



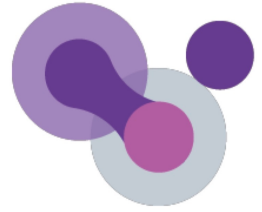
Oct 27, 2020

Version 1

GHRU (Genomic Surveillance of Antimicrobial Resistance) RetrospeBioinformatics Methods V.1

DOI

<https://dx.doi.org/10.17504/protocols.io.bp2l6b11kgqe/v5>



Anthony Underwood¹

¹Centre for Genomic Pathogen Surveillance



Anthony Underwood

Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

OPEN  ACCESS



DOI: <https://dx.doi.org/10.17504/protocols.io.bp2l6b11kgqe/v5>

Protocol Citation: Anthony Underwood 2020. GHRU (Genomic Surveillance of Antimicrobial Resistance) RetrospeBioinformatics Methods. [protocols.io https://dx.doi.org/10.17504/protocols.io.bp2l6b11kgqe/v5](https://dx.doi.org/10.17504/protocols.io.bp2l6b11kgqe/v5)

License: This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: In development

We are still developing and optimizing this protocol

Created: October 27, 2020

Last Modified: October 27, 2020

Protocol Integer ID: 43872

Keywords: genomic surveillance of antimicrobial resistance, analysing genome data, genome sequence data, retrospebioinformatics method, genomic surveillance, genome data, genome, based snp phylogeny, antimicrobial resistance, foundational analyses of de novo assembly, snp phylogeny, de novo assembly, nihr global health research unit, de novo, examination of the pipeline code, description of the pipeline, pipeline code, associated docker

Abstract

A description of the pipelines used for analysing genome data from the retrospective 1 project of the NIHR Global Health Research Unit (Genomic Surveillance of Antimicrobial Resistance)

All genome sequence data were processed using versioned Nextflow [1] workflows and associated Docker [2] containers covering the foundational analyses of de novo assembly, mapping-based SNP phylogeny, MLST assignment and AMR determinant detection (table 1). The exact steps performed can be derived from examination of the pipeline code but each workflow will be described in brief.

1 **De novo assembly**

reads trimming and adapter removal using trimmomatic (0.38) [3], read correction using lighter (1.1.1) [4], downsampling to 100x coverage using seqtk (1.3) [5], read merging using flash (1.2.11) [6], assembly using SPAdes (3.12.0) [7]. Quality control was performed using fastqc (0.11.8) [8], multiqc (1.7) [9] and qualifyr (1.4.4) [10]. Species identification was carried out by bactinspector (0.1.3) [11] and contamination checked using Confindr (0.7.2) [12].

2 **Mapping based phylogeny**

Reads were trimmed as described for de novo assembly and mapped to a reference with bwa mem (0.7.17) [13], variants called and filtered using bcf tools (1.9) [14] and the filtering conditions for a low quality position being '%QUAL<25 || FORMAT/DP<10 || MAX(FORMAT/ADF)<2 || MAX(FORMAT/ADR)<2 || MAX(FORMAT/AD)/SUM(FORMAT/DP)<0.9 || MQ<30 || MQOF>0.1'. A pseudoalignment where each sample has a base relative to the reference sequence. Missing bases and low quality bases are encoded using the - and N characters. The alignment was used to generate a maximum likelihood tree using iqtree (1.6.8) [15,16] and ultrafast bootstraps (parameters -m GTR+G -alrt 1000 -bb 1000).

3 **AMR determinant detection**

The ARIBA software (2.14.4) [17] was used to detect acquired genes using the NCBI database [18] (downloaded 2019-10-30) and the pointfinder database (downloaded 2020-12-11) adapted for Ariba [19].

4 **MLST detection**

The ARIBA software (2.14.4) [17] was used to determine the 7-locus MLST type using the profile and alleles found in the pubmlst database [20,21] (downloaded 2020-12-20).

5 **Table 1: Nextflow workflows**

	Workflow name	Workflow link	Docker hub Container(s) used	Version at publication
	De novo assembly	https://gitlab.com/cgps/ghru/pipelines/assembly	bioinformant/ghru-assembly:versionORregistry.gitlab.com/cgps/ghru/pipelines/assembly:version	1.5.5
	Mapping based SNP phylogeny	https://gitlab.com/cgps/ghru/pipelines/snp_phylogeny	bioinformant/ghru-snp-phylogeny:versionORregistry.gitlab.com/cgps/ghru/pipelines/snp_phylogeny:version	1.2.2

AMR determinant detection	https://gitlab.com/cgps/ghru/pipelines/dsl2/pipelines/amr_prediction	bioinformant/ghru-amr-prediction:versionORregistry.gitlab.com/cgps/ghru/pipelines/dsl2/pipelines/amr_prediction	1.0
MLST	https://gitlab.com/cgps/ghru/pipelines/dsl2/pipelines/mlst	bioinformant/ghru-mlst:versionORregistry.gitlab.com/cgps/ghru/pipelines/dsl2/pipelines/mlst:version	1.0

6 References

1. Di Tommaso P, Chatzou M, Floden EW, Barja PP, Palumbo E, Notredame C. Nextflow enables reproducible computational workflows. *Nat Biotechnol* 2017; 35:316–319.
2. Merkel, D. Docker: lightweight linux containers for consistent development and deployment. 2014; 239.
3. Bolger AM, Lohse M, Usadel B. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinforma Oxf Engl* 2014; 30:2114–2120.
4. Song L, Florea L, Langmead B. Lighter: fast and memory-efficient sequencing error correction without counting. *Genome Biol* 2014; 15:509.
5. Li H. lh3/seqtk. 2020. Available at: <https://github.com/lh3/seqtk>. Accessed 26 October 2020.
6. Magoč T, Salzberg SL. FLASH: fast length adjustment of short reads to improve genome assemblies. *Bioinformatics* 2011; 27:2957–2963.
7. Bankevich A, Nurk S, Antipov D, et al. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol J Comput Mol Cell Biol* 2012; 19:455–477.
8. FastQC A Quality Control tool for High Throughput Sequence Data. Available at: <https://www.bioinformatics.babraham.ac.uk/projects/fastqc/>. Accessed 7 October 2020.
9. Ewels P, Magnusson M, Lundin S, Käller M. MultiQC: summarize analysis results for multiple tools and samples in a single report. *Bioinforma Oxf Engl* 2016; 32:3047–3048.
10. Qualifyr. Available at: <https://gitlab.com/cgps/qualifyr>. Accessed 7 October 2020.
11. BactInspector. Available at: <https://gitlab.com/antunderwood/bactinspector>. Accessed 26 October 2020.
12. Low AJ, Koziol AG, Manninger PA, Blais B, Carrillo CD. ConFindr: rapid detection of intraspecies and cross-species contamination in bacterial whole-genome sequence data. *PeerJ* 2019; 7:e6995.
13. Li H. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM. *ArXiv13033997 Q-Bio* 2013; Available at: <http://arxiv.org/abs/1303.3997>. Accessed 26 October 2020.
14. samtools/bcftools. samtools, 2020. Available at: <https://github.com/samtools/bcftools>. Accessed 26 October 2020.



15. Nguyen L-T, Schmidt HA, von Haeseler A, Minh BQ. IQ-TREE: A Fast and Effective Stochastic Algorithm for Estimating Maximum-Likelihood Phylogenies. *Mol Biol Evol* 2015; 32:268–274.
16. Hoang DT, Chernomor O, von Haeseler A, Minh BQ, Vinh LS. UFBoot2: Improving the Ultrafast Bootstrap Approximation. *Mol Biol Evol* 2018; 35:518–522.
17. Hunt M, Mather AE, Sánchez-Busó L, et al. ARIBA: rapid antimicrobial resistance genotyping directly from sequencing reads. *Microb Genomics* 2017; 3:e000131.
18. Bacterial Antimicrobial Resistance Reference Gene. Available at: <https://www.ncbi.nlm.nih.gov/bioproject/PRJNA313047>. Accessed 26 October 2020.
19. ariba_amr_databases. Available at: https://gitlab.com/cgps/ghru/pipelines/data_sources/ariba_amr_databases. Accessed 26 October 2020.
20. Jolley KA, Chan M-S, Maiden MCJ. mlstdbNet - distributed multi-locus sequence typing (MLST) databases. *BMC Bioinformatics* 2004; 5:86.
21. Jolley KA, Maiden MCJ. BIGSdb: Scalable analysis of bacterial genome variation at the population level. *BMC Bioinformatics* 2010; 11:595.