

Jun 14, 2024

Genotyping Wheat Grains to Identify Parental Donation of Alleles

DOI

<https://dx.doi.org/10.17504/protocols.io.6qpvr84xplmk/v1>

Delfi Dorussen¹, Samuel Burrows¹, Giorgia Di Santolo², Philippa Borrill¹

¹John Innes Centre; ²Sant'Anna School of Advanced Studies



Delfi Dorussen

John Innes Centre

Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

OPEN  ACCESS



DOI: <https://dx.doi.org/10.17504/protocols.io.6qpvr84xplmk/v1>

Protocol Citation: Delfi Dorussen, Samuel Burrows, Giorgia Di Santolo, Philippa Borrill 2024. Genotyping Wheat Grains to Identify Parental Donation of Alleles. **protocols.io** <https://dx.doi.org/10.17504/protocols.io.6qpvr84xplmk/v1>

License: This is an open access protocol distributed under the terms of the **Creative Commons Attribution License**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: May 13, 2024

Last Modified: June 14, 2024

Protocol Integer ID: 99658

Keywords: dna extraction from wheat grain, genotyping wheat grain, wheat research, genotypes of progeny, genotype, dna extraction, parental donation of allele, genotypes of interest, genotyping of the endosperm, missing genotype, genotyping, capable of germination, wheat grain, allele, germination, particular combinations of allele, seedling leaf tissue, dna, progeny, seed formation, embryo, kompetitive allele specific pcr, allowing adult plant, adult plant

Funders Acknowledgements:

BBSRC Doctoral Training Partnership Studentship

Grant ID: BB/T008717/1

BBSRC Research Grant

Grant ID: BB/T013524/1

Designing Future Wheat Institute Strategic Programme

Grant ID: BB/P016855/1

Abstract

In wheat research, KASP (Kompetitive Allele Specific PCR) is widely used to determine the genotypes of progeny from large, segregating populations. However, DNA extraction and genotyping is generally performed on seedling leaf tissue, and thus genotypes that affect seed formation/germination cannot be detected. We have developed a method that relies on DNA extraction from wheat grains to capture these missing genotypes. Furthermore, DNA extraction and genotyping of the endosperm, rather than the embryo, means that we can determine, for each individual, whether an allele is inherited maternally or paternally. This can be used to determine whether an allele is deleterious when inherited either maternally or paternally, or whether particular combinations of alleles are deleterious when inherited from the same parent, but not two separate parents. Moreover, the embryo remains intact throughout this protocol and is capable of germination, allowing adult plants to be recovered from genotypes of interest.



Materials

Reagents

Grain sterilisation & dissection

Sterile diH₂O

Sodium hypochlorite solution

70% ethanol

DNA extraction

DNA extraction buffer (0.1 M Tris-HCl, 0.05 M EDTA, 1.25% SDS):

- 100 mL 1.0 M Tris-HCl, pH 7.5
- 100 mL 0.5 M EDTA, pH 8.0
- 125 mL 10% SDS
- 675 mL sterile diH₂O

6 M ammonium acetate (stored at 4°C)

70% ethanol

Isopropanol

Genotyping

KASP Primers (100 µM stocks)

KASP/PACE mix

Materials

Grain sterilisation & dissection

15 mL Falcon tubes

Shallow 96-well plate (e.g. Nunc™ MicroWell™ 96-Well, NuncIon Delta-Treated, Flat-Bottom Microplate)

Deep well 96-well DNA plate (e.g. Thermo Scientific™ Abgene™ 96 Well 1.2mL Polypropylene DeepWell™ Sample Processing & Storage Plate)

Parafilm

Aluminium foil

Razor blade

Sticky plate seal (e.g. Thermo Scientific™ Nunc™ Adhesive Plate Seal AB-0580)

DNA extraction

Deep well 96-well DNA plate (e.g. Thermo Scientific™ Abgene™ 96 well 1.2mL Polypropylene DeepWell™ Sample Processing & Storage Plate)

Tungsten carbide beads

Sticky plate seal (e.g. Thermo Scientific™ Nunc™ Adhesive Plate Seal AB-0580)

Caps (e.g. Qiagen Collection Microtube Caps)

Genotyping

384-well KASP plate (e.g. FrameStar® 384 Well Skirted PCR Plate clear)

Optically clear sticky plate seal (e.g. Xtra-Clear Advanced Polyolefin StarSeal (qPCR))



Equipment

Vortexer

Orbital shaker

Tissue homogeniser (e.g. Geno Grinder)

Plate centrifuge

Thermocycler

Plate reader

Grain Surface Sterilisation

- 1 Make up the sterilisation solution: add 5 mL sodium hypochlorite solution to 100 mL sterile diH₂O.
- 2 Place the grains in a 15 mL Falcon tube and add 5 mL 70% ethanol. Vortex the tube for 10 seconds, leave for 5 minutes, and vortex again for 10 seconds.
- 3 Pour off the ethanol.
- 4 Add 5 mL of the sterilisation solution to the Falcon tube and shake to dislodge the grains from the bottom of the tube. Place the tube on a shaker for 15 minutes.
- 5 Pour off the sterilisation solution.
- 6 Wash the grains four times with sterile diH₂O, using 10 mL of diH₂O for each wash.

Grain Dissection & Germination

- 7 Working in a flow hood, add 50 µL of sterile diH₂O to each well of the shallow 96-well plate.
- 8 Cut each grain laterally in half with a razor blade (**Figure 1**).

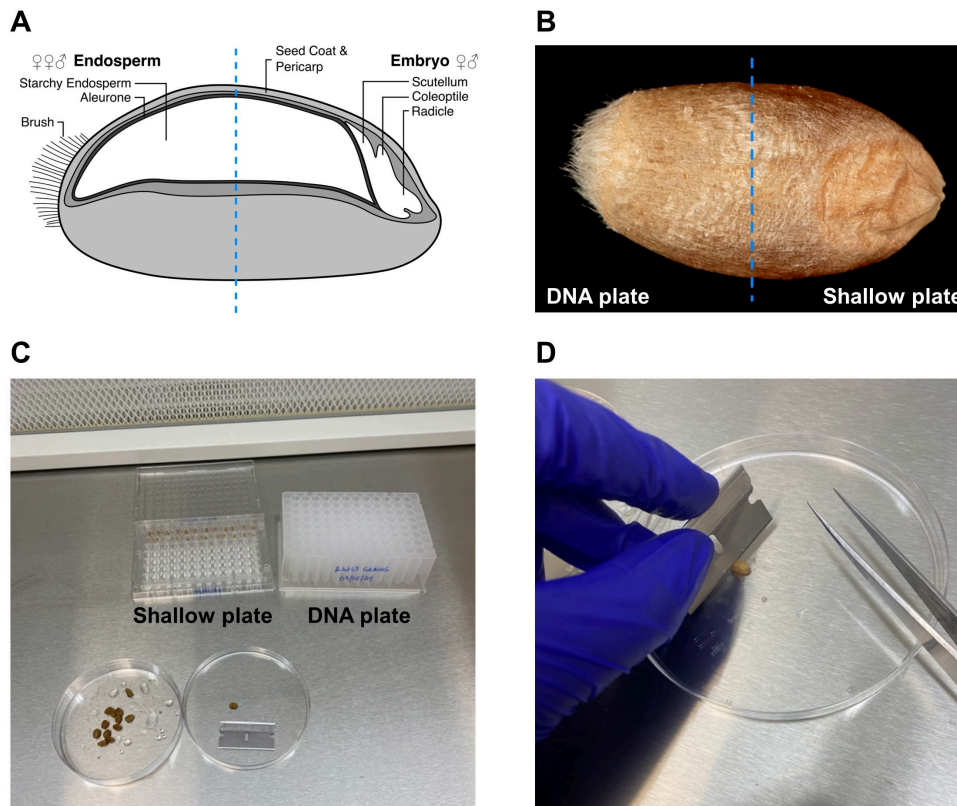


Figure 1: Grain dissection.

A. Cross-section of wheat grain, showing the position of the cut (blue dotted line).
B. Top view of wheat grain, showing the position of the cut (blue dotted line) and in which plate to place each half.
C. Set up when working in the flow hood.
D. Dissection of a wheat grain.

- 9 Place the half of the grain containing the embryo in a well of the shallow 96-well plate. Place the half of the grain containing only the endosperm in a well of the 96-well DNA plate. Ensure that the two grain halves are placed in corresponding wells in the two plates.
- 10 Once all of the grains are dissected, cover the 96-well DNA plate with a sticky seal. If DNA extraction will be completed the same day, the DNA plate can be stored in the fridge at 4°C. Otherwise, store the DNA plate at -20°C.
- 11 Seal the shallow 96-well plate containing the embryo grain halves with parafilm and wrap the plate in aluminium foil. Place the plate in the fridge (4°C) for two days.
- 12 After two days, take the shallow 96-well plate out of the fridge, unwrap the aluminium foil, and leave at room temperature for three to four days to promote germination.

- 13 Sow the embryo grain halves in a 96 cell tray filled with standard seedling compost (**Figure 2**).



Figure 2: Sowing the grains.

Embryo grain half that has germinated and is about to be sown in soil.

DNA Extraction

- 14 If the endosperm grain halves were stored at -20°C (step 10), leave to defrost at room temperature before continuing.
- 15 Preheat the DNA extraction buffer to 65°C .
- 16 Place a tungsten carbide bead in each well of the DNA plate and cap the wells with microtube caps. Shake in a tissue homogeniser (e.g. Geno Grinder) at 1250 rpm for 2 minutes.
- 17 Add 333 μL of DNA extraction buffer to each well of the DNA plate and cap the wells (ensuring the same cap is used for each well as in step 16).
- 18 Shake the DNA plate in a tissue homogeniser at 1750 rpm for 2 minutes.

- 19 Briefly spin the DNA plate in a centrifuge until 3000 rpm is reached.
- 20 Incubate the plate at 65°C for 45 minutes. During the incubation, invert the plate a few times every 15 minutes.
- 21 During the incubation, aliquot 240 μ L of isopropanol into each well of a new 96-well DNA plate. Seal with a sticky plate seal and store at -20°C.
- 22 Once the plate has been incubated at 65°C for 45 minutes, place it in the fridge (4°C) for 15 minutes.
- 23 Briefly spin the DNA plate in a centrifuge until 5000 rpm is reached.
- 24 Add 167 μ L of 6 M ammonium acetate (stored at 4°C) to each well. Seal the plate with caps and shake vigorously for 15 seconds. Store the plate at -20°C for 15 minutes.
- 25 Centrifuge the plate for 8 minutes at 6000 rpm.
- 26 Transfer 400 μ L of the supernatant from each well into the plate containing the isopropanol (prepared in step 21). Seal the plate with caps and shake vigorously for 15 seconds.
- 27 Briefly spin the DNA plate in a centrifuge until 5000 rpm is reached. Remove the caps from the plate and replace with a sticky plate seal.
- 28 Place the DNA plate at -20°C for 15 minutes to precipitate the DNA.
- 29 Centrifuge the plate for 12 minutes at 5200 rpm.
- 30 Remove the plate seal and invert the plate to pour off the supernatant. Place the inverted plate on a paper towel for 10 seconds to remove the remaining liquid.
- 31 Add 350 μ L of 70% ethanol to each well. Centrifuge the plate for 12 minutes at 5200 rpm and pour off the supernatant as before (step 30).



- 32 Leave the plate (uncovered) at room temperature overnight, or at 65°C for 30 minutes.
- 33 Add 200 µL of sterile diH₂O to each well. Seal with a sticky plate seal and vortex the plate.
- 34 Leave the DNA to resuspend at room temperature for 15 minutes or overnight in the fridge (4°C).
- 35 Vortex the plate and then centrifuge for 10 minutes at 5200 rpm.

Genotyping

- 36 Prepare the primer mix by combining the following, and vortex to mix:

A	B
100 µM Vic primer	12 µL
100 µM Fam primer	12 µL
100 µM common primer	30 µL
Water	46 µL
Total	100 µL

Amount of each component of the primer mix.

- 37 Prepare the master mix, scaled for the total number of reactions. For one reaction, combine 2.5 µL PACE/KASP mix with 0.07 µL primer mix. When preparing a large number of reactions, prepare 15% additional master mix. Vortex to mix.
- 38 Add 2.5 µL of master mix into each well of a 384-well plate.
- 39 Add 2.5 µL of DNA to each well and pipette up and down to mix with the master mix. Seal the plate with an optically clear sticky seal.
- 40 Briefly spin the plate in a centrifuge until 5000 rpm is reached.

- 41 Place the 384-well plate in a thermocycler with the following run conditions:

	A	B	C	D
	Denaturation	94°C	15:00 min	
	Denaturation	94°C	00:20 min	x10
	Annealing/ extension	65-57°C	01:00 min	
	Denaturation	94°C	00:20 min	x35
	Annealing/ extension	57°C	01:00 min	
	Hold	12°C		

Run conditions for KASP genotyping.

- 42 Read the plate in a plate reader and visualise the results using a cluster calling software (e.g. KlusterCaller). Four clusters should be visible on the plot, corresponding to the homozygous Vic genotype, heterozygous with maternal donation of the Vic genotype, heterozygous with maternal donation of the Fam genotype, and the homozygous Fam genotype (**Figure 3**).

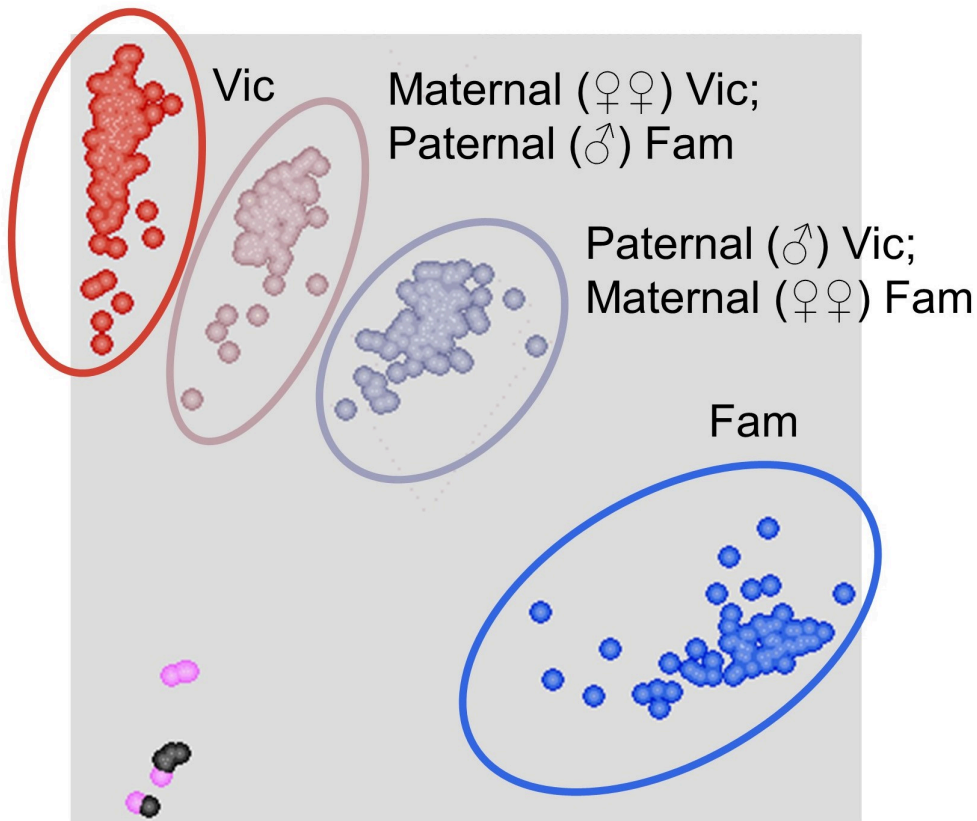


Figure 3: Genotype clusters in KlusterCaller.

Plot showing the four clusters produced from the grain genotyping - homozygous Vic (red), heterozygous with maternal donation of Vic allele (pale pink), heterozygous with maternal donation of Fam allele (lilac), and homozygous Fam (blue).