

Dec 07, 2023

# **©** General Microtome Sectioning of Formalin-Fixed Paraffin Embedded (FFPE) blocks

DOI

dx.doi.org/10.17504/protocols.io.bp2l69morlge/v1



Toby J Curless<sup>1,2</sup>, Zane Jaunmuktane<sup>1,3,2</sup>

<sup>1</sup>Department of Clinical and Movement Neurosciences, UCL Institute of Neurology, Queen Square, London WC1N 3BG, UK;

<sup>2</sup>Aligning Science Across Parkinson's (ASAP) Collaborative Research Network, Chevy Chase, MD, 20 815, USA.; <sup>3</sup>QSBB, 1 Wakefield Street London Greater London WC1N 1PJ, UK



## **Toby J Curless**

Department of Clinical and Movement Neurosciences, UCL Insti...

## Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account





DOI: https://dx.doi.org/10.17504/protocols.io.bp2l69morlqe/v1

Protocol Citation: Toby J Curless, Zane Jaunmuktane 2023. General Microtome Sectioning of Formalin-Fixed Paraffin Embedded (FFPE) blocks. protocols.io https://dx.doi.org/10.17504/protocols.io.bp2169morlqe/v1



License: This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: February 17, 2023

Last Modified: May 31, 2024

Protocol Integer ID: 77172

Keywords: ASAPCRN, Tissue, Sectioning, Parkinson's disease, Microtome, Formalin-fixed Paraffin Embedded, general microtome sectioning of formalin, general microtome sectioning, further tissue processing such as immunohistochemistry, fixed paraffin embedded, further tissue processing, immunohistochemistry, blocks in preparation, fixed paraffin, tissue, embedded block, steps for the sectioning, formalin, ffpe

#### **Funders Acknowledgements:**

The Michael J. Fox Foundation for Parkinson's Research (MJFF) and the Aligning Science Across Parkinson's (ASAP) Initiative Grant ID: ASAP-000478

### Abstract

This protocol describes the steps for the sectioning of formalin-fixed paraffin embedded blocks in preparation for further tissue processing such as immunohistochemistry.

#### **Materials**

Superfrost<sup>TM</sup> charged microscope slides Microtome Fine tweezers Fine paintbrush (optional) Water bath Slide holder

## **Troubleshooting**

## Safety warnings



• Follow approved local risk assessment and institutional policies on the use of sharps.



### Before start

- Check all equipment, supplies and consumables
- Set-up water bath to required temperature
- Ensure blade is fit-for-purpose and clean
- Label/barcode SuperFrost Plus slides



## **Preparation**

- Check all equipment, consumables and supplies are in place.
- 2 Place a clean blade into the blade holder of the microtome.
- 3 Place the FFPE block onto ice, with the face of the block on the surface of the I On ice until the block is cold through.
- 4 Heat  $\perp 1$  distilled  $H_2O$  to  $\mid 40 \circ C \mid$  in a water bath.

## **Sectioning**

- 12h
- 5 Once both block and water are at the correct temperature, insert the block into the chuck of the microtome.
- 6 Adjust the levers on the microtome to position the face of the block parallel with the blade.
- 7 Lower the block until it is level with the blade and move the block closer to the blade using the handle on the microtome.
- 8 Select the thickness of choice on the microtome (usually  $\rightarrow$  4-7 µm.
- 9 Section and trim into the block until the whole tissue cross-section is present in the sections.
- 10 Using tweezers or a fine paintbrush, gently pull sections down until a ribbon of tissue sections has formed.
- 11 Pick up the ribbon with tweezers and place into the water bath to allow wrinkles to disappear from the tissue.





A ribbon of four consecutive sections floating in the water bath to remove wrinkles.

- Separate sections using tweezers and collect individual sections on SuperFrost  $^{\mathsf{TM}}$ 12 charged microscope slides.
- 13 Allow slides to drain and air dry vertically at Room temperature for > 12:00:00 prior to further processing.



