

Oct 09, 2023

Version 2

Golden Gate Assembly V.2

DOI

dx.doi.org/10.17504/protocols.io.kqdg3xknqg25/v2

NUS iGEM1

¹National University of Singapore



NUS IGEM

National University of Singapore

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Protocol Citation: NUS iGEM 2023. Golden Gate Assembly. protocols.io

https://dx.doi.org/10.17504/protocols.io.kqdg3xknqg25/v2
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Protocol status: Working

We use this protocol and it's working

Created: October 08, 2023



Last Modified: October 09, 2023

Protocol Integer ID: 88975

Keywords: Golden Gate, Assembly, DNA Assembly, DNA, Bsal, Restriction site, Restriction enzyme, type iis restriction enzyme, containing golden gate restriction site, golden gate assembly 2023 nus, golden gate assembly, dna oligo, same restriction site, plasmid backbone, restriction

Abstract

2023 NUS-Singapore iGEM team followed this protocol to assemble DNA oligos containing Golden Gate restriction sites with a plasmid backbone that contains the same restriction sites. The restriction enzymes utilised in this protocol are the Type IIS restriction enzymes, specifically Bsal. The use of Type IIS restriction enzymes ensures that there is no scar or extra sequence at the junctions between the assembled fragments.

Guidelines

This protocol outlines the Golden Gate procedures with a sample volume of 20 µL per reaction.

Protocol materials

⊠ Bsal-HF®v2 **New England Biolabs Catalog** #R3733S

X T4 DNA Ligase New England Biolabs Catalog #M0202S

Troubleshooting

Safety warnings



- Proper lab PPE must be worn at all times.
- Thermal gloves shall be worn when handling items from \$\mathbb{L} -20 \cdot \mathbb{C} fridge.



Golden Gate Assembly

- 1 Prepare an ice box.
- 2 Place the 2 10X NEB T4 DNA ligase buffer New England Biolabs ,
 - S Bsal-HF®v2 New England Biolabs Catalog #R3733S , and
 - X T4 DNA Ligase New England Biolabs Catalog #M0202S in ice.
- 3 Add the following reagents into a PCR tube:

| Item | Volume |
|-------------------|--------|
| DI Water | 10μL |
| Plasmid | 1μL |
| PCR Extract Oligo | 5μL |
| Bsal-HFv2 Enzyme | 1μL |
| T4 DNA Ligase | 1μL |
| T4 Ligase Buffer | 2μL |

Note

Reagents with enzymes such as Bsal-HFv2 Enzyme and T4 DNA Ligase must be kept at a low temperature (in ice) when they are in-use to prevent the enzymes from denaturation.

4 Put the sample into the Thermal Cycler and run it with the following conditions:

*Set "Lid Temperature" to \$\ 105 °C and set "Volume" to \$\ 20 \mu L

| Temperature | Duration |
|---|-----------|
| 37°C | 5 minutes |
| 16°C | 5 minutes |
| Go to step 1, repeat the cycle 40 times | |



| Temperature | Duration |
|-------------|---------------|
| 37°C | 1 hour |
| 60°C | 15 minutes |
| 12°C | Infinite Loop |

Transformation 1h 15m 45s

- 5 Prepare a box of ice.
- 6 Take an Eppendorf tube that contains pre-made competent cells from the 4 -80 °C fridge.
- 7 Immediately place the Eppendorf tube with competent cells into the ice box for **(2)** 00:05:00
- 8 Add the whole Golden Gate Assembly product (\perp 20 μ L) o into the Eppendorf tube containing the competent cells.
- 9 Tap the bottom of the Eppendorf tube to mix the solution.
- 10 Leave the Eppendorf tube in ice for 00:10:00 .
- 11 Place the Eppendoft tube into a foam floating.
- 12 Place them into the water bath for 00:00:45 at 42 °C for heat shock.
- 13 Place the Eppendorf tube into the ice immediately
- 14 Add <u>Add</u> 1 mL of the LB media into the Eppendorf tube.

5m

10m

45s



- Place the Eppendoft tube into the incubator at \$\\$37 \circ\$ for \(\frac{\circ}{\circ} \) 01:00:00 for recovery.
- 1h

16 Centrifuge the Eppendorf tube to form a cell pellet (no specific speed and time).

Plating and Incubation

1h

- 17 Prepare an LB agar plate with the correct antibiotics.
- 18 Remove $\underline{\underline{A}}$ 950 $\mu \underline{L}$ of the LB solution from the Eppendorf tube that contains the cell pellet, leaving about $\underline{\underline{A}}$ 100 $\mu \underline{L}$ in the Eppendorf tube.
- 19 Resuspend the cells by pipetting the solution.
- 20 Spread the cells onto the agar with the L-spreader.
- Place the petri dish in the incubator at 37 °C for Overnight to allow the colonies to grow.

1h