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Protocol status: Working
 We use this protocol and it's working

HPA Cell Atlas Standard Immunostaining Protocol

Molecular Biology Reports

Peter Thul¹, Ulrika Axelsson²

¹Science for Life Laboratory / KTH - Royal Institute of Technology;

²Stockholm Royal Institute of Technology, KTH

Human Protein Atlas

Tech. support email: contact@proteinatlas.org

Peter Thul

Science for Life Laboratory / KTH - Royal Institute of Techn...

ABSTRACT

The Cell Atlas, part of the Human Protein Atlas project, systematically investigate the spatiotemporal subcellular distribution of human proteins. This is our standard protocol for indirect immunostaining of cells cultured in 96-well glass bottom plates.

GUIDELINES

All volumes in the protocol refer to volume per well in a 96-well microplate. Unspecified incubation temperatures are at room temperature. Immunostaining can be performed manually or with the help of a liquid handling robot.

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MATERIALS

MATERIALS

⊗ Disodium phosphate **Sigma Aldrich Catalog #S7907**

⊗ Monopotassium phosphate **Sigma Aldrich Catalog #P9791**

Buffers are prepared using MilliQ water

10X PBS

160 g NaCl

4 g KCl

28.8g Na₂HPO₄

4.8 g KH₂PO₄

Dissolve in 1600 ml Milli-Q. Adjust pH to 7.2 and add Milli-Q up to two liters. Filter and autoclave the buffer.

4% PFA/1X PBS

Use lab coat, eye protection and gloves and work in a fume hood.

1. Pre-heat 150 ml 10% FBS/1X PBS in a 250 ml beaker to 60°C on a hot plate with magnetic stirrer.
2. Add 50 ml 16% paraformaldehyde to the warm PBS and leave stirring for 20 min at 60°C.
3. Add concentrated NaOH until the solution reaches pH ~11.
4. Allow the solution to cool down to room temperature.
5. Set pH to 7.2-7.3 using first concentrated HCl, and then diluted HCl for fine adjustment.
6. Aliquot and store at -20°C. Thaw at room temperature right before use.

DAPI

1. Dissolve 10 mg of DAPI powder in 2 ml MilliQ water to a 5 mg/ml (14.3 mM) concentrated stock solution. Store at -20°C.
2. Mix 80 µl 5 mg/ml DAPI stock solution with 920 µl MilliQ water to a pre-diluted 400 µg/ml DAPI solution. Aliquot and store at -20°C.

Glycerol/10X PBS

1. Add 5 ml 10X PBS to 45 ml of glycerol. Mix.
2. Autoclave or filter sterilize.

SAFETY WARNINGS

- ⚠ Paraformaldehyde (PFA) is harmful in contact with skin or if inhaled, causes severe skin burns and eye damage, may cause an allergic skin reaction, may cause respiratory irritation, is suspected of causing genetic defects and may cause cancer.

BEFORE START INSTRUCTIONS

Cells are grown over night in 80 ul cell culture media (according to providers specifications) in 96-well glass bottom microplates coated with fibronectin (12.5 ug/ml). Optimal seeding concentration/confluency upon staining varies between cell lines and need to be tested in each setting.

Prior to immunostaining, primary antibodies generated within the Human Protein Atlas (HPA) project are diluted to 2 ug/ml in blocking buffer (4% FBS/1XPBS) together with 1 ug/ml tubulin marker (mouse anti-alpha tubulin, Ab7291, Abcam) and 1.25 ug/ml ER marker (chicken anti-calreticulin, Ab2908, Abcam).

Fixation

- 1 Remove the growth medium (i.e. aspirate ~ 80 µL from all wells).
- 2 Wash the cells once with 40 µL 1X PBS.
- 3 Fix the cells by incubating in 40 µL 4% PFA/1XPBS for 15 minutes. 00:15:00

Permeabilization

- 4 Remove the PFA and permeabilize the cells by incubating with 40 µL 0.1 % Triton X-100/1X PBS for 3 x 5 minutes. 00:05:00 00:05:00 05:00:00

- 5 Remove the Triton X-100 and wash the cells once with 40 μ L 1XPBS.

Primary Antibody Incubation

- 6 Add 40 μ L blocking buffer with diluted primary antibodies; 2 ug/ml HPA antibody, 1 ug/ml anti-alpha tubulin (Abcam, Ab7291), and 1.25 ug/ml anti-calreticulin (Abcam, Ab2908). Incubate over night at 4°C
or for two hours at room temperature.

4 °C 16:00:00 overnight

or

20 °C (RT) 02:00:00

Secondary Antibody Incubation

- 7 Remove the blocking buffer with primary antibodies and wash the cells with 40 μ L 1X PBS for 4 x 10 minutes. 00:10:00 00:10:00 00:10:00 00:10:00

- 8 Add 40 μ L blocking buffer with diluted secondary antibodies; 2.5 ug/ml anti-rabbit Alexa Fluor 488 (Thermo Fisher Scientific, A11034), 2.5 ug/ml anti-mouse Alexa Fluor 555 (Thermo Fisher Scientific, A21424), and 2.5 ug/ml anti-chicken Alexa Fluor 647 (Thermo Fisher Scientific, A21449). Incubate at room temperature for 90 minutes while keeping dark. 01:30:00

DAPI Nuclear Staining and Mounting

- 9 Remove the blocking buffer and incubate with 40 μ L 0.2 ug/ml DAPI/1X PBS for 10 minutes. 00:10:00

- 10** Wash the cells with 40 μ L 1X PBS for 4 x 10 minutes. 00:10:00 00:10:00
00:10:00 00:10:00
- 11** Fill the wells with glycerol/10X PBS and seal the plate with an adhesive aluminium PCR plate seal. Plates can be stored at 4°C for at least two weeks.