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Version 2

## hsqc\_metab.nan V.2

Forked from a private protocol



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**We use this protocol and it's working**

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## Abstract

This is a protocol for running the Bruker pulse program "hsqcetgpsisp2".

## Guidelines

This protocol intends to provide concise instructions to carry out the experiment. For more comprehensive information, see Bruker's documentation "Basic NMR Experiments" by clicking **? → Manuals (docs)** on the menu bar on TopSpin. See also "Pulse Program Catalogue. 1D/2D" for the details about the pulse program used in this protocol.

## Troubleshooting



## Before start

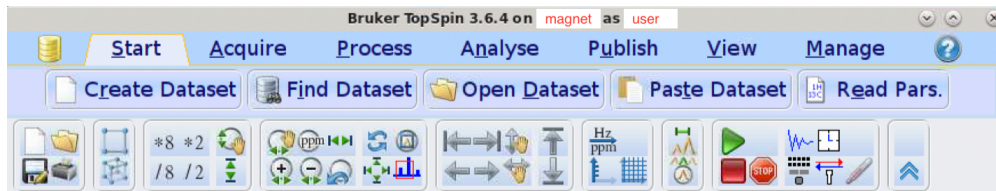
This protocol assumes:

- Your sample is loaded, locked, tuned for both proton and carbon channels, and shimmed in the magnet
- The calibrated 90° pulse value for proton (i.e., P1) for the sample has been collected

## Create a new dataset

1

- 1.1 On the menu bar on TopSpin, click on **Start → Create Dataset**



(This protocol uses TopSpin 3.6.4, and the interface may look different on other TopSpin versions.)

### Note

You can also use the **new** command in the command line to do this step.

- 1.2 Enter
- **NAME**: Name of a set of datasets (e.g., human\_serum\_study1). Use a single string.
  - **EXPNO**: Dataset number. Use a positive integer.

Select

- **Directory**: Your directory.



Create New Dataset - new

Prepare for a new experiment by creating a new data set and initializing its NMR parameters according to the selected experiment type. For multi-receiver experiments several datasets are created. Please define the number of receivers in the Advanced.

Dataset

NAME

EXPNO

Directory

☐ Open in new window

Parameters

☐ Use current parameters

☒ Read parameterset

☒ Set solvent

Additional action

☒ No additional action

☐ Execute getprosol

☐ Keep parameters

Advanced

Number of datasets (receivers)

Title

### Note

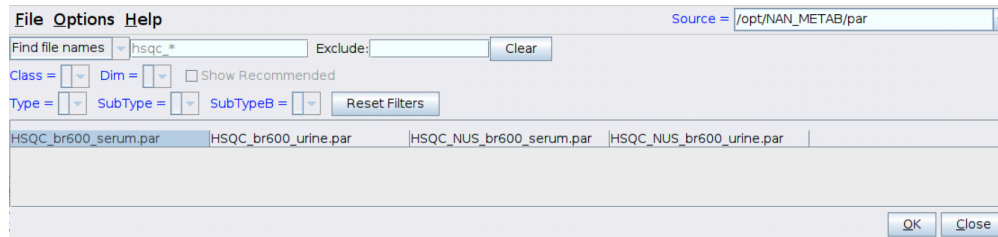
Your new dataset will be stored in **Directory/NAME/EXPNO**

### 1.3 Select Read parameterset

Click the button

**Select**

### 1.4 A new window opens. On the right top bar, select Source = /opt/NAN\_METAB/par



In the list, select the one you want to use:

For serum and plasma samples:

- **HSQC\_br600\_serum.par**: Parameter set using an acquisition mode "traditional planes"
- **HSQC\_NUS\_br600\_serum.par**: Parameter set using an acquisition mode "non-uniform sampling (NUS)". Higher resolution on the indirect dimension

For urine samples:

- **HSQC\_br600\_urine.par**: Parameter set using an acquisition mode "traditional planes"
- **HSQC\_NUS\_br600\_urine.par**: Parameter set using an acquisition mode "non-uniform sampling (NUS)". Higher resolution on the indirect dimension

#### Note

Parameter set names in the list vary between spectrometers (e.g., HSQC\_br800\_serum.par).

Click

**OK**

1.5

Click

**OK**

2

Go to the **"USE DEFAULT"** tab below to proceed with the default optimized parameters.

## STEP CASE

Use default parameters: 6 steps

This step case uses the default optimized parameters to acquire a spectrum.



3

- 3.1 Load the calibrated P1 using the following command in the command line.



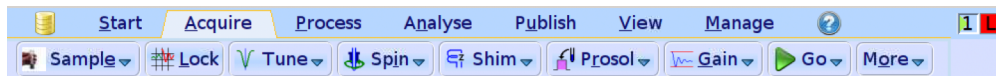
```
getprosol 1H [calibrated P1 value] [power level for P1]
```

(e.g., getprosol 1H 10.01 -7.45)

#### Note

**[power level for P1]** varies between spectrometers. Never use a wrong **[power level for P1]**

- 3.2 Click on  
**Acquire** → **Gain**  
in the menu bar to automatically set the receiver gain.



#### Note

You can also use the **rga** command in the command line.


- 3.3 Click  
**Go**  
in the menu bar to acquire a spectrum.

#### Note

You can also use the **zg** command in the command line.

- 3.4 After the run, click on  
**Process** → **Proc. Spectrum**  
in the menu bar to execute an automated processing macro.



- 3.5 If you want to modify parameters to improve your spectrum,  and move to the step case "MODIFY PAR".