 Equilibrate the samples in RIMS containing 0.5% sodium azide for at least  12:00:00 before imaging.  
Transfer to an imaging dish or chamber slide using RIMS as mounting media.

## Imaging

9 Perform confocal microscopy. Each microscope will require optimization of imaging parameters.

## Equipment

LSM 710

NAME

confocal laser microscope

TYPE

Zeiss

BRAND

LSM 710

SKU

<https://www.zeiss.com/microscopy/us/products/confocal-microscopes.html><sup>LINK</sup>



10 The following lasers were used for imaging each antigen in these images:

Track 1- 488- Collagen IV

Track 2- 405- GCG, 561- SMA

## Results

11

### Expected result

Islet alpha-cells- GCG Stain - Specific Stain, Intensity Good, Background Low  
Collagen IV- basement membranes- Specific Stain, Intensity Good, Background medium  
SMA- smooth muscle cells surrounding vessels - Specific Stain, Intensity Good, Background Low

12 Analyze islet microvascular density by contouring the GCG+ area for islet volume followed by collagen IV and SMA morphometry using Vesselucida360.