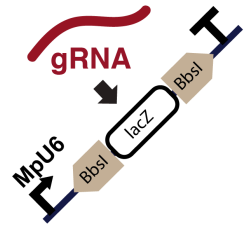


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gRNA design and cloning with BbsI into Loop plasmid L1_lacZgRNA-Ck2/3 V.1

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We use this protocol and it's working

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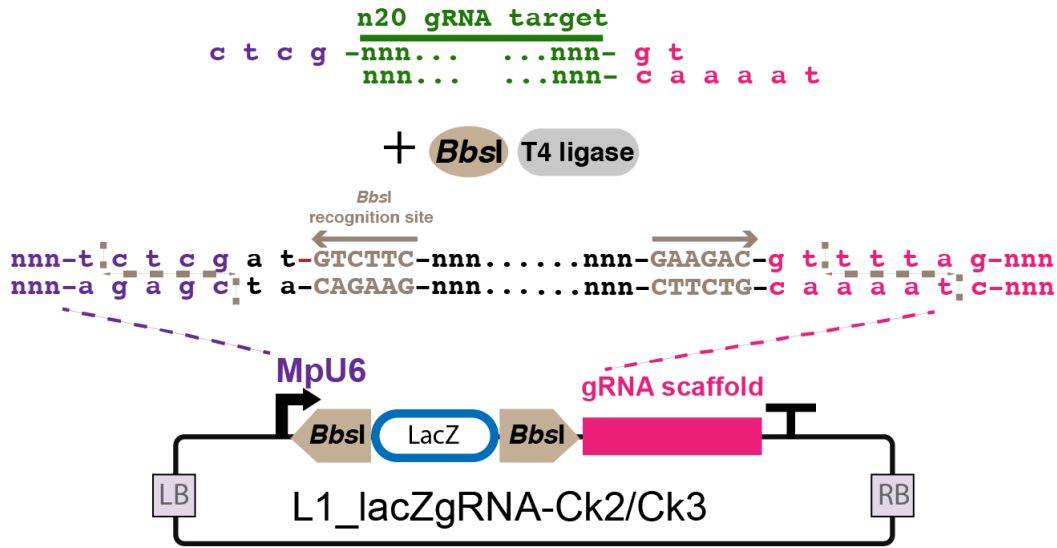


Abstract

This protocol explains how to design and clone the guide RNA target sequence into a L1 plasmid ready to accept the gRNA by cloning with BbsI. L1 plasmids are L1_lacZgRNA-Ck2 and L1_lacZgRNA-Ck3.

If one gRNA target sequence is cloned into the Ck2 plasmid and another one into Ck3 one, the two L1_gRNA transcription units can be combined with an antibiotic resistance transcription unit and a MpEF1 α :Cas9 transcription unit via **L2 SapI Loop assembly**. This allows for dual gRNA editing.

1



Design of oligos for gRNA BbsI mediated cloning into L1_lacZgRNA-Ck2 or L1_lacZgRNA-Ck3 vectors. The L1_lacZgRNA-Ck2/3 BbsI digested vectors have GAGC and TTTA overhangs. Therefore, oligos for gRNA should be designed such that the forward strand has a 5' overhang of CTCG and the reverse strand has a 5' overhang of gt-AAAT (addition of "gt" nucleotides is necessary to reconstitute the full sequence of the gRNA scaffold in pink). Light brown arrows: BbsI recognition site. Light brown dashed lines: BbsI cleavage site. LacZ: lacZa cassette for blue-white screening of colonies.

2 gRNA oligo design

Order two oligos that contain the forward and reverse guide sequence plus the overhangs necessary for ligation (highlighted with bold) into L1_lacZgRNA-Ck2 or L1_lacZgRNA-Ck3:

oligo F: 5'- CTCG-NNNNNNNNNNNNNNNNNNNNNN-gt 3'

oligo R: 5'- TAAAc-NNNNNNNNNNNNNNNNNNNNNN-3'

Note: Standard de-salted oligos are ok

3 Oligo annealing.

Mix oligos with water as follow:



oligo F (100 μ M) 1 μ l
oligo R (100 μ M) 1 μ l
water 8 μ l
Total volume 10 μ l

Anneal in a thermocycler using the following parameters: 37°C for 30 min, 95°C for 5 min and then ramp down to 25°C at 5°C per min. After annealing the gRNA can be directly cloned into L1_lacZgRNA-Ck2 or L1_lacZgRNA-Ck3 plasmid without the need of any further processing (step 4).

4 **Cloning into backbone vector**

In a 0.2 mL tube set up the following reaction:

Component	Volume (μ L)
Sterile water	6
10x T4 ligase buffer (NEB)	1
1 mg/mL bovine serum albumin (NEB)	0.5
T4 DNA ligase (5 U/ μ L) (Thermo Fisher)	0.25
SapI (LguI) (5 U/ μ L) (Thermo Fisher)	0.25
L1_lacZgRNA-Ck2 or L1_lacZgRNA-Ck3 (25-50 ng)	1
annealed oligo	1
Final volume	10

- Place samples on the thermocycler and incubated using the following program:

Assembly: 15 cycles: 3 minutes at 37°C and 4 minutes at 16°C

Termination: 5 minutes at 50°C and 10 minutes at 80°C



- Transform chemically competent using 1 μL of reaction and plate on LB agar plates with 100 $\mu\text{g/mL}$ spec and X-gal 40 $\mu\text{g/mL}$ for blue-white screening. Incubate at 37°C for 16 h.
- Confirm with sequencing