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ÕGI Bio Oxygen Flask Cleaning V.2

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Protocol status: Working

We use this protocol and it's working

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Disclaimer

The cleaning protocols provided by OGI Bio are intended as general guidelines for the maintenance of our micro bioreactor equipment. These protocols are designed to ensure optimal performance and longevity of the equipment under standard operating conditions.

OGI Bio assumes no liability for any damage, contamination, or malfunctions resulting from deviations from these protocols, improper use, or the use of incompatible cleaning agents or methods. It is the responsibility of the user to verify that all cleaning agents and procedures are compatible with local regulations, organizational standards, and the materials of the equipment in use.

Abstract

This protocol is for the cleaning of your OGI BioReactor Oxygen Measurement Flasks.

Guidelines

If the oxygen measurement flasks are autoclaved, they should be left for 7 days before being used again and have their 100% point recalibrated prior to next use, as per our manual. They should be autoclaved at temperatures no higher than 121°C for no more than 15 minutes.

Ethanol should not be left in the flasks for more than 5 minutes.

The oxygen measurement flasks should be stored away from direct light.

Materials

70% Ethanol

Ultrapure (18 MOhm) Water

Equipment

Oxygen Measurement Flask

NAME

Accessory

TYPE

OGI Bio

BRAND

OGI-A-1008

SKU

<https://ogibio.co.uk/product/replacement-oxygen-measurement-flask/>^{LINK}

Safety warnings

- ⚠ Autoclaving may damage the oxygen spots by inducing delamination of their layers. If this protocol is not suitable and you must autoclave the flasks, check the spots for visible signs of damage after autoclaving. Due to the photosensitive nature of the spots, these flasks **may not** be sterilized via exposure to UV light.

Before start

Ensure protocol is carried out in sterile conditions.

Ensure you have familiarised yourself with the guidelines and warnings for this protocol.



Cleaning Oxygen Measurement Flask

- 1 Retaining the magnetic stir bar, empty the contents of each flask into the waste flask.
- 2 Remove stir bars and wipe thoroughly with 70% ethanol. Leave aside. 
- 3 Rinse out any remaining cell culture with 10-20 mL ultrapure water. A vortex mixer can be used if there are stubborn residues. **N.B. ensure that the stir bar is not in the flask if vortexing, this will destroy the spot.**
- 4 Repeat Step 3 if there are still residues, dislodging with a gentle jet of water from a pipette if necessary.
- 5 Vigorously swirl 10-20 mL of 70% ethanol around the flask. A vortex mixer can be used if there are stubborn residues on the flask. Take care to avoid leaving the ethanol in the flask for longer than 5 minutes.
- 6 Pour the ethanol from the flasks and leave to dry inverted, with the stir bars under the flasks. If using caps, spray them with ethanol and leave to dry inverted.

Storage

- 7 Store the flasks with their stir bars inside them with the caps tightly screwed on in a dark place. They should be left for at least 24 hours before being used again.