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## Homemade Gibson Mastermix

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External link: [https://openwetware.org/wiki/Gibson\\_Assembly](https://openwetware.org/wiki/Gibson_Assembly)

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**Protocol status:** Working

We use this protocol and it's working

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### Abstract

Recipe for homemade 1.33 x Gibson Mastermix.

## Materials

### MATERIALS

- ☒ beta-Nicotinamide adenine dinucleotide (NAD+) - 0.2 ml New England Biolabs Catalog #B9007S
- ☒ Q5 High-Fidelity DNA Polymerase - 100 units New England Biolabs Catalog #M0491S
- ☒ Taq DNA Ligase - 2,000 units New England Biolabs Catalog #M0208S
- ☒ T5 Exonuclease - 5,000 units New England Biolabs Catalog #M0363L
- ☒ PEG-8000
- ☒ DTT (Dithiothreitol) (> 99% pure) Protease free Gold Biotechnology Catalog #DTT
- ☒ Deoxynucleotide Solution Set - 25 umol of each New England Biolabs Catalog #N0446S

## Preparation of 5x isothermal reaction buffer

- 1 Recipe for 4 mL:

Component	Molarity / Concentration	Amount	Final concentration
Tris-HCl, pH 7.5	1 M	2 mL	500 mM
MgCl <sub>2</sub>	1 M	200 µL	50 mM
dATP	100 mM	40 µL	1 mM
dCTP	100 mM	40 µL	1 mM
dGTP	100 mM	40 µL	1 mM
dTTP	100 mM	40 µL	1 mM
DTT	1 M	200 µL	50 mM
PEG-8000	-	1 g	25 %
NAD <sup>+</sup>	100 mM	200 µL	5 mM
H <sub>2</sub> O	-	to final volume of 4 mL	

- 2 Mix dNTPs, NAD<sup>+</sup>, Tris-HCl, MgCl<sub>2</sub> and DTT.
- 3 Slowly add PEG-8000 to mixture and mix well, until completely dissolved. Add H<sub>2</sub>O to a final volume of 4 mL.
- 4 Prepare aliquots of the 5x isothermal buffer as required, e.g. 100 µL.  
Store at -20 °C.

## Preparation of 1,33x Assembly Mastermix

- 5 Recipe for 25 × 15 µL aliquots:

Component	Concentration	Amount	Final concentration (after adding DNA)
5x isothermal rxn buffer	5x	100 µL	1x
Taq DNA Ligase	40 U/µL	50 µL	4 U/µL
T5 Exonuclease	1 U/µL	2 µL	4 U/mL
Q5 Hi-Fi DNA Polymerase	2 U/µL	6.25 µL	25 U/mL

H <sub>2</sub> O		216.75	
<b>Total volume</b>		<b>375 µL</b>	

## Preparation of 1.33x Assembly Mastermix

- 6 Work on ice. Mix H<sub>2</sub>O and 5x buffer, then add enzymes.
- 7 Prepare 25 × 15 µL aliquots in PCR tubes. Store at -20 °C.  
These aliquots are concentrated 1.33 x - add your DNA in a volume of 5 µL to a final volume/concentration of 20 µL / 1x.

## Gibson assembly

- 8 After addition of DNA, incubate Gibson assembly mix at 50 °C for 45 min.  
 00:45:00 Gibson assembly

## Transformation

- 9 Transform chemically competent cells with an aliquot of your assembly mix.