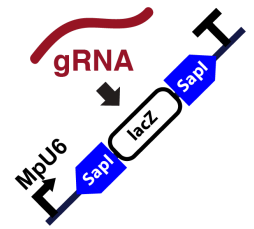


Dec 04, 2019

# gRNA design and cloning with Sapl into Loop plasmid L2\_lacZgRNA-Cas9-CsA

Forked from [gRNA design and cloning into Loop L2 plasmids \(L2\\_gRNA-Cas9-CsA and L2\\_gRNA-CsA plasmids\)](#).



DOI  
[dx.doi.org/10.17504/protocols.io.93wh8pe](https://dx.doi.org/10.17504/protocols.io.93wh8pe)  
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DOI: [dx.doi.org/10.17504/protocols.io.93wh8pe](https://dx.doi.org/10.17504/protocols.io.93wh8pe)

**Protocol Citation:** Eftychis Frangedakis, Marta Marta Tomaselli, Susana Sauret-Gueto 2019. gRNA design and cloning with Sapl into Loop plasmid L2\_lacZgRNA-Cas9-CsA. **protocols.io** <https://dx.doi.org/10.17504/protocols.io.93wh8pe>

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**Protocol status:** Working

**We use this protocol and it's working**

**Created:** December 04, 2019

**Last Modified:** December 04, 2019

**Protocol Integer ID:** 30550



## Abstract

This protocol explains how to design and clone the guide RNA target sequence into a L2 plasmid ready to accept the gRNA by cloning with SapI (L2 plasmid also contains a cassette to express Cas9)





oligo R (100 $\mu$ M) 1 $\mu$ l

water 8 $\mu$ l

Total volume 10 $\mu$ l

Anneal in a thermocycler using the following parameters: 37°C for 30 min, 95°C for 5 min and then ramp down to 25°C at 5°C per min. After annealing the gRNA can be directly cloned into L2\_lacZgRNACas9-CsA plasmid without the need of any further processing (step 4).

#### 4 **Cloning into backbone vector**

In a 0.2 mL tube set up the following reaction:

Component	Volume ( $\mu$ L)
Sterile water	5
10x Tango buffer (Thermo Fisher)	1
1 mg/mL bovine serum albumin (NEB)	0.5
T4 DNA ligase (5 U/ $\mu$ L) (Thermo Fisher)	0.25
10mM ATP (SIGMA)	1
SapI (LguI) (5 U/ $\mu$ L) (Thermo Fisher)	0.25
L2_lacZgRNACas9-Csa (25-50 ng)	1
annealed oligo	1
Final volume	10

- Place samples on the thermocycler and incubated using the following program:

Assembly: 15 cycles: 3 minutes at 37°C and 4 minutes at 16°C



Termination: 5 minutes at 50°C and 10 minutes at 80°C

- Transform chemically competent using 1 µL of reaction and plate on LB agar plates with 100 µg/mL spec and X-gal 40. Incubate at 37 °C for 16 h.
- Confirm with sequencing