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Human and mouse alpha-synuclein protein expression and purification

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We use this protocol and it's working well

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Abstract

The protocol for is designed for high-yield purification of recombinant α -synuclein monomer. It is recommended to always store the protein on ice, and once the purification process has started, it should not be stopped.



Materials

⊗ BL21-Gold (DE3) Competent Cells **Agilent Technologies Catalog #230132**

⊗ Nalgene™ Rapid-Flow™ Sterile Single Use Vacuum Filter Units **Thermo Fisher Scientific Catalog #565-0010**

⊗ SnakeSkin Dialysis Tubing 3.5K MWCO 35 **Thermo Fisher Scientific Catalog #88244**

⊗ Endotoxin detection kit LAL **Genscript Catalog #95045-024**

⊗ ToxinEraser™ Endotoxin Removal Kit **Genscript Catalog #89233-330**

⊗ ToxinEraser™ Endotoxin Removal Resin **Genscript Catalog #L00402**

⊗ HiPrep Q HP anion exchange chromatography column **Cytiva Catalog #29018182**

⊗ MilliporeSigma™ Amicon™ Ultra-15 Centrifugal Filter Units **Catalog #MilliporeSigma™ UFC901024**



Protocol materials

☒ Nalgene™ Rapid-Flow™ Sterile Single Use Vacuum Filter Units **Thermo Fisher Scientific Catalog #565-0010**

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☒ ToxinEraser™ Endotoxin Removal Resin **Genscript Catalog #L00402**

☒ Endotoxin detection kit LAL **Genscript Catalog #95045-024**

☒ BL21-CodonPlus (DE3)-RIL **Agilent Technologies Catalog #230245-41**

☒ 1.5mL Micro Centrifuge Tube; endotoxin-free **CELLTREAT Catalog #50-202-024**

☒ ToxinEraser™ Endotoxin Removal Kit **Genscript Catalog #89233-330**

☒ ToxinEraser™ Endotoxin Removal Resin **Genscript Catalog #L00402**

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☒ SnakeSkin Dialysis Tubing 3.5K MWCO 35 **Thermo Fisher Scientific Catalog #88244**

☒ Endotoxin detection kit LAL **Genscript Catalog #95045-024**

☒ SOC Medium **Merck MilliporeSigma (Sigma-Aldrich) Catalog #S1797-10X5ML**

☒ Ampicillin sodium salt **Merck MilliporeSigma (Sigma-Aldrich) Catalog #A0166**

☒ Agar powder **Grainger Catalog #31FZ34**

☒ LB Broth, Miller (Granulated) **Fisher Scientific Catalog #BP9723-500**

☒ IPTG **Research Products International Corp (RPI) Catalog #I56000-5.0**

☒ SODIUM CHLORIDE **Fisher Scientific Catalog #S2711**

☒ EDTA, disodium salt, dihydrate **Fisher Scientific Catalog #S312-500**

☒ cOmplete™, EDTA-free Protease Inhibitor Cocktail MIDI **Merck MilliporeSigma (Sigma-Aldrich) Catalog #4693132001**

☒ Tris Base **Research Products International Corp (RPI) Catalog #T60040-5000.0**

Troubleshooting

Transformation

1d

1 Thaw down an aliquot of plasmid construct (pRK172) encoding WT-human-a-synuclein or mouse-a-synuclein [M] 0.3 mg/mL ⚠ On ice

15m

2 Thaw down ⚠ On ice an aliquot of BL21 (DE3) RIL competent E Coli cells

15m

⊗ BL21-CodonPlus (DE3)-RIL **Agilent Technologies Catalog #230245-41**

3 Add 🧪 1 µL of plasmid construct to the thawed competent cells and gently mix by flicking the bottom of the tube with a finger a few times



Safety information

do not resuspend

3.1 Incubate the reaction mix ⚠ On ice

15m



4 Perform heat-shock transformation ⚠ 42 °C in water bath incubator with manually shaking at 🌀 100 rpm, 00:00:45

1m



Equipment

Precision™ General Purpose Water Bath

NAME

Water Bath

TYPE

Thermo Scientific

BRAND

TSGP10

SKU

5 Immediately transfer the tube on ice and incubate for 1 min.

1m

6 Add  1000 μL of SOC media to a chilled reaction

10s

 SOC Medium **Merck MilliporeSigma (Sigma-Aldrich) Catalog #S1797-10X5ML**

7 Incubate the bacteria  200 rpm, 37°C, 00:30:00

30m

Equipment

ThermoMixer® C

NAME

ThermoMixer


TYPE

Eppendorf

BRAND

5382000023

SKU


7.1 Prepare sterile 10cm LB agar plate containing  0.1 mg/mL of ampicillin

 Ampicillin sodium salt **Merck MilliporeSigma (Sigma-Aldrich) Catalog #A0166**


 Agar powder **Grainger Catalog #31FZ34**

8 Collect 50 ul of cell suspension (Tube #1 5% of cells)

3m

Centrifuge 950 cell suspension at  500 x g, 10°C, 00:03:00

Save the pellet with approximately  50 μL of media (Tube #2 95% of cells)

8.1 Spread tubes #1 and #2 onto a selection plate and incubate overnight at  37 °C in bacterial incubator

10m



Equipment

Isotemp™ Microbiological Incubator, 178 L, Stainless Steel^{NAME}



Microbiological Incubator^{TYPE}

Fisherbrand^{BRAND}

15-103-0513^{SKU}

Protein expression

12h

- 9 Pick one colony and transfer into  10 mL LB media with  0.1 mg/mL of ampicillin start in the morning (9:00 am)

 LB Broth, Miller (Granulated) Fisher Scientific Catalog #BP9723-500

- 9.1 Incubate the bacteria  250 rpm, 37°C, 05:00:00 until it reaches OD 0.2-0.3

5h

Equipment

Natural convection incubator^{NAME}

Bacterial shaker^{TYPE}

Innova^{BRAND}

M1335-0000^{SKU}



- 10 Transfer a starter culture to 2X2L flasks filled with 0.5L LB media with [M] 0.1 mg/mL of ampicillin
5 mL to each flask

Equipment

New Brunswick™ Innova® 42

NAME

Incubated Benchtop Shakers

TYPE

Eppendorf

BRAND

M1335-0004

SKU



- 11 Incubate the culture at the same conditions until it reaches OD 0.8 (use nanodrop or cuvette) (reaches optimal density at 6-7 pm)

5h

- 12 Induce protein expression by adding [M] 0.05 millimolar (mM) IPTG, incubate at

12h

🌡️ 18 °C for ⌚ 12:00:00 overnight



⚙️ IPTG Research Products International Corp (RPI) Catalog #I56000-5.0

Note

To cool down the grown culture, transfer the flasks into ice-bath and incubate until it reaches desired temperature

Cell lysis

11m 45s

- 13 Collect the pellets by centrifugation (JA14 rotor) at 🌀 5000 x g x g at 🌡️ 4 °C for
⌚ 00:10:00 . Used 250 ml Beckman tubes
Usually get 10-12 g from 2L

10m

Equipment

Avanti® J-E Centrifuge

NAME

Avanti Centrifuge

TYPE

Beckman

BRAND

369005

SKU



- 14 Add to pellets 80 ml of lysis buffer (total): [M] 10 millimolar (mM) ThisHCl pH 7.6 , [M] 750 millimolar (mM) NaCl, [M] 1 millimolar (mM) EDTA, [M] 1 millimolar (mM) PMSF (add just before using, have aliq frozen [M] 0.1 Molarity (M)), protease inhibitors (use MAXI version, need only one tablet);

⊗ SODIUM CHLORIDE Fisher Scientific Catalog #S2711

⊗ Tris Base Research Products International Corp (RPI) Catalog #T60040-5000.0

⊗ EDTA, disodium salt, dihydrate Fisher Scientific Catalog #S312-500

⊗ cComplete™, EDTA-free Protease Inhibitor Cocktail MIDI Merck MilliporeSigma (Sigma-Aldrich) Catalog #4693132001

- 15 Carefully resuspend the pellets to homogenize the solution

- 15.1 Heat up 1 L of water in a high temperature resistant glass beaker (turn heat to the max on the magnetic stirrer)

- 16 While waiting on water to get to the boiling point sonicate the lysates (use thick prob-tip) for 00:01:00 , 30%, 00:00:15 ON 00:00:30 OFF of amplitude then go to next falcon, had 3 falcons (repeat 3 times, avoid overheating)

10m



- 17 After sonication samples need to get boiled thereby put the falcon tubes into glass beaker and boil for 00:25:00 . Use tweezers to pull out the tubes 25m
- 18 Transfer boiled homogenates into new 50 mL falcon tubes; chill down suspensions at room temperature for 20 min 20m
- 19 Prepare 4 L of buffer 10 millimolar (mM) TrisHCl 7.6 ,
 50 millimolar (mM) NaCl, 1 millimolar (mM) EDTA, 1 millimolar (mM) PMSF for dialysis
- 20 Centrifuge the homogenates at 20000 x g for 01:00:00 at 4 °C 1h
- 21 Filter the supernatant using 0.45 um filter unit
 Nalgene™ Rapid-Flow™ Sterile Single Use Vacuum Filter Units **Thermo Fisher Scientific Catalog #565-0010**
- 22 Transfer filtered supernatant into dialysis bag which is: SnakeSkin Dialysis tubing, 3.5K MWCO, 35 mm dry I.D., 35 feet.
Measure the dialysis tube taking into consideration that 5 cm length of tube holds 48 mL of the sample (plus 2.5cm at each end for closure). Clip the tube using green clips, make sure it does not leak.
Place the dialysis bag into 4 L plastic beaker filled with dialysis buffer, incubate overnight on magnetic plate on the slow mode (Chromatography fridge)
 SnakeSkin Dialysis Tubing 3.5K MWCO 35 **Thermo Fisher Scientific Catalog #88244**



Equipment

ÄKTA pure 25 L1

NAME

ÄKTA pure chromatography system

TYPE


Cytiva

BRAND

29018225

SKU

Protein purification (anion-exchange chromatography)

23 After a night of dialysis ( 4 °C slow mixing) collect the suspension into 100 mL glass bottle (filter the sample before running on the column, 0.22 um filter).

24 Column - HiPrep Q HP 16/10 column 1×20 ml (stored in 70% ethanol);

 HiPrep Q HP anion exchange chromatography column **Cytiva Catalog #29018182**

24.1 Wash the column 2V of miliQ degassed water

24.2 Wash the column with 2V of **STARTING BUFFER** [M] 10 millimolar (mM) TrisHCl


 7.6 , [M] 50 millimolar (mM) NaCl

24.3 Activate with 1V of [M] 10 millimolar (mM) TrisHCl  7.6 , [M] 1 Molarity (M) NaCl

24.4 Equilibrate with 3V of starting buffer


25 Load  80 mL of suspension and then washed with 100 ml [M] 50 millimolar (mM)





NaCl [M] 10 millimolar (mM) TrisHCL,  300 mL of gradient elution (0-100%), 2 ml/min flow rate. Collected samples using fraction collector 2, every fraction 4 ml (use 10 ml glass tubes)

26 Place supernatant into channel A1 (was previously use for starting buffer, do not generate bubbles)



27 Place starting buffer in channel A2 (clean the tubing using the program mode)

28 Place elution buffer in channel B1 ([M] 10 millimolar (mM) TrisHCl  7.6 ,
[M] 1 Molarity (M) NaCl)
Collected samples using fraction collector 2, every fraction 4 ml (use 10 ml glass tubes);

29 Analyze the fractions eluted at 250-350 mM salt (20 RFU conductivity) though SDS-PAGE (stain with Coomassie).
Combine  10 μ L of each fraction with  10 μ L of 2X laemmli buffer and analyze fractions by SDS-PAGE with 4-20% gradient gels, followed by coomassie staining/destaining


30 Measure A280/260 for the fractions containing single a-syn band, avoid collecting samples with A280/260 > 0.85

31 Combine the evaluated factions and measure total protein concentration using nanodrop.

32 Dialyze with  4 L of [M] 10 millimolar (mM) TrisHCl  7.6 ,
[M] 50 millimolar (mM) NaCl (follow instruction for dialysis)



Further purification

33 Repeat section 'Protein purification (anion-exchange chromatography)' for the further fractionation of the purified preparation 




Protein concentration

10m

34 Concentrate dialyzed protein sample to approximately [M] 30 mg/mL aliquot



Add  3 μL of 10x diluted aliquot in PBS onto nanodrop pedestal;

Parameters:

- other proteins; coefficient extinction: 5.98; MW: 14.4 kDA (**for wild-type human α -synuclein**)

- other proteins; coefficient extinction: 7.45; MW: 14.4 kDA (**for wild-type mouse α -synuclein**)

Perform two measurements and confirm <10% standard error between two measurements

If necessary, prepare 20X and 30X dilutions to confirm findings.

Equipment

NanoDrop™ One/OneC Microvolume UV-Vis Spectrophotometer^{NAME}

UV-Vis Spectrophotometer^{TYPE}

Thermo Scientific^{BRAND}




ND-ONE-W^{SKU}

Prepare the ultra-concentration system



35 Use 50 mL ultra centrifugation units with 3K cutoff



MilliporeSigma™ Amicon™ Ultra-15 Centrifugal Filter
Units **Catalog #**MilliporeSigma™ UFC901024

36 Wash off the unit with miliQ water through centrifugation at  5000 x g at  4 °C
for  00:05:00 , JA10 rotor

5m

37 Load first  15 mL of the sample into ultracentrifugation unit (max load of the unit is approx.  15 mL)

38 Centrifuge at  5000 x g at  4 °C for  00:05:00 , JA10 rotor

5m



39 Resuspend concentrated sample, add more of protein sample and concentrate until the total volume is ~ 5 mL

40 Store at -80 °C . Yield should be approximately 80 mg per 2 L culture



Endotoxin removal

- 41 Follow instructions for
ToxinEraser™ Endotoxin Removal Kit **Genscript Catalog #89233-330** with modifications
For a more successful endotoxin removal, add 1 mL of
ToxinEraser™ Endotoxin Removal Resin **Genscript Catalog #L00402** before the regeneration process
- 42 Collect the eluate into 5 mL endotoxin-free tube and save 2 aliquots (10 µL and 50 µL) for protein concentration and endotoxin measurements
1.5mL Micro Centrifuge Tube; endotoxin-free **CELLTREAT Catalog #50-202-024**

Endotoxin quantification

- 43 Follow instructions for Endotoxin detection kit LAL Genscript
Endotoxin detection kit LAL **Genscript Catalog #95045-024**